Free leptin index as a marker for leptin action in diabetes and obesity: leptin and soluble leptin receptors relationship with HbA1c

Maciej OWECKI¹, Elżbieta NIKISCH², Anna MICZKE³, Danuta PUPEK-MUSIALIK³, Jerzy SOWIŃSKI¹

- 1 Department of Endocrinology, Metabolism and Internal Medicine, University of Medical Sciences in Poznań, Poland
- 2 Department of Informatics and Statistics, University of Medical Sciences in Poznań, Poland
- 3 Department of Internal Diseases, Metabolic Disturbances and Hypertension, University of Medical Sciences in Poznań, Poland

Assoc. Prof. Maciej Owecki, MD., PhD.
Department of Endocrinology, Metabolism and Internal Medicine
ul. Przybyszewskiego 49, 60-355 Poznań, Poland.
теl: +48 61 8691330; FAX: +48 61 8691682; E-MAIL: mowecki@ump.edu.pl

Submitted: 2010-05-12 Accepted: 2010-08-25 Published online: 2010-12-05

Key words: metabolism; leptin; adipocytokines; diabetes; obesity

Neuroendocrinol Lett 2010; 31(5):679-684 PMID: 21173748 NEL310510A07 © 2010 Neuroendocrinology Letters • www.nel.edu

Abstract **OBJECTIVES:** Glycosylated haemoglobin (HbA1c), leptin, leptin soluble receptors (sOB-R) and free leptin index (FLI) may all be influenced by diabetes, but their associations remain unclear. Therefore, we put forward a hypothesis that serum leptin, sOBR and FLI might be parallel to Hb1c, as they all reflect the metabolic status. **DESIGN AND SETTING:** We measured leptin and sOB-R concentrations in 97 obese non-diabetic (47 women and 50 men), and 65 obese diabetic (32 women and 33 men) humans, and examined whether they were related to HbA1c. Under the condition, the presence of diabetes was the only differentiating factor between two groups of frankly obese humans. **RESULTS:** Non-diabetic vs. diabetic, median and interquartile range, respectively: Leptin (ng/ml), 30.83, 37.27vs.28.24, 23.34; *p*>0.05; sOB-R (ng/ml), 17.62, 17.05vs.21.81, 16.61, p<0.05; FLI, 231.23, 310.00vs.131.76, 157.68, p<0.05. To investigate the influence of HbA1c on leptin and sOB-R, both groups were divided into tertiles based on HbA1c. In diabetics, leptin did not differ between the high, intermediate, and low HbA1c levels subgroups, *p*>0.05, and leptin was not influenced by HbA1c levels: r=0.086; *p*>0.05. For sOB-R, respectively: *p*>0.05; r=0.080; *p*>0.05. In non-diabetics, respectively: *p*<0.05; r=0.2923; *p*<0.05 for leptin; and *p*<0.0001, r=0.5103; *p*<0.0001, for s-OB-R. **CONCLUSIONS:** Not leptin alone but serum sOB-R and FLI are the markers of leptin action impairment in type 2 diabetes. Further, HbA1c is not associated with metabolic status of leptin in obese diabetic patients, whereas this association is found in obese non-diabetic humans.

Abbreviations:

FLI- the free leptin indexinsulin resistanceHbA1c- glycosylated haemoglobin A1csOB-R- soluble leptin receptors	BMI FLI HbA1c	- the body mass index - the free leptin index - glycosylated haemoglobin A1c	HOMA sOB-R	 the homeostasis model assessment of insulin resistance soluble leptin receptors 	
--	---------------------	--	---------------	--	--

INTRODUCTION

Facing an epidemic of type 2 diabetes, clinicians seek valuable markers that could serve as a tool in the management of this disorder. Glycosylated haemoglobin (HbA1c), leptin and leptin soluble receptors (sOB-R) may all be influenced by diabetes and insulin treatment (Aas *et al.* 2009), and thus their measurements may help manage diabetic patients. Nonetheless, whereas HbA1c is the cornerstone of a modern approach to diabetes, the role of leptin and sOB-R in diabetes arouses controversy (Al-Shoumer KA *et al.* 2008; Kratzsch *et al.* 2004).

Leptin, encoded by the ob gene, is a protein that regulates body weight by suppressing food intake and increasing energy expenditure (Ahima & Flier 2000; Campfield et al. 1995; Considine et al. 1995; Halaas et al. 1995; Myer et al. 2008; Sinha et al. 1996a; Zhang et al. 1994). Serum leptin levels show a positive correlation with body fat, being increased in obesity (Considine et al. 1995; Hamilton et al. 1995; Lonnqvist et al. 1995) and decreased in states of severe malnutrition such as anorexia nervosa (Ferron et al. 1997); furthermore, serum leptin decreases postprandially (Stirban et al. 2008) but is not influenced by exercise (Malkova et al. 2008). Therefore, leptin is an adipocyte-synthesized hormone, the role of which is to inform the brain of the amount of the adipose tissue present in the body (Jeanrenaud & Jeanrenaud 1996). In addition, leptin has also a range of other physiological functions including regulation of puberty, placental function, peripheral insulin sensitivity, and interaction with other hormonal mediators and regulators (Meier 1996).

Leptin binds to its receptor, the leptin receptor that exists as membrane-bound isoforms and a soluble form (sOB-R). The latter, of the reported molecular mass of approximately 140 kDa (Liu *et al.* 1997), has been proposed to be a binding protein, and an important regulator of leptin activity (Zastrow *et al.* 2003). The soluble form of the leptin receptor is generated by cleavage of the membrane-bound forms (Maamra *et al.* 2001). It has been proposed that the balance between leptin and sOB-R, or the free leptin index (FLI), might be a useful tool to assess leptin activity (Kratzsch *et al.* 2002).

Elevated concentrations of circulating leptin have been associated with type 2 diabetes, obesity and insulin resistance (Ruhl *et al.* 2001; Zimmet el al. 1996; Koebnick *et al.* 2008; Widjaja *et al.* 1997), whereas data regarding the correlations of sOB-R and FLI with obesity and diabetes are more limited, and conflicting particularly with regard to the markers of glycaemic control in diabetes (Kiess *et al.* 1998; Kratzsch *et al.* 2004). Therefore, we have put forward a hypothesis that serum leptin, sOBR and FLI might be parallel to Hb1c, as they all reflect the metabolic status. To prove this hypothesis, we aimed to measure leptin and sOB-R concentrations in non-diabetic and diabetic humans with a similar range of trunkal obesity, and to examine whether they were related to the levels of HbA1c. Under the condition, the presence of diabetes was the only differentiating factor between two groups of frankly obese humans

MATERIALS AND METHODS

The study group consisted of 97 obese non-diabetic subjects (47 women and 50 men), and 65 obese and diabetic subjects (32 women and 33 men), recruited by local advertising. Both groups were sex- and age-matched. Their age was 51±6.9, and 52±6.7 years, respectively. Obesity was defined according to the WHO criterion $(BMI, >30 \text{ kg/m}^2)$. In all non-diabetics, diabetes was excluded by means of the standard oral glucose tolerance test: 75 grams of glucose was administered orally and the plasma glucose concentration was measured in 2 hours. In the diabetes group, all subjects presented with type 2 diabetes mellitus. Diabetes was defined according to the WHO criterion (fasting glucose of 126 mg/dL, or 6.99 mmol/L, on two various days). All subjects were treated with oral sulfonylureas. All subjects were otherwise healthy according to history, clinical examination, and routine laboratory findings. In particular, none of the studied subjects had evidence of acute or chronic inflammatory disease, had a history of alcohol overconsumption, was completely sedentary, or involved in athletics. All subjects were examined in the morning (at 08:00) after an overnight fast. For women, the examination was made in the middle of the menstruation cycle according to self report. All women had no history of menstrual disturbances or infertility and showed no clinical features of hyperandrogenism or PCOS. Height and weight were measured, and the percentage of body fat was determined with a bioimpedance technique (Bodystat 1500, by BODYSTAT Isle of Man Ltd., U.K.). Repeated bioimpedance tests were performed at the same time after an overnight fast (08:00 A.M.). Exercise, and alcohol or caffeine consumption was banned 24 hrs prior to the bioimpedance test. Premenstrual women and women during menstruation were excluded, as well as patients using diuretics. A venous blood sample was obtained for the measurement of plasma levels of glucose (expressed as mmol/ litre, mmol/l) at the hospital's routine chemistry laboratory. Serum levels of leptin, sOB-R and insulin were determined at the Department of Clinical Biochemistry of the Poznań University of Medical Sciences using commercial ELISA kit (R&D Systems Inc., MN, USA); microplater reader SunriseTM (Tecan Group Ltd., Männedorf, Switzerland), and were expressed as nanograms per millilitre (ng/ml), and as milliunits per litre (mU/l), for leptin and sOB-R, and for insulin, respectively. Leptin and sOB-R were measured according to the manufacturer's recommended protocol - in duplicate, with a sensitivity of 0.0078 ng/ml and 0.057 ng/ml, respectively. Intra-assay coefficients of variation (CV) were respectively for leptin and sOB-R: 4.87% and 5.20%, whereas inter-assay CVs were 5.97% for leptin and 6.13% for sOB-R. The homeostasis model assessment (HOMA) approach was used to calculate the *in vivo* insulin sensitivity with the formula: fasting plasma glucose (mmol/l) × fasting plasma insulin (mU/l) × 22.5^{-1} (Matthews *et al.* 1985).

The values are given as the median and interquartile range. Mann-Whitney test, Kruskal-Wallis test, Dunn test (for post hoc analysis), and Spearman correlation analysis were used for statistical examinations. Standard computer software (STATISTICA 6.0 by Statsoft, Inc.) was applied to the statistical analyses. *p*-values of less than 0.05 were considered statistically significant.

The study was approved by the ethics committee of the Poznań University of Medical Sciences. All subjects gave informed consent to participate.

RESULTS

The clinical data of the diabetic and non-diabetic subjects are shown in Table 1. Both study groups were of similar age, and their body mass indices and percent body fat did not differ. However, as expected, diabetic subjects showed higher levels of HbA1c and plasma glucose. FLI and sOB-R concentrations differed between diabetic and non-diabetic subjects, p<0.05, whereas leptin did not, p>0.05.

To investigate the influence of HbA1c on serum concentrations of leptin and sOB-R, both study groups were divided into tertiles according to their HbA1c levels. In diabetics, as seen in Table 2, except HbA1c, all parameters did not differ between the tertile subgroups. The plasma concentrations of leptin did not differ significantly between the high (H), intermediate (I), and low (L) HbA1c levels subgroups (in Kruskal-Wallis test: p=0.3613; Dunn test: H vs. I, p=0.4615; H vs. L, p=1.000; I vs. L, p=1.000, respectively). Moreover, the serum



Fig. 1. Correlations between leptin and soluble leptin receptors (Leptin sR), and HbA1c in diabetic subjects. Spearman correlation analysis was used, and *p*-values of less than 0.05 were considered statistically significant.

concentration of leptin was not influenced by HbA1c levels: in Spearman's correlation coefficient analysis, the serum leptin levels were not correlated with HbA1c levels (r=0.086; p=0.5179). Moreover, the plasma concentrations of sOB-R in diabetic patients did not differ significantly between the high (H), intermediate (I), and low (L) HbA1c levels subgroups (in Kruskal-Wallis test: p=0.9171; Dunn test: H vs. I, p=1.000; H vs. L, p=1.000; I vs. L, p=1.000, respectively). In addition,

Tab. 1. Clinical characteristics of obese diabetic and non diabetic subjects examined. Data are given as median and interquartile range. The abbreviations used are as follows: n, number of subjects; BMI, the body mass index; P, plasma; HbA1c, glycosylated haemoglobin; sOB-R, soluble leptin receptors; FLI, free leptin index.

	Non-diabetics	Diabetics	مبرامير
n	97	65	<i>p</i> -value
BMI	35.35,	36.20,	0.996
(kg/m²)	10.30	8.50	
Body fat	44.8,	43.85,	0.663
(%)	13.50	12.80	
P glucose	5.5,	7.30,	<0.001
(mmol/liter)	0.90	2.63	
HbA1c	5.90, 0.90	7.40, 1.30	<0.0001
P insulin	25.72,	20.25,	0.163
(mU/liter)	26.05	22.61	
Leptin	30.83,	28.24,	0.574
(ng/ml)	37.27	23.34	
sOB-R	17.62,	21.81,	<0.05
(ng/ml)	17.05	16.61	
FLI	231.23, 310.00	131.76, 157.68	<0.05



Fig. 2. Correlations between leptin and soluble leptin receptors (Leptin sR), and HbA1c in non diabetic subjects. Spearman correlation analysis was used, and *p*-values of less than 0.05 were considered statistically significant.

Tab. 2. Diabetic subjects divided into tertiles based on their HbA1c levels. Data are given as median and interquartile range. The abbreviations used are as follows: n, number of subjects; BMI, the body mass index; P, plasma; HbA1c, glycosylated haemoglobin; sOB-R, soluble leptin receptors; FLI, free leptin index.

Tab. 3. Non-diabetic subjects divided into tertiles based on their HbA1c levels. Data are given as median and interquartile range. The abbreviations used are as follows: n, number of subjects; BMI, the body mass index; P, plasma; HbA1c, glycosylated haemoglobin; sOB-R, soluble leptin receptors; FLI, free leptin index.

	high	intermediate	low	
n	21	22	22	<i>p</i> -value
BMI	36.80,	35.10,	35.35,	0.854
(kg/m²)	7.60	9.00	10.60	
Body fat	48.80,	41.90, 1	42.45,	0.135
(%)	7.50	4.70	9.80	
P glucose	9.27,	7.51,	6.70,	0.301
(mmol/liter)	0.01	2.62	1.20	
HbA1c	9.00, 1.40	7.40, 0.50	6.50, 0.90	<0.0001
P insulin	22.86,	21.78,	18.31,	0.451
(mU/liter)	42.24	20.10	13.74	
Leptin	34.63,	21.50,	27.79,	0.361
(ng/ml)	15.96	35.28	24.18	
sOB-R	25.82,	20.33,	21.04,	0.917
(ng/ml)	21.47	10.78	14.44	
FLI	159.95, 98.63	113.67, 150.45	131.76, 145.39	0.376

	high	intermediate	low	
n	28	36	33	<i>p</i> -value
BMI	39.75,	33.50,	36.60,	<0.05
(kg/m ²)	7.30	10.20	12.40	
Body fat	50.0,	38.25,	43.30,	<0.01
(%)	7.55	17.85	12.00	
P glucose	6.0,	5.55,	5.56,	<0.05
(mmol/liter)	1.0	1.10	1.10	
HbA1c	6.45, 0.30	5.90, 0,25	5.20, 0.60	<0.0001
P insulin	27.03,	28.54,	25.49,	0.692
(mU/liter)	21.37	31.32	24.65	
Leptin	55.66,	25.11,	27.74,	<0.01
(ng/ml)	38.73	31.47	30.36	
sOB-R	24.83,	15.22,	7.96,	<0.0001
(ng/ml)	14.78	18.96	14.89	
FLI	250.9, 225.8	182.5, 295.4	328.5, 508.0	<0.05

the serum concentration of sOB-R was not influenced by HbA1c levels: in Spearman's correlation coefficient analysis, the serum sOB-R levels did not correlate with HbA1c levels (r=0.080; p=0.5492). FLI did not differ significantly between the high (H), intermediate (I), and low (L) HbA1c levels subgroups (in Kruskal-Wallis test: p=0.3761; Dunn test: H vs. I, p=0.6238; H vs. L, p=1.0000; I vs. L, p=0.7495, respectively), and it did not correlate with HbA1c levels (r=0.008; p=0.9521).

The results in non-diabetic subjects are shown in Table 3. Unlike in diabetics, leptin was influenced by HbA1c levels because it differed between the high (H), intermediate (I), and low (L) HbA1c levels subgroups (in Kruskal-Wallis test: P =0.0047; Dunn test: H vs. I, p=0.0073; H vs. L, p=0.0256; I vs. L, p=1.000, respectively), and it correlated with HbA1c levels (r=0.2923; p=0.0073). Further, sOB-R was influenced by HbA1c levels, as it differed between the high (H), intermediate (I), and low (L) HbA1c levels subgroups (in Kruskal-Wallis test: *p*=0.0000; Dunn test: H vs. I, *p*=0.0242; H vs. L, *p*=0.00002; I vs. L, *p*=0.2408, respectively), and it correlated with HbA1c levels (r=0.5103; p<0.0001). FLI did not differ between the high (H), intermediate (I), and low (L) HbA1c levels subgroups (in Kruskal-Wallis test: P =.0431; Dunn test: H vs. I, p=1.0000; H vs. L, p=0.1422; I vs. L, p=0.0667, respectively). However, FLI it correlated with HbA1c levels (r=-0.239; p=0.0294).

In order to evaluate the influence of sex on leptin and sOB-R levels, and gender relation to HbA1c, we estimated all correlations for diabetic and no diabetic females and males separately, in an identical manner. Results were similar to those in the whole cohort. However, due to relatively low numbers of participants we decided not to present the data as reliable conclusions could not have been drawn on their basis.

DISCUSSION

In this study in a homogenous group of obese human subjects with and without type 2 diabetes mellitus, we demonstrate that serum concentrations of leptin are similar despite the presence of type 2 diabetes, whereas the levels of soluble receptors for leptin are significantly higher in diabetic humans as compared with non diabetic individuals. Consequently, the ratio of leptin to its soluble receptors, or the free leptin index (FLI), is significantly decreased in diabetic individuals. That sOB-R levels are higher in diabetic patients is discordant with previous reports (Sun et al. 2010). Furthermore, it is also contrasting with the fact that sOB-R has been reported lower in obese individuals as compared with lean subjects (Considine et al. 1996; Maffei et al. 1995; Sinha et al. 1996b), because obesity is a well established risk factor for the development of type 2 diabetes mellitus.

As a consequence of the above findings, FLI levels were lower in diabetic patients as compared with nondiabetic subjects. As FLI reflects leptin activity, one may conclude that leptin activity was lower in diabetic patients, in spite of similar leptin concentrations. This finding may be explained not only by pure mathematical division. Rather, we postulate that some regulatory mechanisms are impaired in obesity with concomitant diabetes: with similar levels of leptin, higher sOB-R levels are needed to guarantee sufficient leptin action in diabetic subjects. Leptin action would be impaired in those patients not only by the presence of obesity, but also by the presence of diabetes.

What we add to the current knowledge, is that we advocate that it is not leptin alone, and not s-OBR alone that should be measured in diabetes. As sOB-R has been reported to function as inhibitor or stabilizer of leptin (Lammert *et al.* 2001; Liu *et al.* 1997; Sinha *et al.* 1996b), its changes might reflect the proposed leptin resistance in obesity and diabetes (Arch *et al.* 1998; Martin *et al.* 2000; Zimmet *et al.* 1999). In this setting, FLI seems to be the best indicator of metabolic abnormalities that develop in diabetes. The relative imbalance between leptin and its receptors may be one of the foundations of type 2 diabetes pathogenesis.

Further, we analyzed whether leptin, sOB-R and FLI were associated with HbA1c, which is a well established marker of diabetes control. Interestingly, we found no correlations between leptin, sOB-R, FLI and HbA1c in the diabetic group but we observed those associations in non diabetics. This was surprising as one would rather expect parallel increases of both leptin resistance and HbA1c in diabetes. This finding seems to be odd at first sight, however it may be explained with the following theory: in type 2 diabetes, many regulatory mechanisms fail to function correctly. Under the circumstance, the precise interactions between adipocytokines and various serum proteins are disturbed, and presumably so is the interaction between sOB-R and leptin, and HbA1c. The latter, with physiological concentrations in non diabetics, seems to be parallel to leptin and sOB-R, whereas non-physiologically secreted in diabetic humans may not be parallel to changes of adipocytokines concentrations. In addition, there also may be some statistical reasons for this finding in our study: in the non-diabetic group, but not in the diabetic group, BMI and the amount of fat tissue differed between various HbA1c subgroups. As a result, BMI differences might influence leptin, sOB-R and FLI in non obese patients, which in turn influenced the overall statistical results in this group. HbA1c might be associated with BMI, and along with this association, with leptin, sOB-R and FLI.

In conclusion, we demonstrate that serum sOB-R levels are higher, FLI lower, whereas leptin concentrations are similar in diabetic patients as compared with non diabetic subjects with the same degree of obesity. Under the circumstance, FLI seems to be a better marker of leptin action impairment in type 2 diabetes mellitus that leptin alone. Moreover, leptin, sOB-R and FLI did not correlate with HbA1c in diabetics, but they all did in non diabetics. Our findings suggest that HbA1c levels are not associated with the metabolic status of leptin in diabetic patients. Instead, they are correlated with Hb1c in obese non diabetic patients. The loss of this association that occurs in obesity after the development of diabetes remains unclear, but presumably it may be related to the presence of a strong relationship between HbA1c and BMI in non diabetic obese humans and the absence of this associations in diabetes that we also show in this study.

ACKNOWLEDGEMENTS

This work was supported with Grant No. 501-01-02221355-08350 from the University of Medical Sciences in Poznań, Poland.

REFERENCES

- 1 Aas AM, Hanssen KF, Berg JP, Thorsby PM, Birkeland KI (2009). Insulin-stimulated increase in serum leptin levels precedes and correlates with weight gain during insulin therapy in type 2 diabetes. J Clin Endocrinol Metab. **94**: 2900–2906.
- 2 Ahima RS, Flier JS (2000). Leptin. Annu Rev Physiol. 62: 413–437.
- 3 Al-Shoumer KA, Al-Asousi AA, Doi SA, Vasanthy BA (2008). Serum leptin and its relationship with metabolic variables in Arabs with type 2 diabetes mellitus. Ann Saudi Med. **28**: 367–370.
- 4 Arch JR, Stock MJ, Trayhurn P (1998). Leptin resistance in obese humans: does it exist and what does it mean? Int J Obes Relat Metab Disord. 22: 1159–1163.
- 5 Campfield LA, Smith FJ, Guisez Y, Devos R, Burn P (1995). Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. Science. **269**: 546–549.
- 6 Considine RV, Considine EL, Williams CJ, Nyce MR, Magosin SA, Bauer TL, et al (1995). Evidence against either a premature stop codon or the absence of obese gene mRNA in human obesity. J Clin Invest. **95**: 2986–2988.
- 7 Considine RV, Sinha MK, Heiman ML, Kriauciunas A, Stephens TW, Nyce MR, et al (1996). Serum immunoreactive-leptin concentrations in normal-weight and obese humans. N Engl J Med. **334**:292–295.
- 8 Ferron F, Considine RV, Peino R, Lado IG, Dieguez C, Casanueva FF (1997). Serum leptin concentrations in patients with anorexia nervosa, bulimia nervosa, and unspecific eating disorders are independent of the respective disease but correlate with the body mass index. Clin Endocrinol (Oxf). **46**: 289–293.
- 9 Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, et al 1995. Weight-reducing effects of the plasma proteins encoded by the obese gene. Science. **269**: 543–546.
- 10 Hamilton BS, Paglia D, Kwan AYM, Deitel M (1995). Increased obese mRNA expression in omental fat cells from massively obese humans. Nat Med. 1: 953–956.
- 11 Jeanrenaud FR and Jeanrenaud B (1996). Obesity, leptin and the brain. N Engl J Med. **334**: 324–325.
- 12 Kiess W, Anil M, Blum WF, Englaro P, Juul A, Attanasio A, et al (1998). Serum leptin levels in children and adolescents with insulin-dependent diabetes mellitus in relation to metabolic control and body mass index. Eur J Endocrinol. **138**: 501–509.
- 13 Koebnick C, Roberts CK, Shaibi GQ, Kelly LA, Lane CJ, Toledo-Corral CM, et al (2008). Adiponectin and leptin are independently associated with insulin sensitivity, but not with insulin secretion or beta-cell function in overweight Hispanic adolescents. Horm Metab Res. **40**: 708–712.
- 14 Kratzsch J, Deimel A, Galler A, Kapellen T, Klinghammer A, Kiess W (2004). Increased serum soluble leptin receptor levels in children and adolescents with type 1 diabetes mellitus. Eur J Endocrinol. **151**: 475–481.
- 15 Kratzsch J, Lammert A, Bottner A, Seidel B, Mueller G, Thiery J, et al (2002). Circulating soluble leptin receptor and free leptin index during childhood, puberty, and adolescence. J Clin Endocrinol Metab. **87**: 4587–4594.

- 16 Lammert A, Kiess W, Bottner A, Glasow A, Kratzsch J (2001). Soluble leptin receptor represents the main leptin binding activity in human blood. Biochem Biophys Res Commun. 283: 982–988.
- 17 Liu C, Liu XJ, Barry G, Ling N, Maki RA, De Souza EB (1997). Expression and characterization of a putative high affinity human soluble leptin receptor. Endocrinology. **138**: 3548–3554.
- 18 Lonnqvist F, Arner P, Nordfors L, Schalling M (1995). Overexpression of the obese (ob) gene in adipose tissue of human obese subjects. Nat Med. 1: 950–953.
- 19 Maamra M, Bidlingmaier M, Postel-Vinay MC, Wu Z, Strasburger CJ, Ross RJ (2001). Generation of human soluble leptin receptor by proteolytic cleavage of membrane-anchored receptors. Endocrinology. **142**: 4389–4393.
- 20 Maffei M, Halaas J, Ravussin E, Pratley RE, Lee GH, Zhang Y, et al (1995). Leptin levels in human and rodent: measurement of plasma leptin and ob RNA in obese and weight-reduced subjects. Nat Med. 1: 1155–1161.
- 21 Malkova D, McLaughlin R, Manthou E, Wallace AM, Nimmo MA (2008). Effect of moderate-intensity exercise session on preprandial and postprandial responses of circulating ghrelin and appetite. Horm Metab Res. **40**: 410–415.
- 22 Margetic S, Gazzola C, Pegg GG, Hill RA (2002). Leptin: a review of its peripheral actions and interactions. Int J Obes Relat Metab Disord. 26: 1407–1433.
- 23 Martin RL, Perez E, He YJ, Dawson Jr R, Millard WJ (2000). Leptin resistance is associated with hypothalamic leptin receptor mRNA and protein downregulation. Metabolism. **49**: 1479–1484.
- 24 Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC (1985). Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia. **28**: 412–419.
- 25 Meier CA. Leptin secretion and action: an update (1996). Eur J Endocrinol. **134**: 543–544.
- 26 Myers MG, Cowley MA, Münzberg H (2008). Mechanisms of leptin action and leptin resistance. Annu Rev Physiol. **70**: 537–556.

- 27 Ruhl CE, Everhart JE (2001). Leptin concentrations in the United States: relations with demographic and anthropometric measures. Am J Clin Nutr. **74**: 295–301.
- 28 Sinha MK, Ohannesian JP, Heiman ML, Kriauciunas A, Stephens TW, Magosin S, et al (1996a). Nocturnal rise of leptin in lean, obese and non-insulin-dependent diabetes mellitus subjects. J Clin Invest. 97: 1344–1347.
- 29 Sinha MK, Opentanova I, Ohannesian JP, Kolaczynski JW, Heiman ML, Hale J, et al (1996b). Evidence of free and bound leptin in human circulation. Studies in lean and obese subjects and during short-term fasting. J Clin Invest. **98**: 1277–1282.
- 30 Stirban A, Negrean M, Götting C, Stratmann B, Gawlowski T, Mueller-Roesel M, et al (2008). Leptin decreases postprandially in people with type 2 diabetes, an effect reduced by the cooking method. Horm Metab Res. **40**: 896–900.
- 31 Sun Q, van Dam RM, Meigs JB, Franco OH, Mantzoros CS, Hu FB (2010). Leptin and soluble leptin receptor levels in plasma and risk of type 2 diabetes in U.S. women: a prospective study. Diabetes. **59**: 611–618.
- 32 Widjaja A, Stratton IM, Horn R, Holman RR, Turner R, Brabant G (1997). UKPDS 20: plasma leptin, obesity, and plasma insulin in type 2 diabetic subjects. J Clin Endocrinol Metab. **82**: 654–657.
- 33 Zastrow O, Seidel B, Kiess W, Thiery J, Keller E, Bottner A, et al (2003). The soluble leptin receptor is crucial for leptin action: evidence from clinical and experimental data. Int J Obes Relat Metab Disord. 27: 1472–1478.
- 34 Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM (1994). Positional cloning of the mouse obese gene and its human homologue. Nature. **372**: 425–432 (Erratum in Nature 1995; 374: 479).
- 35 Zimmet P, Boyko EJ, Collier GR, de Courten M (1999). Etiology of the metabolic syndrome: potential role of insulin resistance, leptin resistance, and other players. Ann NY Acad Sci. **892**: 25–44.
- 36 Zimmet P, Hodge A, Nicolson M, Staten M, de Courten M, Moore J, et al (1996). Serum leptin concentration, obesity, and insulin resistance in Western Samoans: cross sectional study. BMJ. **313**: 965–969.