

Effects of certain antioxidants on lipid peroxidation process in lung homogenates of L-thyroxine-receiving rats

Joanna WIKTORSKA¹, Andrzej LEWIŃSKI², Michal STUSS¹,
Dariusz NOWAK³, Tadeusz PIETRAS⁴, Ewa SEWERYNEK¹

¹ Department of Endocrine Disorders and Bone Metabolism, Medical University of Lodz, Poland

² Department of Endocrinology and Metabolic Diseases, Medical University of Lodz, Poland

³ Department of Physiology, Medical University of Lodz, Poland

⁴ Clinic of Pneumology and Allergology, Medical University of Lodz, Poland

Correspondence to: Prof. Ewa Sewerynek, MD., PhD.
Department of Endocrine Disorders and Bone Metabolism
Medical University of Lodz
Żeligowskiego Str 7/9, 90-752 Lodz, Poland.
TEL/FAX: +48 (42) 6393127;
E-MAIL: ewa.sewerynek@wp.pl; ewa.sewerynek@umed.lodz.pl

Submitted: 2009-07-01 *Accepted:* 2009-12-15 *Published online:* 2010-02-16

Key words: **lipid peroxidation; hyperthyroidism; hypothyreosis; antioxidants; lungs**

Neuroendocrinol Lett 2010; **31**(1):137-146 PMID: 20150865 NEL310110A03 © 2010 Neuroendocrinology Letters • www.nel.edu

Abstract

OBJECTIVE: A possible role of antioxidants in thyreotoxicosis was investigated. We examined the parameters of lipid peroxidation (LPO): conjugated dienes (CD), malondialdehyde (MDA), Schiff bases (SB) in lung homogenates of male Wistar rats.

METHODS: Two control groups were created: Group 1 – intact animals and Group 2 – animals injected with 0,9% NaCl. In Experiment I, the animals received L-thyroxin (LT4) i.p. (Groups 3–7). After one week the rats received additionally: Group 4 – melatonin (MEL); Group 5 – propylthiouracil (PTU); Group 6 – Ambroxol (AMB); Group 7 – N-acetylcysteine (NAC). In Experiment II, the animals received only antioxidants.

RESULTS: In Experiment I, we noticed a significantly higher MDA and SB level in Group 2, compared to that in Group 1. Moreover, we observed a significantly higher MDA and SB level in Group 3, vs. that in Group 1, but SB level was lower in Group 3 than in Group 2. Melatonin, PTU and NAC reduced CD; PTU, AMB diminished MDA and MEL, AMB lowered SB levels as compared to Group 3. In Experiment II, we observed significantly higher MDA and SB level in Group 2, vs. that in Group 1. Melatonin, AMB and NAC decreased MDA and SB level, when compared to Group 2 but PTU elevated MDA and SB level vs. that in Group 1.

CONCLUSIONS: 1) L-T4 suppresses LPO, 2) MEL, AMB and NAC protect against LPO, 3) PTU is an antioxidant in thyreotoxicosis, however, when administered alone, it enhances LPO, 4) stress accelerate LPO.

INTRODUCTION

Free radicals (FR) occur as by-products of mitochondrial metabolism in the mechanism of electron leak from the respiratory chain. Lipid peroxidation (LPO) is the best-known chain free radical process. The following products result, among others, from subsequent stages of this process: conjugated dienes (CD), aldehydes, such as: malondialdehyde (MDA) and Schiff bases (SB) (Asayama *et al.* 1987).

An excess of thyroid hormones accelerates the basic metabolism, especially aerobic metabolism in mitochondria (Fernandez *et al.* 2006; Harper and Seifert 2008). Triiodothyronine (T_3) – induced enhanced oxygen use leads to excessive production of aerobic free radicals and – subsequently – to an increased use of intracellular antioxidants, as well as to inactivation of antioxidative enzymes, inducing, in this way, oxidative stress in tissues, both in thyroid hormone receiving animals and in people with hyperthyroidism (Fernandez *et al.* 2006). Excessive LPO in hyperthyreosis results from enhanced cellular respiration; euthyreosis restoration normalises the process (Bianchi *et al.* 1999).

The cells are equipped with an antioxidative protection system, which is responsible for systemic protection against detrimental effects of free radicals (Sewerynek and Wiktorska 1997). However, the efficiency of this system fails in cases of an excessive severity of oxidative stress (Bartosz 2003). Studies of many authors concentrate on searching for substances which, when, administered, could suppress the production of free radicals and reduce the degree of FR-induced damage. Examples of such compounds include drugs, such as prophythiouracil (PTU), ambroxol (AMB), N-acetylcysteine (NAC) or certain hormones, such as melatonin (MEL).

Prophythiouracil is well known anti-thyroid drug. It has also got antioxidative properties (Hicks *et al.* 1992; Wiktorska *et al.* 2002). A number of authors claim that a therapy with PTU application reduces LPO severity in patients with hyperthyroidism (Bianchi *et al.* 1999). It is not clear, however, if PTU directly “sweeps” FR and LPO products (Hicks *et al.* 1992) or if this activity results from regained euthyreosis (Bianchi *et al.* 1999).

Melatonin (5-methoxy-N-acetyltryptamine) is a hormone, synthesised in the pineal gland. During recent years, much attention has been paid to its antioxidative and anti-neoplastic potential (Karbownik *et al.* 2001). Many authors highlight the fact that MEL effectively suppresses LPO, induced by excessive levels of thyroid hormones in experimental animals (Sewerynek *et al.* 1999; Wiktorska *et al.* 2005; 2006a), while pinealectomy enhances this process (Mogulkoc *et al.* 2005c).

Ambroxol (trans-4-(2)-amino-3,5-dibromobenzylaminocyclohexane hydrochloride) and NAC are known mucolytic drugs. It has been shown that AMB possesses anti-oxidative properties (Nowak *et al.* 1994, 1995). It should be stressed that this agent reveals

affinity to pulmonary tissue, where its concentrations are much higher than in blood. There are also many reports on anti-oxidative effects of NAC (Nowak *et al.* 1993; 1995). The most significant NAC activity is associated with increasing the resources of intracellular GSH (Gillissen and Nowak 1998). It should be stressed that the lungs, as an organ, directly exposed to contacts with the outer environment, are at the highest risk of being the target for many harmful biological and chemical agents, contained in the air. Smaller basic intensification of LPO (López-Torres *et al.* 1993) and smaller – than in remaining studied rats’ organs – activity of cytochrome oxidase (Pérez-Campo *et al.* 1993) evidence about efficiency of antioxidant defense mechanisms in lung.

The goal of this study was an evaluation of LPO severity in the lung of rats, administered with levothyroxin ($L-T_4$) injections. Either the levels or the concentrations of LPO – CD, MDA and SB products in lung homogenates were measured, as well as the antioxidative effects of PTU, MEL, AMB and NAC were determined in suppressing the process. The study aimed also at an analysis of the antioxidative effects of PTU, MEL, AMB and NAC on the basic level of LPO products.

MATERIAL AND METHODS

The experiments were performed on male Wistar rats with initial body weight of 220 ± 20 g. The animals were kept at a room with constant temperature (21–23°C) and controlled light conditions (12 h light/12 h darkness, the light phase beginning at 6:00 a.m.). The rats had constant access to standard feed (LSM-Agropol, Motycz) and tap water.

All the injections were done intraperitoneally (i.p.) between 4:00–4:15 p.m. The active substances were administered, following their prior solution in 0.9% sodium chloride (NaCl) (the volume of injected solutions – 0.8 ml). Melatonin was, at first, solved in a small ethanol (absolute alcohol) volume and then in 0.9% solution of NaCl (ethanol concentration in the final solution was not above 1%). The solutions of used agents were prepared every day, *ex tempore*.

After experiment time period, planned for each study group, the animals were anaesthetised, weighed and decapitated. Immediately before decapitation, blood was aspirated by heart apex puncture. Then, lungs were collected from all the animals, which were stored in -80°C until homogenised and biochemically assayed. The homogenates were prepared in 1:10 dilution (1 weighed part of defrozen tissue: 10 volumetric parts of homogenate), using redistilled water of temperature close to 0°C . In blood plasma, thyroid hormone concentrations – free thyroxin (FT_4) and free triiodothyronine (FT_3) – were measured by the immunochemoluminescent method. The following LPO products were measured in tissue homogenates: MDA (acc. to Rice and Evans (1991)) and CD (acc. to Buege and Aust

(1978)) – by the spectrophotometric method and SB (acc. to Buege and Aust (1978)) – by the spectrofluorometric method.

Two experiments were performed. Because of different ways of antioxidant administration, two control groups were introduced in both experiments (Group 1 and 2). The animals of Group 1, not receiving any injections (intact animals), were in the control group for the rats, which received PTU in their drinking water. Additionally, Group 1 was regarded as the control for the effects of other substances (NaCl, L-T₄, MEL, AMB and NAC). The rats, receiving injections of 0.9% NaCl solution (Group 2), were regarded to be the control for the animals, receiving L-T₄, MEL, AMB and NAC also by injections. In Experiment I, the animals received L-T₄ in dose of 100 µg/kg b.w. per day, i.p., for 14 days (Group 3–6) and, additionally, antioxidants for a 7-day, decapitation preceding period: PTU in drinking water – 0.1% solution (Group 4), MEL in dose of 5 mg/kg b.w. per day, i.p. (Group 5), AMB in dose of 0.169 mmol/kg b.w. per day, i.p. (Group 6) or NAC in dose of 0.169 mmol/kg b.w. per day, i.p. (Group 7). In Experiment II, the animals received only antioxidants for 7 days before decapitation: PTU in drinking water – 0.1% solution (Group 3), MEL in dose of 5 mg/kg b.w. per day, i.p. (Group 4), AMB in dose of 0.169 mmol/kg b.w. per day, i.p. (Group 5) or NAC on dose of 169 mmol/kg b.w. per day, i.p. (Group 6).

The results, obtained in both experiments, were submitted to statistical analysis by means of the one-way ANOVA test (a one-way analysis of variance). Newman-Keuls's test was applied in order to identify statistically different groups and to determine statistical significance. The calculations were performed by the "STATISTICA" computer program. The results have been presented as figures, on which the levels of statistical significance (*p*) are indicated.

RESULTS

Experiment I

After the experiment, a lower body mass was found in the rats of the group administered with L-T₄ and PTU (Group 4), if compared with either the group, receiving L-T₄ (Group 3) or both control groups (Group 1 and 2) (Figure 1).

In all the studied groups of animals, which were injected L-T₄ (Groups 3–7), an increase of FT₄ concentration was indicated with reference to both control groups (Group 1 and 2). In the rats, submitted to the activity of L-T₄ and PTU (Group 4), L-T₄ and AMB (Group 6), L-T₄ and NAC (Group 7), the concentration of FT₄ was higher in comparison with the animals, which received L-T₄ (Group 3) (Figure 2). Higher FT₃ concentration was observed in the rats, receiving L-T₄ (Group 3) in comparison with Groups 1 and 2. FT₃ concentration was lower in the rats, exposed to the activity of L-T₄ and PTU (Group 4) and L-T₄ and MEL (Group 5), when referred to the rats, injected with L-T₄ (Group 3) and was not statistically different from the concentrations, observed in Groups 1 and 2. FT₃ concentration in the animals, receiving L-T₄ and NAC (Group 7) was higher in comparison with the rats, receiving L-T₄ (Group 3). In the animals, injected with L-T₄ and AMB (Group 6) or L-T₄ and NAC (Group 7), higher concentrations of FT₃ were obtained than in Groups 1 and 2 (Figure 3).

It was also found that PTU (Group 4), MEL (Group 5) and NAC (Group 7) decreased CD level with regards to the rats, receiving L-T₄ (Group 3) (Figure 4).

A difference of MDA and SB concentrations was obtained between the control groups – the parameters reached higher values in the animals, injected with 0.9% NaCl solution (Group 2), when compared with intact animals (Group 1) (Figures 5 and 6).

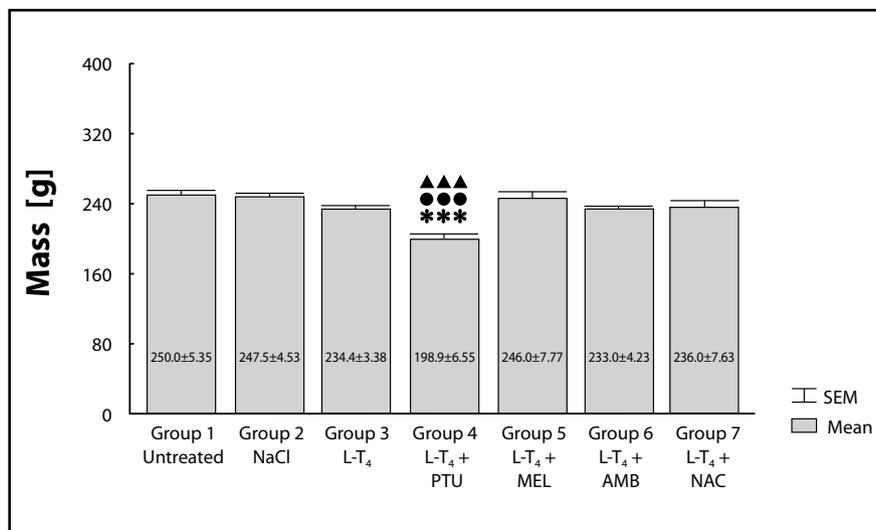


Fig. 1. Mass of rats.

Data represent means ± SEM.

Level of significance:

▲▲▲ - vs. untreated controls, *p* < 0.0005

●●● - vs. saline treated group, *p* < 0.0005

*** - vs. L-T₄ - treated group, *p* < 0.0005

Fig. 2. Concentrations of FT₄ in the serum of rats.
Data represent means ± SEM.
Level of significance:
▲▲▲ - vs. untreated controls, *p*<0.0005
●●● - vs. saline treated group, *p*<0.0005
*** - vs. L-T₄ - treated group, *p*<0.0005
** - vs. L-T₄ - treated group, *p*<0.005
* - vs. L-T₄ - treated group, *p*<0.05

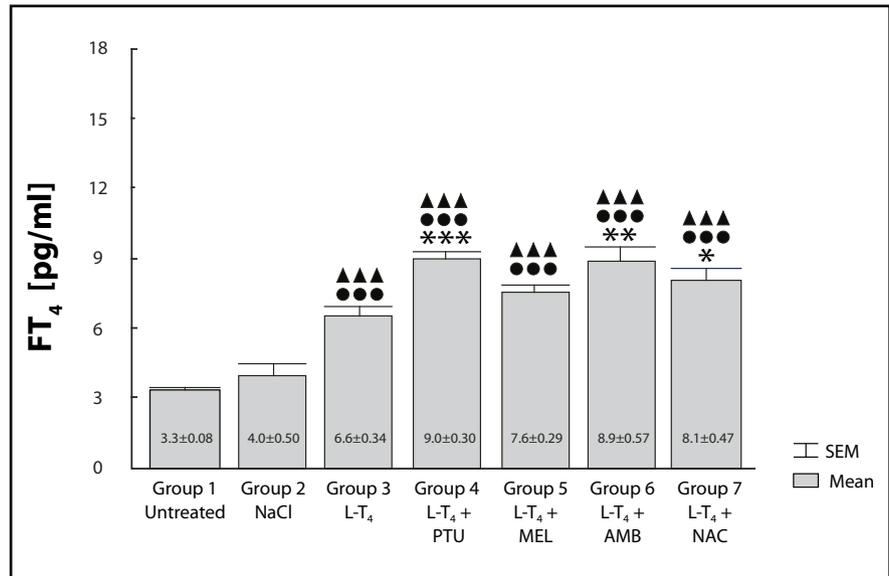


Fig. 3. Concentrations of FT₃ in the serum of rats.
Data represent means ± SEM.
Level of significance:
▲▲▲ - vs. untreated controls, *p*<0.0005
▲▲ - vs. untreated controls, *p*<0.005
●●● - vs. saline treated group, *p*<0.0005
●● - vs. saline treated group, *p*<0.005
*** - vs. L-T₄ - treated group, *p*<0.0005

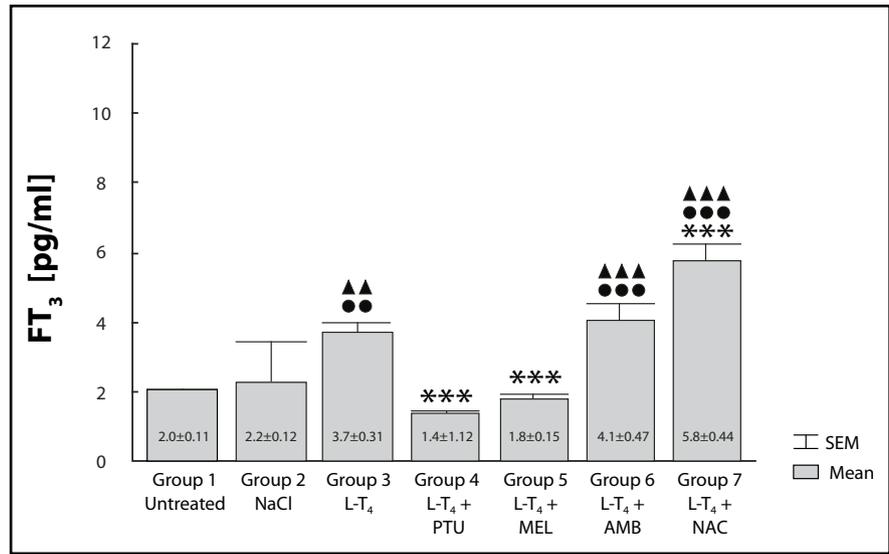
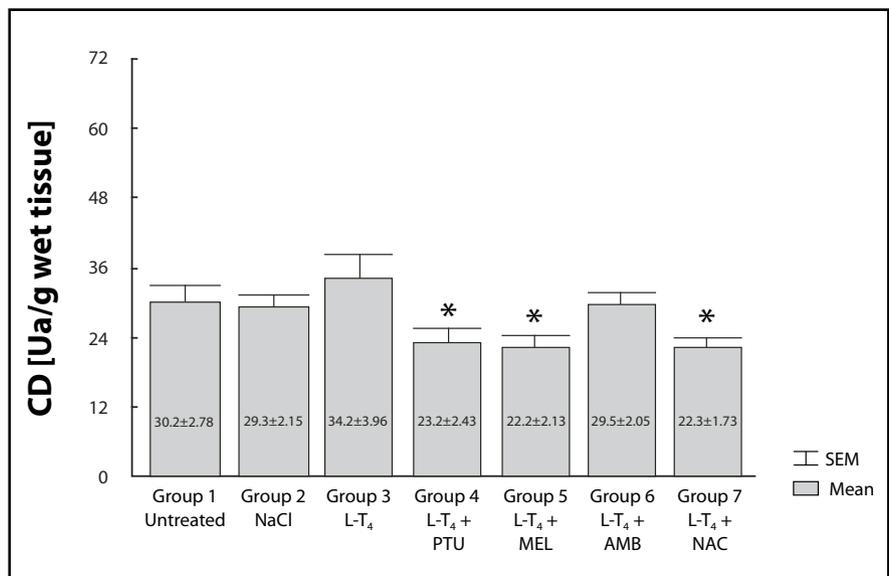


Fig. 4. Levels of CD in the lungs of rats.
Data represent means ± SEM.
Level of significance:
* - vs. L-T₄ - treated group, *p*<0.05



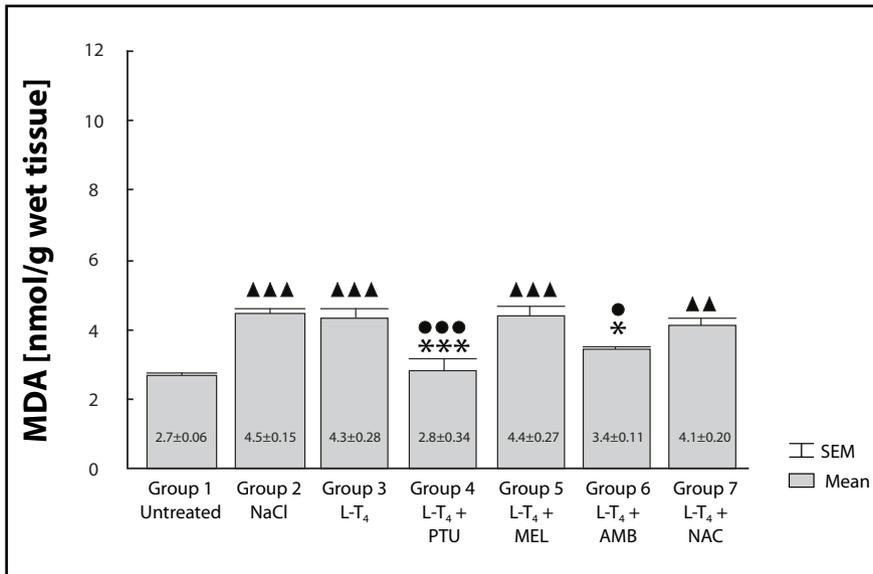


Fig. 5. Concentrations of MDA in the lungs of rats. Data represent means \pm SEM. Level of significance: ▲▲▲ - vs. untreated controls, $p < 0.0005$; ▲▲ - vs. untreated controls, $p < 0.005$; ●●● - vs. saline treated group, $p < 0.0005$; ● - vs. saline treated group, $p < 0.05$; *** - vs. L-T₄ - treated group, $p < 0.0005$; * - vs. L-T₄ - treated group, $p < 0.05$.

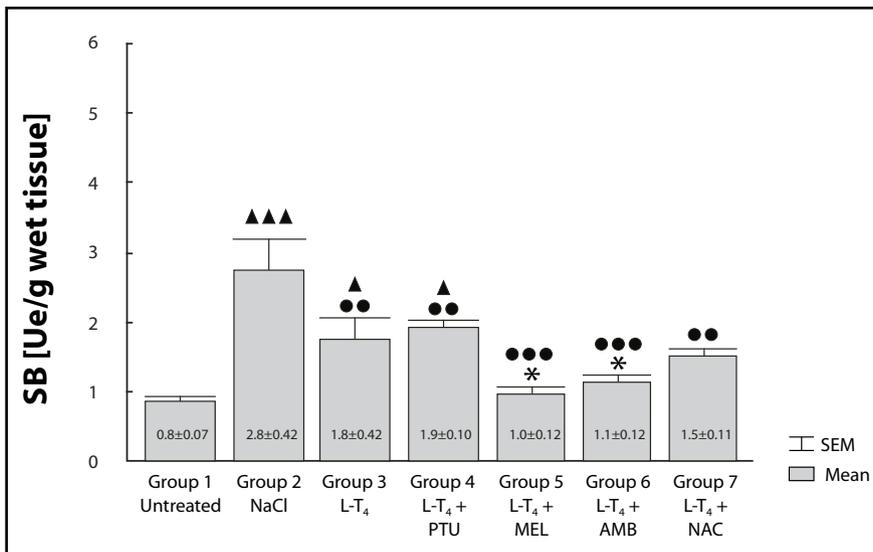


Fig. 6. Levels of SB in the lungs of rats. Data represent means \pm SEM. Level of significance: ▲▲▲ - vs. untreated controls, $p < 0.0005$; ▲▲ - vs. untreated controls, $p < 0.05$; ●●● - vs. saline treated group, $p < 0.0005$; ●● - vs. saline treated group, $p < 0.005$; * - vs. L-T₄ - treated group, $p < 0.05$.

Malondialdehyde concentration was higher in the rats, which had been administered L-T₄ (Group 3) than in Group 1. Prophylthiouracil (Group 4) and AMB (Group 6) decreased MDA concentration in comparison to both the group, receiving L-T₄ (Group 3) and Group 2, down to a value not significantly differing from that, observed in Group 1. In the animals, submitted to the joint activity of L-T₄ and MEL (Group 5) or of L-T₄ and NAC (Group 7), the value of that parameter was higher with regards to Group 1 (Figure 5).

Shiff bases level had the lowest values in the rats, which had been injected L-T₄ (Group 3), when referred to Group 2, while it demonstrated a higher value from that, obtained in Group 1. The administration of MEL (Group 5) and AMB (Group 6) decreased SB level, both with regards the group receiving L-T₄ (Group 3) and Group 2, down to a value not significantly different from that, found in Group 1. Moreover, in the ani-

mals submitted to the joint activity of L-T₄ and PTU (Group 4) and of L-T₄ and NAC (Group 7), the value of that parameter was lower in comparison with Group 2. However, in case of the rats, which had been administered L-T₄ and PTU (Group 4), SB level was higher from that in Group 1 (Figure 6).

Experiment II

After the experiment, a lower body mass was found in the rats of the PTU administered group (Group 3) in comparison to both control groups (Groups 1 and 2) (Figure 7).

Lower concentration of FT₄ was obtained in the PTU receiving animals (Group 3) than in both control groups (Groups 1 and 2). NAC (Group 6) decreased the value of that parameter, with reference to the rats, injected with 0.9% NaCl solution (Group 2) (Figure 8). Also, a decrease in FT₃ concentration was observed in

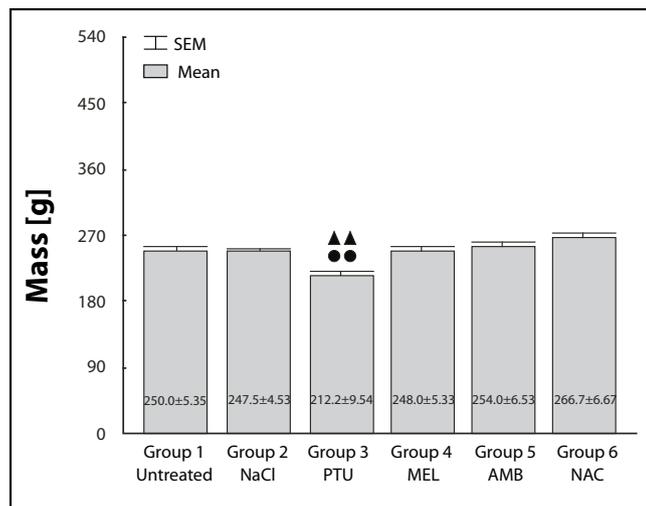


Fig. 7. Mass of rats. Data represent means ± SEM.
Level of significance:
▲▲ - vs. untreated controls, $p < 0.005$
●● - vs. saline treated group, $p < 0.005$

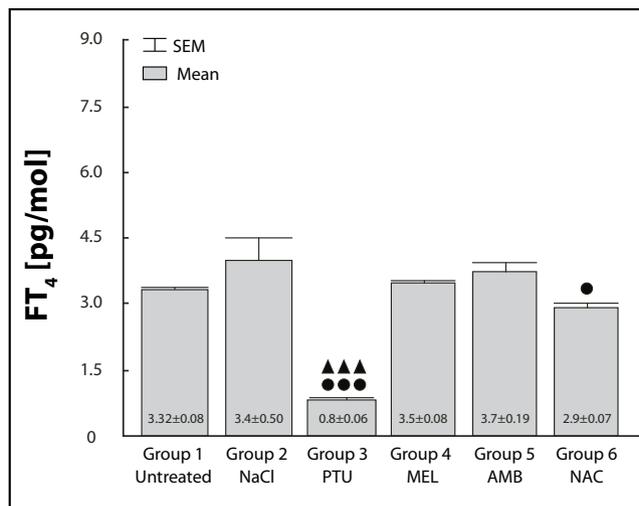


Fig. 8. Concentrations of FT₄ in the serum of rats.
Data represent means ± SEM.
Level of significance:
▲▲▲ - vs. untreated controls, $p < 0.0005$
●●● - vs. saline treated group, $p < 0.0005$
● - vs. saline treated group, $p < 0.05$

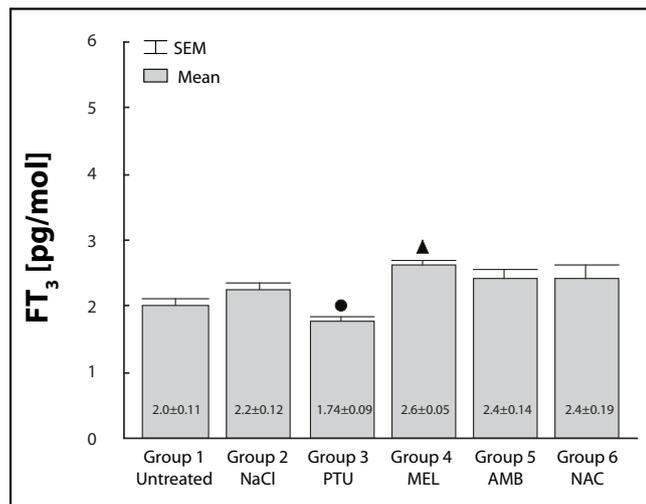


Fig. 9. Concentrations of FT₃ in the serum of rats.
Data represent means ± SEM.
Level of significance:
▲ - vs. untreated controls, $p < 0.05$
● - vs. saline treated group, $p < 0.05$

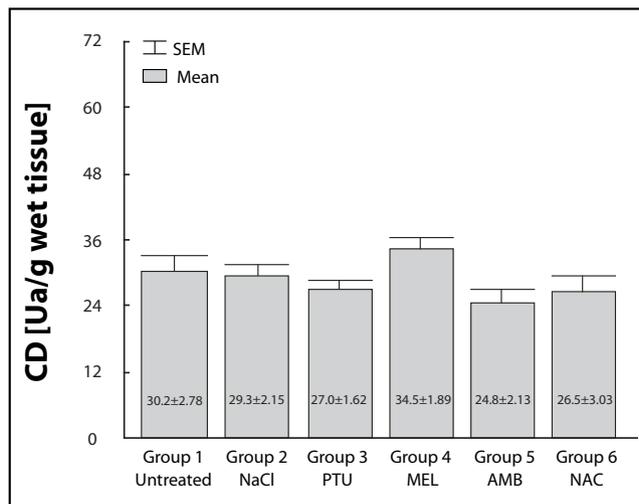


Fig. 10. Levels of CD in the lungs of rats.
Data represent means ± SEM.
Level of significance:

the animals, exposed to PTU activity, when compared with that in Group 2. The studied parameter reached a higher values in the MEL administered rats (Group 4), when compared to intact animals (Group 1) (Figure 9).

No statistically significant differences were noted in CD level (Figure 10).

It was found, however, that MDA concentration and SB level were higher in Group 2 than in Group 1 (Figures 11 and 12).

The administration of MEL (Group 4), AMB (Group 5), NAC (Group 6) decreased MDA concentration in comparison with Group 2, however, that parameter had a higher value from that, observed in Group 1. Prophythiouracil (Group 3) increased MDA concen-

tration in comparison with the respective control group (Group 1); however, that parameter demonstrated a lower value, when compared to the one in Group 2 (Figure 11).

Melatonin (Group 4) and NAC (Group 6) decreased the value of that parameter with regards to Group 2, down to a value not significantly different from that, obtained in Group 1. In turn, AMB (Group 5) decreased SB level, when compared to that in Group 2; that parameter reached a higher value from that in Group 1. It was demonstrated that PTU administration (Group 3) increased SB level in comparison to the respective control group (Group 1), although that parameter had a lower value from that, observed in Group 2 (Figure 12).

DISCUSSION

In the present paper, body mass decrease was demonstrated in the rats, administered either PTU alone or PTU together with L-T₄, when compared with the animals of the other groups. Similar observations were made by other authors (Asayama *et al.* 1987; Kariya *et al.* 1983, Sahoo *et al.* 2008). The suppression of body mass growth in the animals after PTU may be explained by the toxic effects of anti-thyroid drugs (Kariya *et al.* 1983). However, no statistically significant difference was observed in body mass of the rats, receiving L-T₄. It is conformable with reports of other authors (Asayama *et al.* 1987).

It is worth noting that the identification of clinical symptoms of thyreotoxicosis in rats is rather difficult. As it has already been mentioned, neither in this paper nor in reports of other authors, no body mass decrease – a phenomenon, typical for hyperthyroidism in humans – was demonstrated. A proof for the efficacy of L-T₄ administration was the statistically significant increase of FT₄ and FT₃ in blood serum, also observed by other authors (Asayama *et al.* 1987; Baydas and Meral 2005; Mano *et al.* 1995; Sewerynek *et al.* 1999).

In the present study, MEL, administered together with L-T₄, decreased FT₃ concentration in blood of the animals, while, when given separately, it demonstrated an opposite activity – increasing the level of FT₃. There are numerous reports, indicating the suppressive effect of MEL on growth processes and synthesis of the thyroid hormones (Karbownik and Lewinski 2003).

In the performed experiments, similarly as in the earlier publication (Wiktorska *et al.* 2005), a differ-

ence was observed between the two control groups in the content of LPO products. The levels of oxidative stress parameters were significantly higher in the animals, receiving injections of 0.9% NaCl solution, when compared with intact animals. Those results suggest a stimulatory effect of stress on LPO. The stress may probably have been induced by manual handling of the animals and by the accompanying laboratory procedures. Smith-Kielland (Smith-Kielland and Morland 1981) claims that the enhanced synthesis of proteins in the livers of rats, receiving intraperitoneal injections of 1 ml of 0.9% NaCl solution resulted from stress and suggests that catecholamines and some steroid hormones mediate these changes.

Surgical stress is a known factor, enhancing LPO in pulmonary tissue (Jablonka *et al.* 1992). Other authors observed amplified oxidative stress in rats, resulting from opening the abdominal cavity and touching the intestine (Thomas *et al.* 2003). Other stressors, intensifying LPO process, include noise (Manikandan *et al.* 2006), sleep deprivation (Singh *et al.* 2008), long-term isolation (Huong *et al.* 2005) and immobilisation and cold (Bozhko and Gorodetskaia 1991).

In the available literature, numerous reports can be found on oxidative stress intensification in the course of hyperthyroidism in humans (Wiktorska *et al.* 2006a). Many of these reports indicate LPO enhancement in hyperthyreosis and normalisation of these parameters under PTU (Bianchi *et al.* 1999; Erdamar *et al.* 2008; Yavuz *et al.* 2004), or tiamazol therapy (Bozhko and Gorodetskaia 1991; Sewerynek *et al.* 2000). It should be noted that some authors have observed enhanced LPO in the course of both hyper- and hypothyroidism in

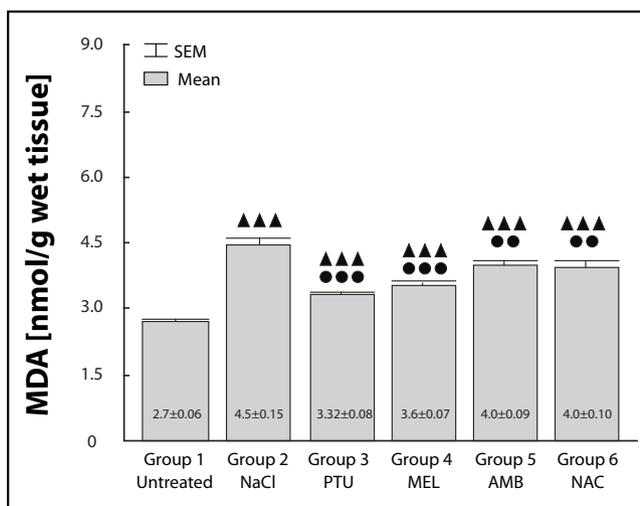


Fig. 11. Concentrations of MDA in the lungs of rats.

Data represent means ± SEM.

Level of significance:

- ▲▲▲ - vs. untreated controls, $p < 0.0005$
- - vs. saline treated group, $p < 0.0005$
- - vs. saline treated group, $p < 0.005$

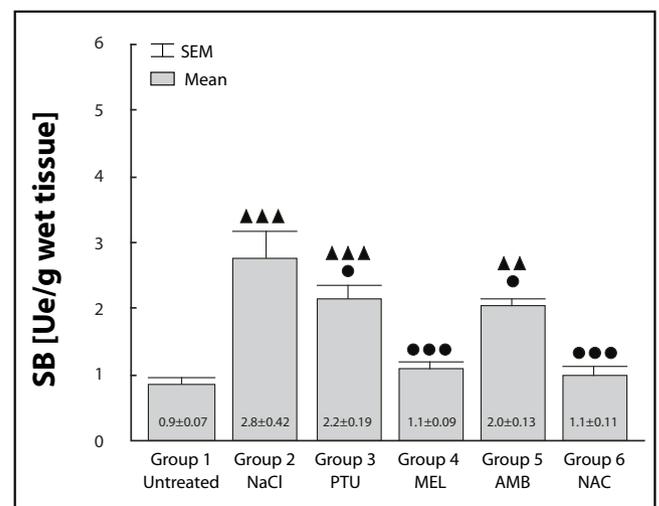


Fig. 12. Levels of SB in the lungs of rats. .

Data represent means ± SEM.

Level of significance:

- ▲▲▲ - vs. untreated controls, $p < 0.0005$
- ▲▲ - vs. untreated controls, $p < 0.005$
- - vs. saline treated group, $p < 0.0005$
- - vs. saline treated group, $p < 0.005$
- - vs. saline treated group, $p < 0.05$

humans (Dumitriu *et al.* 1988; Erdamar *et al.* 2008; Oge *et al.* 2004). There are also numerous papers, reporting that L-T₄ administration to rats enhances LPO in blood (Baydas and Meral 2005), in the brain (Wiktorska *et al.* 2005; Mogulkoc *et al.* 2005c), in the heart (Asayama *et al.* 1987; Mogulkoc *et al.* 2005c), in the liver (Mogulkoc *et al.* 2005c) and in kidneys and testes of rats (Mogulkoc *et al.* 2005a).

Unexpectedly, in this study, no LPO induction was demonstrated in lung homogenates of the rats injected with L-T₄; on the contrary, the level of SB in those animals was lower, when compared with the control group, receiving 0.9% NaCl solution. LPO decrease in thyrotoxicosis is in conformity with reports of other authors. According to Bozhko and Gorodestkaia (1991), an administration of thyroid hormones in small doses inhibits LPO in the cardiac muscle of rats, submitted to immobilisation and cold. There are also reports that L-T₄, administered for 4 weeks, decreased LPO in the liver (Guerrero *et al.* 1999) and in the brain (Mano *et al.* 1995) of rats.

It was observed in the present study that PTU had decreased CD level and MDA concentration in lung homogenates from the L-T₄ administered animals. Similar observations were made in brain homogenates (Wiktorska *et al.* 2005). In another study PTU decreased SB concentrations in liver and lung homogenates after the administration of pharmacological doses of iodide (Sewerynek *et al.* 2006). According to Rodriguez-Pierce *et al.* (1994), PTU, when administered to pregnant rats, reduced the formation of MDA and of pathological changes in the lungs and increased the survival of new-borns, submitted to hyperoxy. There are also studies, confirming the inhibitory effect of this drug on oxidative processes in the course of hyperthyreosis in humans (Bianchi *et al.* 1999; Erdamar *et al.* 2008; Yavuz *et al.* 2004).

However, PTU, administered to animals, which did not receive L-T₄ before, induced hypothyreosis and increased the levels of MDA and SB in lung homogenates. It is known that hypothyreosis causes profound changes in oxidative processes of organisms (Wiktorska *et al.* 2006b). Intensification of oxidative stress was observed in studies on hypothyroidism in humans (Dumitriu *et al.* 1988; Erdamar *et al.* 2008; Nanda *et al.* 2008; Oge *et al.* 2004). The authors assume that the excessive LPO results from slower metabolic changes (Dumitriu *et al.* 1988). Also in the studies, performed on animals, PTU enhanced LPO in internal organs: the brain (Rahaman *et al.* 2001), the liver (Yilmaz *et al.* 2003), the testis (Sahoo *et al.* 2008) and in the heart (Chattopadhyay *et al.* 2003). Supposedly, PTU has also got prooxidative properties per se (Hicks *et al.* 1992; Wiktorska *et al.* 2002).

There are also papers, reporting that hypothyreosis caused LPO suppression in the brain (Mogulkoc *et al.* 2005b; Wiktorska *et al.* 2005), in the liver (Mogulkoc *et al.* 2005b), in the heart (Mogulkoc *et al.* 2005b; Yilmaz

et al. 2003), in the testis (Mogulkoc *et al.* 2005d; Sahoo *et al.* 2008), in the kidney (Mogulkoc *et al.* 2005d) and in the thyroid (Yilmaz *et al.* 2003). Moreover, hypothyreosis in experimental animals minimises oxidative pulmonary damage, associated with the application of ionising radiation (Sener *et al.* 2006) and hyperoxia (Rodriguez-Pierce *et al.* 1994). Probably, PTU administration is advantageous only in thyrotoxicosis and in such cases which, in a certain degree, simulate the condition, leading to increased metabolism (Wiktorska *et al.* 2006b). Furthermore, Sahoo *et al.* (2008) has claim that transient hypothyroidism induced by PTU treatment for 30 days decreased LPO in rat testis, but LPO were elevated during persistent hypothyroidism (90 days treatment).

In the present study, MEL decreased CD and SB levels in lung homogenates of the animals, receiving injections of L-T₄. Also in other experiments, suppression of excessive LPO was observed in the brains (Mogulkoc *et al.* 2006; Wiktorska *et al.* 2006b), in kidneys (Mogulkoc *et al.* 2005a; Sewerynek *et al.* 1999), in the testes (Mogulkoc *et al.* 2005a) in the heart and in the liver (Mogulkoc *et al.* 2006; Popov *et al.* 2008), as well as in the blood (Baydas and Meral 2005; Popov *et al.* 2008) of rats in experimentally induced hyperthyreosis. Moreover in our laboratory, noticed that MEL decreased SB concentrations in liver and lung of rats after the treatment with different doses of iodide, but – unexpectedly – the level of SB increased in lung in the group with the highest dose of iodine in diet (Świerczyńska-Machura *et al.* 2004).

In the present study, MEL decreased the basic levels of MDA and SB in lung homogenates. It appears from earlier reports that this hormone reduced the basic LPO in the brain (Wiktorska *et al.* 2005) and in kidneys (Sewerynek *et al.* 1999).

It was observed in the performed studies that AMB had decreased MDA and SB levels in lung homogenates of rats after L-T₄ injections. Similar observations were made in brain homogenates of the rats, receiving L-T₄ (Wiktorska *et al.* 2005). It appears from the studies, performed by Nowak *et al.* (1993) that AMB reduced CD formation in the lungs and in the heart of mice, induced by administration of endotoxins. In another experiment, those authors demonstrated that a single AMB administration had brought back CD and MDA levels to their normal values in the cardiac muscle, however, it failed to protect against dexorubicine-induced LPO in the lungs and the livers of mice (Nowak *et al.* 1995). Moreover, this drug, when intraperitoneally injected to mice, reduced LPO in lung homogenates, submitted to the temperature of 50 °C and to H₂O₂ (Nowak *et al.* 1994).

It should be noted that in Experiment II, AMB decreased the basic concentration of MDA, as well as SB levels in lung homogenates. In other experiments, a decrease of the basic level of CD and SB was obtained in the brains of rats (Wiktorska *et al.* 2005) and of CD

in homogenates of the lungs, the heart and the liver in mice, following the administration of this medication (Nowak *et al.* 1994).

In our present study, NAC reduced CD level in lung homogenates of the L-T₄ receiving animals. It remains in conformity with other reports, demonstrating that this drug used to suppress LPO in the brains of L-T₄ receiving rats (Wiktorska *et al.* 2005), as well as in the hearts of rats after T₃ (Venditti *et al.* 1998). In earlier experiments, protective activity of NAC was observed in the organ in question, suppressing endotoxin (Nowak *et al.* 1993), 50°C temperature (Nowak *et al.* 1994) and nicotine (Sudheer *et al.* 2008) induced LPO.

In the present study, NAC, administered separately, decreased values of the majority of studied LPO parameters in lung homogenates. A decrease of basic CD level in homogenates of the lungs, the heart and the liver, following the administration of this drug, was also noted by Nowak *et al.* (Nowak *et al.* 1994).

The results of the performed studies indicate that: 1) the administration of L-T₄ suppresses LPO in the lungs of rats, 2) MEL, AMB and NAC effectively protect against LPO, 3) PTU is an effective antioxidant in thyreotoxicosis, however, when administered alone, it induces hypothyreosis and enhances LPO, 4) the differences in LPO, observed between the two applied control groups, suggest a stimulatory effect of stress on oxidative processes in the lungs.

ACKNOWLEDGEMENTS

The study was financially supported by the Medical University of Lodz (Grant No. 502-11-554).

REFERENCES

- Asayama K, Dobashi K, Hayashibe H, Megata Y, Kato K. (1987). Lipid peroxidation and free radical scavengers in thyroid dysfunction in the rat: A possible mechanism of injury to heart and skeletal muscle in hyperthyroidism. *Endocrinology*. **121**: 2112–2118.
- Bartosz G. (2003). *Druga Twarz Tlenu. Wolne Rodniki w Przyrodzie*. Wydawnictwo Naukowe PWN. Warszawa.
- Baydas B, Meral I. (2005). Effects of melatonin on lipid peroxidation and anti-oxidant enzyme activity in rats with experimentally induced hyperthyroidism. *Clin. Exp. Pharmacol. Physiol.* **32**: 541–544.
- Bianchi G, Solaroli E, Zaccheroni V, Grossi G, Bargossi AM, Melchionda N, Marchesini. (1999). Oxidative stress and anti-oxidant metabolites in patient with hyperthyroidism: effect of treatment. *Horm. Metab. Res.* **31**: 620–624.
- Bozhko AP, Gorodetskaia IV. (1991). The enhancement of body resistance to combined exposure to immobilization and cold with thyroid hormones. *Nauchnye. Doki. Vyss. Shkoly. Biol. Nauki*. 80–86.
- Buege JA, Aust SD. (1978). Microsomal lipid peroxidation. *Methods Enzymol.* **52**: 302–310.
- Chattopadhyay S, Zaidi G, Das K, Chainy GB. (2003). Effects of hypothyroidism induced by 6-n-propylthiouracil and its reversal by T₃ on rat heart superoxide dismutase, catalase and lipid peroxidation. *Indian J. Exp. Biol.* **41**: 846–849.
- Dumitriu L, Bartoc R, Ursu H, Purice M, Ionescu V. (1988). Significance of high levels of serum malonyldialdehyde (MDA) and ceruloplasmin (CP) in hyper- and hypothyroidism. *Endocrinologie*. **26**: 35–38.
- Erdamar H, Demirci H, Yaman H, Erbil MK, Yakar T, Sancak B, Elbeg S, Biberoglu G, Yetkin I. (2008). The effect of hypothyroidism, hyperthyroidism, and their treatment on parameters of oxidative stress and antioxidant status. *Clin. Chem. Lab. Med.* **46**: 1004–1010.
- Fernandez V, Tapia G, Varela P, Romanque P, Cartier-Ugarte D, Videla LA. (2006). Thyroid hormone-induced oxidative stress in rodents and humans: a comparative view and relation to redox regulation of gene expression. *Comp. Biochem. Physiol. C. Toxicol. Pharmacol.* **142**: 231–239.
- Gillissen A, Nowak D. (1998). Characterization of N-acetylcysteine and ambroxol in anti-oxidant therapy. *Respir. Med.* **92**: 609–623.
- Guerrero A, Pamplona R, Portero-Otin M, Barja G, Lopez-Torres M. (1999). Effect of thyroid status on lipid composition and peroxidation in the mouse liver. *Free Radic. Biol. Med.* **26**: 73–80.
- Harper ME, Seifert EL. (2008). Thyroid hormone effects on mitochondrial energetics. *Thyroid*. **18**: 145–156.
- Hicks M, Wong LS, Day RO. (1992). Antioxidant activity of propylthiouracil. *Biochem. Pharmacol.* **43**: 439–444.
- Huong NT, Murakami Y, Tohda M, Watanabe H, Matsumoto K. (2005). Social isolation stress-induced oxidative damage in mouse brain and its modulation by majonoside-R2, a Vietnamese ginseng saponin. *Biol Pharm Bull.* **28**: 1389–1393.
- Jablonka S, Ledwozyw A, Kadziolka W, Jablonka A, Nestorowicz A. (1992). The influence of Ambroxol on peroxidative processes in lung and plasma in dogs after pneumectomy. *Arch. Vet. Pol.* **32**: 57–66.
- Karbownik M, Lewinski A, Reiter RJ. (2001). Anticarcinogenic actions of melatonin which involve antioxidative processes: comparison with other antioxidants. *Int. J. Biochem. Cell. Biol.* **33**: 735–753.
- Karbownik M, Lewinski A. (2003). The role of oxidative stress in physiological and pathological processes in the thyroid gland; possible involvement in pineal-thyroid interactions. *Neuroendocrinol. Lett.* **24**: 293–303.
- Kariya K, Dawson E, Neal RA. (1983). Toxic effects of propylthiouracil in the rat. *Res. Commun. Chem. Pathol. Pharmacol.* **40**: 333–336.
- Lopez-Torres M, Pérez-Campo R, Cadenas S, Rojas C, Baria G. (1993). A comparative study of free radicals in vertebrates – II. Non-enzymatic antioxidants and oxidative stress. *Comp. Biochem. Physiol.* **105B**: 757–763.
- Manikandan S, Padma MK, Srikumar R, Jeya Parthasarathy N, Muthuvel A, Sheela Devi R. (2006). Effects of chronic noise stress on spatial memory of rats in relation to neuronal dendritic alteration and free radical-imbalance in hippocampus and medial prefrontal cortex. *Neurosci. Lett.* **399**: 17–22.
- Mano T, Sinohara R, Sawai Y, Oda N, Nishida Y, Mokumo T, Asano K, Ito Y, Kotake M, Hamada M. (1995). Changes in lipid peroxidation and free radical scavengers in the brain of hyper- and hypothyroid aged rats. *J. Endocrinol.* **147**: 361–365.
- Mogulkoc R, Baltaci AK, Oztekin E, Aydin L, Sivrikaya A. (2006). Melatonin prevents oxidant damage in various tissues of rats with hyperthyroidism. *Life Sci.* **79**: 311–315.
- Mogulkoc R, Baltaci AK, Oztekin E, Aydin L, Tuncer I. (2005a). Hyperthyroidism causes lipid peroxidation in kidney and testis tissues of rats: Protective role of melatonin. *Neuroendocrinol. Lett.* **28**: 26.
- Mogulkoc R, Baltaci AK, Aydin L, Oztekin E, Sivrikaya A. (2005b). The effect of thyroxine administration on lipid peroxidation in different tissues of rats with hypothyroidism. *Acta Physiol. Hung.* **92**: 39–46.
- Mogulkoc R, Baltaci AK, Aydin L, Oztekin E, Sivrikaya A. (2005c). Pinelectomy inhibits antioxidant system in rats with hyperthyroidism. *Neuroendocrinol. Lett.* **26**: 795–798.

- 27 Mogulkoc R, Baltaci AK, Oztekin E, Ozturk A, Sivrikaya A. (2005d). Short-term thyroxine administration leads to lipid peroxidation in renal and testicular tissues of rats with hypothyroidism. *Acta Biol. Hung.* **56**: 225–232.
- 28 Nanda N, Bobby Z, Hamide A. (2008). Association of thyroid stimulating hormone and coronary lipid risk factors with lipid peroxidation in hypothyroidism. *Clin. Chem. Lab. Med.* **46**: 674–679.
- 29 Nowak D, Antczak A, Pietras T, Białasiewicz P, Król M. (1994). Protective effect of ambroxol against heat- and hydrogen peroxide-induced damage to lung lipids in mice. *Eur. Resp. J.* **7**: 1629–1634.
- 30 Nowak D, Pierściński G, Drzewowki J. (1995). Ambroxol inhibits Doxorubicin-induced lipid peroxidation in heart of mice. *Free Rad. Biol. Med.* **19**: 659–663.
- 31 Nowak D, Pietras T, Antczak A, Król M, Piasecka G. (1993). Ambroxol inhibits endotoxin-induced lipid peroxidation in mice. *Pol. J. Pharmacol.* **45**: 317–322.
- 32 Oge A, Sozmen E, Karaoglu AO. (2004). Effect of thyroid function on LDL oxidation in hypothyroidism and hyperthyroidism. *Endocr Res.* **30**: 481–489.
- 33 Pérez-Campo R, López-Torres M, Rojas C, Cadenas S, Baria G. (1993). A comparative study of free radicals in vertebrates – I. Antioxidant enzymes. *Comp. Biochem. Physiol.* **105B**: 749–755.
- 34 Popov SS, Pashkov AN, Popova TN, Zoloedov VI, Semenikhina AB, Rakhmanova TI. (2008). Melatonin influence on free radical homeostasis in rat tissues at thyrotoxicosis. *Biomed. Khim.* **54**: 114–121.
- 35 Rahaman SO, Ghosh S, Mohanakumar KP, Das S, Sarkar PK. (2001). Hypothyroidism in the developing rat brain is associated with marked oxidative stress and aberrant intraneuronal accumulation of neurofilaments. *Neurosci. Res.* **40**: 273–279.
- 36 Rice-Evans CA, Diplock AT, Symons MCR. (1991). *Techniques in Free Radical Research*. Elsevier, Amsterdam.
- 37 Rodriguez-Pierce M, Sosenko IRS, Whitney P, Frank L. (1994). Propylthiouracil treatment decreases the susceptibility to oxygen radical-induced lung damage in newborn rats exposed to prolonged hyperoxia. *Pediatr. Res.* **34**: 530–535.
- 38 Sahoo DK, Roy A, Bhanja S, Chainy GB. (2008). Hypothyroidism impairs antioxidant defence system and testicular physiology during development and maturation. *Gen. Comp. Endocrinol.* **156**: 63–70.
- 39 Sener G, Kabasakal L, Atasoy BM, Erzik C, Velioğlu-Oğünç A, Cetinel S, Contuk G, Gedik N, Yeğen BC. (2006). Propylthiouracil-induced hypothyroidism protects ionizing radiation-induced multiple organ damage in rats. *J Endocrinol.* **189**: 257–269.
- 40 Sewerynek E, Świerczyńska-Machura D, Lewiński A. (2006). Effect of propylthiouracil on the level of Schiff's bases in tissues of rats on diet with different doses of potassium iodide. *Neuroendocrinol. Lett.* **27**: 595–599.
- 41 Sewerynek E, Wiktorska JA. (1997). Stres oksydacyjny: I. Wolne rodniki i antyoksydacyjny system obrony. *Endokrynol. Pol. – Polish J. Endocrinol.* **48**: 157–163.
- 42 Sewerynek E, Wiktorska J, Nowak D, Lewiński A. (2000). Methimazole protection against oxidative stress induced by hyperthyroidism in Graves' disease. *Endocrine Reg.* **34**: 82–89.
- 43 Sewerynek E, Wiktorska J, Lewiński A. (1999). Effects of melatonin on the oxidative stress induced by thyrotoxicosis in rats. *Neuroendocrinol. Lett.* **20**: 157–163.
- 44 Singh R, Kiloung J, Singh S, Sharma D (2008). Effect of paradoxical sleep deprivation on oxidative stress parameters in brain regions of adult and old rats. *Biogerontology.* **9**: 153–162.
- 45 Smith-Kielland A; Morland J. (1981). Reduced hepatic protein synthesis after long-term ethanol treatment in fasted rats. Dependence on animal handling before measurement. *Biochem. Pharmacol.* **30**: 2377–2379.
- 46 Sudheer AR, Muthukumaran S, Devipriya N, Devaraj H, Menon VP. (2008). Influence of ferulic acid on nicotine-induced lipid peroxidation, DNA damage and inflammation in experimental rats as compared to N-acetylcysteine. *Toxicology.* **243**: 317–329.
- 47 Świerczyńska-Machura D, Lewiński A, Sewerynek E. (2004). Melatonin effects on Schiff's base levels induced by iodide administration in rats. *Neuroendocrinol. Lett.* **25**: 70–74.
- 48 Thomas S, Pulimood A, Balasubramanian KA. (2003). Heat preconditioning prevents oxidative stress-induced damage in the intestine and lung following surgical manipulation. *Br. J. Surg.* **90**: 473–481.
- 49 Venditti P, De-Leo T, Di-Meo S. (1998). Antioxidant-sensitive shortening of ventricular action potential in hyperthyroid rats is independent of lipid peroxidation. *Mol. Cell. Endocrinol.* **142**: 15–23.
- 50 Wiktorska JA, Sewerynek E, Lewiński A. (2006a). Wpływ stanu tyreometabolicznego na proces peroksydacji lipidów (I): hipertyreoz. *Clin. Exp. Med. Lett.* **47**: 9–15.
- 51 Wiktorska JA, Sewerynek E, Lewiński A. (2006b). Wpływ stanu tyreometabolicznego na proces peroksydacji lipidów (II): hipotyreoz. *Clin. Exp. Med. Lett.* **47**: 79–83.
- 52 Wiktorska JA, Lewiński A, Sewerynek E (2005). Effects of different antioxidants on lipid peroxidation in brain homogenates induced by L-thyroxine administration in rats. *Neuroendocrinol. Lett.* **26**: 704–708.
- 53 Wiktorska JA, Lewiński A, Sewerynek E. (2002). Antyoksydacyjna rola propylotiouracylu. *Endokrynol. Pol. – Polish J. Endocrinol.* **53**: 189–196.
- 54 Yavuz DG, Yuksel M, Deyneli O, Ozen Y, Aydin H, Akalin S (2004). Association of serum paraoxonase activity with insulin sensitivity and oxidative stress in hyperthyroid and TSH-suppressed nodular goitre patients. *Clin. Endocrinol. (Oxf)*. **61**: 515–521.
- 55 Yilmaz S, Ozan S, Benzer F, Canatan H. (2003). Oxidative damage and antioxidant enzyme activities in experimental hypothyroidism. *Cell. Biochem. Funct.* **21**: 325–330.