

# Action of melatonin on bone marrow depression induced by cyclophosphamide in acute toxicity phase

Nicola PACINI, Fabio BORZIANI

“Prof. Luigi Di Bella” Private Physiology Laboratory, via S.G. Marianini n.45, 41100 Modena, Italy.

Correspondence to: Nicola Pacini,  
via Primo Maggio n.6, 50050 Stabbia Cerreto Guidi (FI), Italy.  
E-MAIL: nicolapacini77@yahoo.it

Submitted: 2009-08-09 Accepted: 2009-10-14 Published online: 2009-11-10

Key words: **melatonin; cyclophosphamide; L-ascorbic acid;  $\alpha$ -tocopheryl acetate; bone marrow; hematopoiesis; leukocyte; erythrocyte; platelet; leukopenia**

Neuroendocrinol Lett 2009; **30**(5):582–591 PMID: 20035269 NEL300509A10 © 2009 Neuroendocrinology Letters • [www.nel.edu](http://www.nel.edu)

## Abstract

**BACKGROUND:** In the course of the last few years, various studies have researched the relations linking melatonin (MLT) with hematopoiesis and the immune-hematopoietic system. Nevertheless, to date there are numerous issues still opened and many questions are yet unanswered. Much emphasis has been recently placed on the reducing role of MLT, which has been demonstrated by many studies to mitigate the genotoxic damage inflicted by various alkylating agents. Although *in vitro* MLT has shown to be effective in limiting the cytological alterations provoked by chemotherapeutic drugs, some clinical studies seem to point to a somewhat lower effectiveness in countering chemotherapy-induced leukopenia and anemia.

**OBJECTIVE:** The aim of this study was to evaluate the activity exerted by pharmacological doses of MLT in limiting leukopenia, anemia and other hemocytometric modifications in animals facing acute toxicity caused by the treatment with cyclophosphamide (CP). Moreover, we have compared the activity of MLT with well-known alpha-tocopherol/ascorbic acid antioxidant system.

**RESULTS:** Our results indicate that overall MLT exerts a remarkable countering activity towards leukopenia and anemia in the early phase of administration of CP.

**CONCLUSIONS:** Our findings suggest possible active involvement of MLT in hematopoiesis and erythrocyte- and leukocyte turnover. This paper summarizes the essential aspects of the available literature, researching the possible relations between MLT and immune-hematopoietic system.

## Abbreviations :

MLT - melatonin  
CP - cyclophosphamide  
DMSO - dimethylsulphoxide  
BM - bone marrow  
CFU-GM - colony forming units for granulocytes and macrophages  
GPCR - G-protein coupled receptors  
APUD - Amine Precursor Uptake Decarboxylase

## INTRODUCTION

Our laboratory has been studying for a long time existing relations between N-acetyl-5-methoxytryptamine or melatonin (MLT), hematopoiesis, blood physiology and the immune system (Di Bella *et al.* 1969; Di Bella *et al.* 1979, a). These studies, begun in the mid '60s by Di Bella and colleagues focused mainly on the effects of MLT on red bone marrow (BM) (Di Bella *et al.* 1976), thrombopoiesis (Di Bella *et al.* 1979, b; Rossi *et al.* 1988), platelet aggregation (Di Bella *et al.* 1980), erythropoiesis (Di Bella *et al.* 1979, a) and lymph circulation (Rossi *et al.* 1974; Di Bella & Gualano, 2006). Subsequent contributions have been made by various authors (Haldar *et al.* 1992, a; Maestroni, 1998; Maestroni, 1999; Labunets *et al.* 2004) on the existence of a link between MLT, hematopoiesis, functions of blood and immunocompetent cells. However, despite intensive research done in this area, there are still many aspects which need to be clarified concerning the role of MLT in hematopoiesis and blood physiology.

MLT is the main product of the pineal gland or epiphysis. It is well known that its maximum secretion and plasmatic peak is reached during the darkness of night. However, in recent years, studies have highlighted its presence and its *de novo* synthesis also in extrapineal organs (Kvetnoy, 1999), including hematopoietic and lymphoid organs, the BM and the thymus (Conti *et al.* 2000; Naranjo *et al.* 2007), and the hematopoietic and differentiated immunocompetent cells, i.e. mononuclear phagocytes, natural killer cells, eosinophils, platelets and, according to some evidence, even red blood cells (Rosengarten *et al.* 1972; Launay *et al.* 1982; Finocchiaro *et al.* 1988; Champier *et al.* 1997; Tan *et al.* 1999; Carrillo-Vico *et al.* 2004; Morera & Abreu, 2005). MLT is also said to be present in organs unrelated to the hematopoietic, immunocompetent and hematologic system, like retina, ovaries and various other organs (Kvetnoy *et al.* 1999; Jaworek *et al.* 2007; Konturek *et al.* 2007). It is also synthesised in the enterochromaffin cells (Raikhlina & Kvetnoy, 1976), in cells belonging to the APUD system and even in structures not belonging to the diffuse neuroendocrine system, so much so that some authors question the real belonging of MLT to the endocrine system in the classic, strict sense of the word. Moreover, the concentrations of extrapineal MLT are not always correlated to the sleep/wake cycle: for example, despite most plasmatic MLT deriving from pineal secretion, in platelets it is synthesised and accumulated independently of the photoperiod (Reiter & Tan, 2003; Morera & Abreu, 2005; Jaworek *et al.* 2007; Konturek *et al.* 2007).

Because of the relatively high concentrations of MLT in the BM (Tan *et al.* 1999), and in many hematopoietic and immunocompetent cells, the hypothesis of a real and active involvement of MLT in hematopoiesis and in immune hematological processes seems to be at least plausible. MLT present in the BM, mononu-

clear phagocytes, platelets, T lymphocytes, thymus and other lymphoid organs appears to be produced and synthesised *in situ* (Rosengarten *et al.* 1972; Launay *et al.* 1982; Finocchiaro *et al.* 1988; Champier *et al.* 1997; Tan *et al.* 1999; Conti *et al.* 2000; Carrillo-Vico *et al.* 2004; Morera & Abreu, 2005; Naranjo *et al.* 2007). Therefore it seems possible that MLT may also act as an autocrine/paracrine factor in the physiology and biochemical dynamics of the BM and of the immune-hematopoietic system, the hypothesis presented also by other authors (Carrillo-Vico *et al.* 2004; Markus *et al.* 2007). This possibility is further supported by the fact that many cells of this system, including platelets and T lymphocytes, synthesise MLT and express its receptors (Vacas *et al.* 1972; Di Bella *et al.* 1979, c; Pozo *et al.* 1997; Drazen *et al.* 2001; Yau *et al.* 2002; Regodón *et al.* 2009). Similarly, in many extra-pineal locations in which it is synthesised, it is supposed to act as a local factor. Despite numerous studies performed on differentiated blood cells and documentation of an independent immune regulation (Carrillo-Vico *et al.* 2005; Szczezanik, 2007), platelet antiaggregation (Di Bella *et al.* 1979, d; Leach & Thorburn, 1980; Kornblihtt *et al.* 1983; Cardinali *et al.* 1993; Di Bella & Gualano, 2006), and protective effect on the erythrocyte membrane (Tesorieri *et al.* 1999; Kedziora-Kornatowska *et al.* 2007), a lot of information still needs to be uncovered about its role in hematopoiesis and in differentiative processes.

MLT has been reported to be useful in the stimulation and regulation of immune-hematopoietic processes: in preliminary clinical trials it has shown a marked tropism for the hematopoietic system, revealing the capacity to attenuate the extent of thrombocytopenia in patients undergoing chemotherapy and radiotherapy (Lissoni *et al.* 1995, a; Lissoni *et al.* 1995, b; Lissoni *et al.* 1996; Lissoni, 2007; Todisco, 2007), and to act directly on the immune responses by stimulating the CD4<sup>+</sup> T helper cells (Maestroni, 1995), which express MLT receptors (Pozo *et al.* 1997; Drazen *et al.* 2001; Carrillo-Vico *et al.* 2003). It has been proven that MLT may significantly influence the pool of cytokines secreted, which in turn influence the activity of helper T cells of both the Th1 and Th2 subtypes (Maestroni, 1995; Liu *et al.* 2001; Carrillo-Vico *et al.* 2003). Although its effects on the immune system appear to be particularly complex and in some instances opposed (Majewska *et al.* 2007), it seems that MLT may also act as a stimulant of cell-mediated immunity, showing a specific modulatory activity on the immune system (Maestroni, 1995; Liu *et al.* 2001; Santello *et al.* 2008). It is also interesting to note the MLT capability of *in vitro* regulation of the proliferation of various lines of lymphoblasts, both normal and transformed, inhibiting the incorporation of tritiated thymidine (Persengiev & Kyurkchiev, 1993). MLT is also active in the inhibition of proliferation of human B lymphoma cells and other tumours of the hematopoietic system (Yu *et al.* 2000; Trubiani *et al.* 2005; Rubio *et al.* 2007; Bejarano *et al.* 2009).

Overall, an immune stimulating effect of MLT has been widely reported, suggesting its use in patients suffering from acquired immune deficiency (Lissoni *et al.* 1995, c; Kast & Altschuler, 2006). It is interesting to note how this stimulation of the immune system has been connected with the genesis of various autoimmune diseases (Sandyk, 1993). Research on heterologous transplants in rats and in subjects affected by rheumatoid arthritis and other immune disorders (Kang *et al.* 2001; Jung *et al.* 2004; Cardell *et al.* 2008; Korkmaz, 2008) suggests that MLT, rather than a generic stimulating influence on immune processes, has a real and direct regulating and modulating role of the immune response: *in vitro* studies showed that MLT is efficacious in the inhibition of proliferation and the induction of apoptosis in human lymphoma cell lines (Trubiani *et al.* 2005; Bejarano *et al.* 2009), as well as useful in individuals suffering from myelodysplasia (Viviani *et al.* 1990). It should be noted that such effects have been ascribed to the induction or the block of apoptosis. Since the latter plays a central role in immunological and hematopoietic processes (Cacciapaglia *et al.* 2009; Droin *et al.* 2009), and since MLT has been widely proven to be able to modulate the apoptotic process in a biphasic mode (Lissoni *et al.* 1995, c; Mayo *et al.* 1998; Yu *et al.* 2000; Scott *et al.* 2001; Trubiani *et al.* 2005; Cheng *et al.* 2006; Feng *et al.* 2006; Rubio *et al.* 2007; Bejarano *et al.* 2009), it cannot be excluded that in addition to its role in the secretion of cytokines (Liu *et al.* 2001) and in endocrine secretion (Mauri *et al.* 1985; Lissoni *et al.* 1987; Aleandri *et al.* 1996; Baltaci *et al.* 2004; Mogulkoc *et al.* 2006), it may also act directly by modulating the apoptotic process during immune responses and hematopoiesis. Such effects appear to rely on its physiological or pharmacological concentrations, and the manner in which the substance is administered.

Moreover some evidence suggests benefit effect of MLT in reduction of anaemia and hemoglobin deficiency in elderly patients chronically treated with MLT (Herrera *et al.* 2001; Hörl, 2002; Labonia *et al.* 2005; Arushanian *et al.* 2006). In mice and Wistar rats MLT has shown to be particularly active in reducing damage to the BM induced by various chemicals and genotoxic agents (Melchiorri *et al.* 1998; Rapozzi *et al.* 1998). MLT showed to be active in reducing cytotoxic damage to the BM in mice after irradiation with gamma rays (Vijayalaxmi *et al.* 1999), but also in increasing the number of circulating blood cells in rats (Koc *et al.* 2002), suggesting that it may act as a regulatory and stimulating factor on macrophages and granulocytes (CUF-GM), thereby fully confirming the hypothesis previously proposed by Haldar, Häussler and Gupta (1992, a; 1992, b). Recently, a similar effect has been suggested by Akbulut *et al.* (1999), while Sutherland *et al.* (2002) have pointed out a possible direct role of MLT in leukocytopoiesis and in the release of white blood cells into circulation.

The reducing properties of MLT has been well documented for years, showing it to be an excellent antioxidant even though the precise mechanism has not yet been fully understood. It seems plausible that this involves the oxidation of carbon 2 on the indolic nucleus with formation of a carbocationic intermediate and the opening of the indolic nucleus with the subsequent formation of kynurenic compounds (Hirata *et al.* 1974; Reiter *et al.* 1999), which also have reducing capability (Goda *et al.* 1999). Widespread research has demonstrated this important antioxidant activity (Sandyk, 1990; Ianăș *et al.* 1991; Pieri *et al.* 1994; Acuña-Castroviejo *et al.* 2001; Reiter *et al.* 2001, a; Martin *et al.* 2002; Baydas *et al.* 2003; Kaptanoglu *et al.* 2003; Leaden & Catalá, 2007), mainly because MLT appears particularly active against hydroxyl radicals, a chemical species particularly injurious to cellular functions and implicated in many physiopathological processes (Poeggeler *et al.* 1993; Brömme *et al.* 2000). Moreover, it has been highlighted the action of MLT in the regulation of gene expression and activity of many antioxidant enzymes (Mayo *et al.* 2002).

MLT is widely expressed throughout the world of living organisms, from animals to plants, whether metazoans or non-metazoans (Korf, 1994; Shedpure & Pati, 1995; Hardeband, 1999; Reiter *et al.* 2001, b; Falcón *et al.* 2009), probably thanks to its marked reducing power, a characteristic which has most likely contributed to its evolutionary success. For this powerful antioxidant role, a finalistic hypothesis has been advanced according to which the high concentration of MLT found in the BM is proposed to have the function of oxidative stress reduction in a metabolically highly active tissue subjected to an extremely high cellular turnover (Tan *et al.* 1999). Thanks to its reducing action it is thought to be able to regulate peroxidase activity of neutrophil and eosinophil granulocytes (Allegra *et al.* 2001; Lu *et al.* 2008).

Notwithstanding this clear and very important role in limiting cellular oxidation, many studies have emphasised the presence of specific MLT receptors, both belonging to the family of G-protein coupled receptors (GPCR) and to that of the nuclear receptors within cells of the immune-hematopoietic system. It should be noted that these receptors are present in platelets, T helper lymphocytes, macrophages, the spleen and thymus (Vacas *et al.* 1992; Pozo *et al.* 1997; Drazen *et al.* 2001; Liu *et al.* 2001; Regodón *et al.* 2009). It is therefore very likely that MLT receptors are present in the differentiated compartment of the immune-hematopoietic system.

In our laboratory we demonstrated and documented the activity of MLT in modulating and attenuating the outward K<sup>+</sup> currents across the megakaryocyte membrane (Di Bella *et al.* 2002). Other evidence suggests its role in many electric transduction processes, both through the direct effect of MLT on ionic channels and through actions mediated by the activation of the

GPCR receptors. It is interesting to note how Mel1 MLT receptor forms dimers with potassium channels at the CNS level (Huan *et al.* 2001; Zhou *et al.* 2003; Hu *et al.* 2005), while in lymphocyte MLT binds to Kv1.3 potassium channels leading to inactivation of ionic currents (Varga *et al.* 2001). In recent years several studies have highlighted the importance of electrophysiological phenomena in the differentiating processes of cells in the immune-hematopoietic system. It cannot be ruled out that amongst its multiple signalling modes there is also an important electrophysiological mechanism.

In the light of these multiple physiological implications, one can easily imagine the difficulties still faced today in understanding the various activities of MLT. Its ubiquitous presence in the organs and systems suggests its dual mode of signalling and acting: systemic on the one hand, because of its pineal secretion, and on the other hand autocrine/paracrine, as a consequence of its local synthesis and activity. It has marked reducing properties and has a vast range of cellular receptors both in the strict sense and in non-classical binding sites (Varga *et al.* 2001). The above mentioned facts is the reason why the complete mechanism of its action in hematopoiesis and in many other processes is not yet fully known.

Effect of MLT on differentiated blood and immunocompetent cells appears to be plausible, though not demonstrated. It remains mostly to be defined as regards to its possible role in the differentiating processes of the hematopoietic compartment such as hematopoietic and primary lymphoid organs.

## OBJECTIVES

The aim of the current study was to assess the effects of MLT at pharmacological doses on limiting leukopenia and anaemia in young rats induced by CP during acute toxicity test at 12, 48 and 72 hours after administration.

CP is a prodrug which is activated through microsomal hydroxylation by cytochrome P450. Once hydroxylated in position 4, the molecule undergoes a non-enzymatic cleavage with the formation of acrolein and phosphoramidic mustard. The latter release orthophosphoric acid and ammonia leading to the formation of nitrogen mustard, a compound which shows an alkylating action towards the N7 position of guanine.

In the rat, unlike in man, at doses of over 50 mg/kg bw CP rapidly induces leukopenia and anaemia just 12–48 hours post-treatment (Paterson & Hanson, 1969; Wheeler *et al.* 2004). Specifically, Wistar and Lewis rats appear to be particularly susceptible to the alkylating action of this drug. Despite various studies have underlined the cytoprotective role of MLT against various genotoxic and alkylating agents (Caroleo *et al.* 1994; Melchiorri *et al.* 1998; Rapozzi *et al.* 1998; Vijayalaxmi *et al.* 1999; Koc *et al.* 2002; Lialiaris *et al.* 2008), and despite the fact that some studies have even shown a possible clinical use in reducing the toxic effects of

chemotherapy (Lissoni *et al.* 1995, a; Lissoni *et al.* 1995, b; Lissoni *et al.* 1996), other clinical studies have put forward the absence of efficacy of MLT in reducing leukopenia and anaemia in patients subjected to anti-cancer chemotherapy, suggesting however its protective role in reducing oxidative stress (Ghielmini *et al.* 1999; Sarma *et al.* 2004).

On the basis of the above considerations, we wondered if, in the short term, MLT could possibly have a protective role against leukopenia and in general against hematic cytopenia induced by treatment with CP, bearing in mind that, longer term, the onset of severe toxic consequences may decrease MLT's trophic effect on the BM. The protective role of MLT was earlier suggested by studies performed on animals irradiated and contemporaneously treated with MLT (Vijayalaxmi *et al.* 1999; Koc *et al.* 2002).

In our study MLT was administered at a dose of 10 mg/kg bw to assess its effect at 12 and 48 hours, while subsequent evaluation at 72 hour post-treatment was carried out following further administrations of MLT at 24 and 48 hours after the first administration. As some of the properties of MLT can be attributed to its reducing action, a parallel evaluation at 72 hours was carried out to determine the action of L-ascorbic acid (vitamin C) and of  $\alpha$ -tocopheryl acetate (vitamin E).

The qualitative and quantitative composition of the blood was studied via a hemocytometric analysis and an examination with the classic Romanowski staining.

## METHODS

In this study 50–65 days old Wistar rats, weighting  $220 \pm 16$  g were utilized. These rats came from breeders specialised in supplying animals for laboratory research. Once in our laboratory, the rats were reared and treated with the utmost respect for their physiology as specified in the relative guidelines: they were fed *ad libitum* with a standard balanced diet and kept in an environment with alternation of light/dark of 12 hours at a temperature of 20–22 °C.

MLT (Sigma Aldrich, purity 98%) was solubilised in dimethylsulphoxide (DMSO) (Sigma Aldrich, purity 98%) and brought up to volume with sterile physiological solution (NaCl 0,9%). Solutions were prepared containing 200 mg/100 ml of MLT in saline solution with 2% w/V DMSO.

CP (Baxter, purity 98%) was dissolved directly in sterile physiological solution. Solutions were prepared containing 500 mg/100 ml of CP in saline solution. These were administered immediately after preparation and care was taken to maintain them at a temperature below 25°C.

L-ascorbic acid (vitamin C) (Roche, purity 98%) was solubilised in sterile physiological solution. Solutions were prepared containing 200 mg/100 ml of vitamin C in saline solution with 2% w/V DMSO.

( $\pm$ )  $\alpha$ -tocopheryl acetate (vitamin E) (Roche, purity 98%) was used with no further preparatory modifications.

MLT, CP and vitamin C were administered subcutaneously in the dorsal panniculus adiposus. Vitamin E was administered orally by gavage.

To assess the effects of MLT on BM depression induced by CP, four groups of randomised rats were used. Two groups of animals received MLT at a dose of 10 mg/kg bw and CP at a dose of 75 mg/kg bw. An hour elapsed between the two administrations. The remaining two groups were administered a sterile physiological solution of 2% w/V DMSO and CP at a dose of 75 mg/kg bw. Similarly to the previous, an hour elapsed between the two administrations.

No animals showed evident signs of idiosyncrasy connected with the various administrations.

After 12 hours samples were taken from two groups of rats, the first treated with CP only and the second with both MLT and CP: the rats were anaesthetised with diethyl ether and subsequently 2–3 ml of blood were taken from the left ventricle of each rat using micro-perfusion needles. Blood was collected into sterile test tubes containing EDTA and temporarily maintained at +4°C. Blood samples were analysed within 6 hours from collection at the Bioanalisi Laboratory in Modena.

A slide for each sample was also prepared according to the classic May-Grunewald-Giemsa staining method: this enabled both direct cytological evaluation of the samples under examination and a double-check of the figures given by the device. Blood slides were subsequently analysed in our laboratory using a Zeiss optic photomicroscope.

The same sampling and laboratory analysis methods were used for the remaining two groups of rats in relation to the 48 hour assessment.

Following this session of study a new work was planned to evaluate the effects of repeated administrations of MLT on BM depression again induced by CP. The effect of antioxidant vitamins on BM depression induced by CP was also investigated.

Three groups of randomised rats were used. The first group of animals received MLT at a dose of 10 mg/kg bw and CP at a dose of 75 mg/kg bw with an hour elapsing between the two administrations. The second group received vitamin C at a dose of 7.6 mg/kg bw and CP at a dose of 75 mg/kg bw. Similarly, an hour elapsed between the two administrations. This group of rats received, in addition, vitamin E at a dose of 30 mg/kg bw. The third group received a sterile physiological solution of 2% w/V DMSO and CP at a dose of 75 mg/kg bw. In this case too, an hour elapsed between the two administrations.

After 24 and 48 hours had elapsed since this first treatment, administrations of MLT at a dose of 10 mg/kg bw were repeated for the first group of animals, vitamin C at a dose of 7.6 mg/kg bw and vitamin E at a dose of 30 mg/kg bw for the second group of rats, and a

sterile physiological solution of 2% w/V DMSO for the third group.

No animals showed evident signs of idiosyncrasy connected with the various administrations.

Seventy-two hours after first administration the test samples were taken from the three groups of rats using the same procedures as described previously.

Statistical analysis of data was carried out using a two-tailed Student t-test for unpaired data. Any variations with  $p < 0.05$  were considered significant.

## RESULTS

### Results after 12 hours

At 12 hours a significant difference between the two groups can be clearly seen in the number of leukocytes: the group treated with both MLT and CP shows a 47% increase in the number of white blood cells compared to that treated with CP alone (Table 1). The Student distribution shows a level of significance with  $p < 0.05$ .

### Results after 48 hours

At 48 hours the difference in the rate of circulating leukocytes appears much less marked than at 12 hours, despite two rats presenting a leukocyte value of over 5000/mm<sup>3</sup> in the group treated with MLT. A significant increase of 16% is instead detected in the number of red blood cells in the group of rats treated with MLT compared to that treated with CP only (Table 2 and 3). The Student distribution indicates a level of significance with  $p < 0.001$  and it is also interesting to note the concurring variations in hemoglobin rate, which in the group treated with MLT is 13% higher ( $p < 0.001$ ). In the same way the group of rats treated with both MLT and CP shows an increase of 15% in hematocrit ( $p < 0.001$ ) compared to the group treated with CP only.

### Results after 72 hours

At 72 hours the hemocytometric analysis does not show statistically significant variations, although, similarly to the previous tests, a greater value of red blood cells corresponds to the MLT treated group, though this is not as great as the corresponding value at 48 hour. In addition, the number of circulating lymphocytes is slightly higher, both in the group treated with MLT and in the group treated with vitamins C and E, respectively +8.1% and +6.5% ( $p < 0.05$ ). With regard to the remaining values, the three groups show fairly homogenous results.

## DISCUSSION

The data outlined above lead to a number of interesting observations. First of all, a single administration of MLT shows itself to be considerably active in delaying the effects of CP, completely annulling leukopenia in the initial hours after treatment, while with the passage of time its effects on leukopenia diminish. It also seems

**Tab. 1.** Hemocytometric analysis after 12 hours.

	CP	CP + MLT
<b>WBC</b> (/mm <sup>3</sup> )	5752.7 ± 577	8460.0 ± 1170
<b>Lymphocytes</b> (%)	80.6 ± 1.3	80.4 ± 1.2
<b>RBC</b> (10 <sup>6</sup> /mm <sup>3</sup> )	6.876 ± 0.194	7.063 ± 0.197
<b>HGB</b> (g/dl)	13.15 ± 0.3	12.20 ± 1.3
<b>HCT</b> (%)	38.63 ± 0.9	39.23 ± 0.9
<b>PLT</b> (10 <sup>3</sup> /mm <sup>3</sup> )	976.2 ± 42.6	957.8 ± 86.2

**Tab. 2.** Hemocytometric analysis after 48 hours.

	CP	CP + MLT
<b>WBC</b> (/mm <sup>3</sup> )	3207.7 ± 262.5	3246.7 ± 338
<b>Lymphocytes</b> (%)	74.7 ± 2.0	73.1 ± 2.4
<b>RBC</b> (10 <sup>6</sup> /mm <sup>3</sup> )	6.005 ± 0.181	6.981 ± 0.160
<b>HGB</b> (g/dl)	11.67 ± 0.216	13.24 ± 0.3
<b>HCT</b> (%)	33.50 ± 0.6	38.62 ± 0.8
<b>PLT</b> (10 <sup>3</sup> /mm <sup>3</sup> )	820.5 ± 56.9	752.9 ± 71.0

**Tab. 3.** Hemocytometric analysis after 72 hours.

	CP	CP + MLT	CP + Vitamins C and E
<b>WBC</b> (/mm <sup>3</sup> )	1154.5 ± 162	1118.2 ± 160	1109.1 ± 131
<b>Lymphocytes</b> (%)	80.03 ± 2.4	86.50 ± 1.2	85.22 ± 1.0
<b>RBC</b> (10 <sup>6</sup> /mm <sup>3</sup> )	6.468 ± 0.137	6.731 ± 0.146	6.499 ± 0.150
<b>HGB</b> (g/dl)	12.32 ± 0.9	12.37 ± 0.2	12.28 ± 0.1
<b>HCT</b> (%)	35.61 ± 0.8	36.53 ± 0.7	35.63 ± 0.7
<b>PLT</b> (10 <sup>3</sup> /mm <sup>3</sup> )	796.2 ± 30.5	783.9 ± 34.7	791.8 ± 32.4

interesting to note its marked action in augmenting the number of circulating red blood cells, an effect which reaches its maximum expression after single administration, 48 hours post-treatment and remains evident in all MLT treated rats albeit to different extents.

Experimental data from the trial with triple administration of MLT during 72 hours did not differ from single dose treatment, suggesting that repeated administration does not affect the results. Similarly, treatment with other reducing substances at pharmacological doses does not appear to significantly influence the action of CP, suggesting that an increase in reducing activity does not influence the effects of CP. However, it should be pointed out that at 72 hours both groups, treated with MLT and treated with antioxidant vitamins, significantly increased the level of circulating lymphocytes, a phenomenon which can be attributed to effects on leukocyte diapedesis. It is interesting to note how vitamin E reduces neutrophil diapedesis, by reducing the expression of P-selectin in the coronary endothelium of rats and ischemic patients subjected to angioplasty (Formigli *et al.* 1997).

These data together indicate that neither does MLT act on the alkylating activity of CP, nor does it reduce the antiproliferative potential of the drug. In the same way it does not influence the BM depression induced by the medication, a conclusion which is in agreement with the work of Ghielmini *et al.* (1999), nor does it seem that its protective action in relation to leukopenia are increased by repeated administrations.

The power of MLT to considerably diminish the toxic effects of CP on BM in the initial hours after administration seems to be, however, evident. The phenomenon observed might suggest that MLT plays a role

in stimulating granulocytogenesis and the activity of T lymphocytes, hypothesising that initially a stimulating effect on the formation of CFU-GM may prevail, the effect suggested by Halder, Häussler and Gupta (1992, a; 1992, b) and other authors as well (Akbulut *et al.* 1999; Currier *et al.* 2000; Sutherland *et al.* 2002). A possible myelostimulating action of MLT might explain a similar modification in the early hours, in a way similar to what was observed in total BM irradiation studies (Koc *et al.* 2002). Since CP expresses its pharmacological effects by stimulating the apoptotic process, and considering that in many studies MLT has shown that it has a modulating action on apoptosis, probably by acting on the mitochondrial membrane, it may be hypothesised that at an initial stage, when the myelosuppressive effect of CP is observed, MLT delays the apoptotic process.

However, a direct effect on leukocyte diapedesis and on chemotaxis cannot be excluded, considering the fact that the plasmatic leukocyte pool is in equilibrium with pool in the tissue and the lymph circulation. Such possibility seems likely. On the other hand MLT seems to be able to modulate chemotaxis and endothelial permeability (Lotufo *et al.* 2006; Peña *et al.* 2007), as well as the expression and regulation of certain adhesion proteins (Kang *et al.* 2001). Similarly, an action of MLT on leukocyte turnover can not be excluded.

The important action of MLT in reducing CP-induced anaemia appears to be evident, the effect which was observed in all three experiments and which proves to reach its peak 48 hours after administration of the substances. Considering this fact, the plasma erythrocytic pool is in direct equilibrium with that of the BM, and to a much lesser extent with that of the splenic pool. These data appear to be significant in indicating

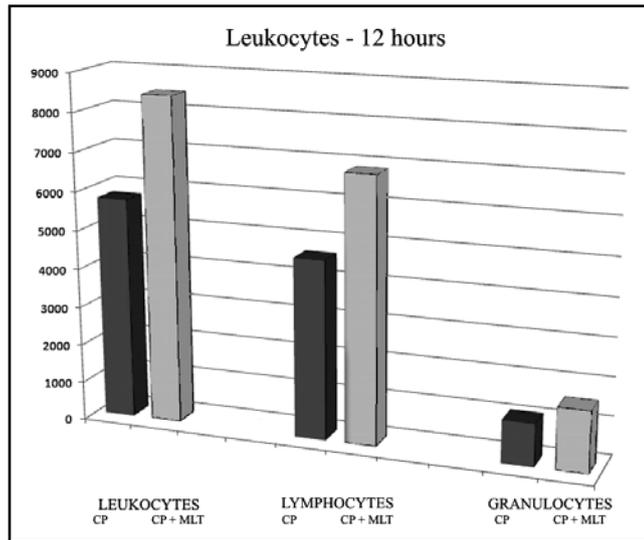


Fig. 1. Leukocytes count 12 hours after administration of CP.

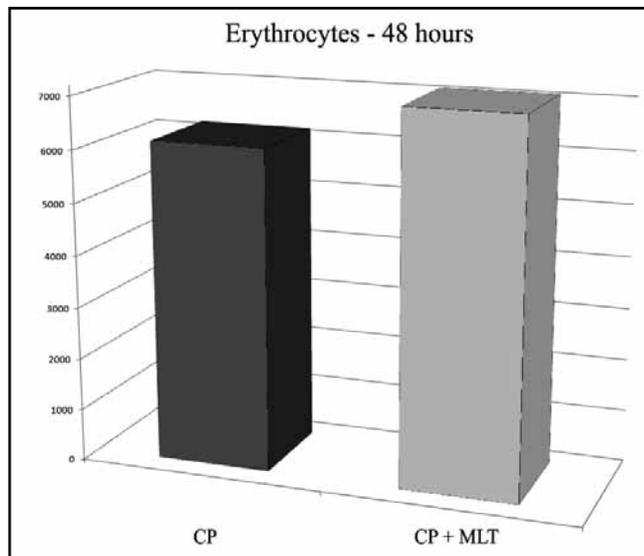


Fig. 2. Erythrocytes count 48 hours after administration of CP.

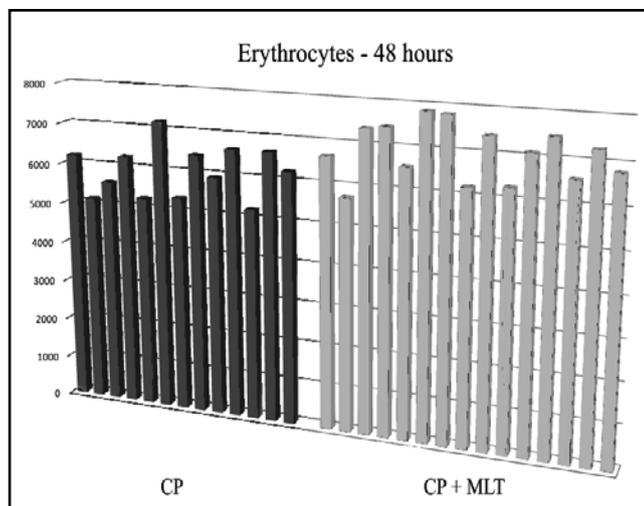


Fig. 3. Erythrocytes count in individuals 48 hours after administration of CP.

an action of MLT in stimulating erythropoiesis, or its direct effect on erythrocyte turnover.

An effective increase in the rate of circulating erythrocytes is also confirmed by the constant increase of the hematocrit and other hemocytometric parameters. The hypothesis of an action of MLT on erythropoiesis and/or on erythrocyte turnover does not seem impossible, especially when we consider the fact that nephrectomy in rats is associated not only with the evident reduction of erythropoietin but also with the massive fall in the levels of MLT, which in turn are normalised following administration of erythropoietin (Vaziri *et al.* 1996). In other clinical studies MLT has proved to be useful in reducing oxidative stress which follows the administration of erythropoietin and ferrous gluconate (Herrera *et al.* 2001; Hörl, 2002), while according to some evidence, chronic treatment with MLT induces an increase of circulating erythrocytes and hemoglobin in the elderly (Arushanian *et al.* 2006). Similarly, chronic administration of MLT in patients undergoing dialysis was shown to be useful in reducing anaemia and in preventing imbalance of iron metabolism (Labonia *et al.* 2005).

It is not superfluous to keep in mind that on circulating erythrocytes MLT was revealed to be efficacious in increasing the levels of reduced glutathione and decreasing those of glutathione reductase, highlighting a marked reduction of the erythrocyte membrane lipid peroxidation (Tesoriere *et al.* 1999; Kedziora-Kornatowska *et al.* 2007). Consequently, it seems that MLT may act to modulate the deformability of the erythrocyte membrane (Yerer *et al.* 2003). It should also be remembered that MLT is able to block erythrocyte carbonic anhydrase (Beydemir & Gülçin, 2004) and to stimulate the action of glucose-6-phosphate dehydrogenase (Ciftçi *et al.* 2001).

From the data acquired in this trial, and from the data published in the literature, it appears probable that MLT may affect erythropoietic process and/or erythrocyte turnover, as indeed suggested in previous studies conducted in our laboratory.

## CONCLUSION

The present study is indicative in confirming the action of MLT on the hematopoietic system, suggesting its involvement not only in leukopoiesis, but also in erythropoiesis and/or in erythrocyte turnover. It is likely that the actions of MLT on the BM are multiple and heterogeneous, exerted through various antioxidant and receptor mechanisms as well as electrophysiological modulation. Lastly, it is clear that MLT significantly influences the effects of CP treatment, but only in the short post-administration period of the CP. Further studies will be needed to understand this phenomenon, which in any case seems indicative of the complex action of MLT, not just connected with its reducing potential, an activity which seems to be influenced by temporal interactions yet to be defined.

## REFERENCES

- 1 Acuña-Castroviejo D, Martín M, Macías M *et al.* (2001). Melatonin, mitochondria, and cellular bioenergetics. *J Pineal Res.* **30**(2): 65–74.
- 2 Akbulut H, Icli F, Büyükelcik A *et al.* (1999). The role of granulocyte-macrophage-colony stimulating factor, cortisol, and melatonin in the regulation of the circadian rhythms of peripheral blood cells in healthy volunteers and patients with breast cancer. *J Pineal Res.* **26**(1): 1–8.
- 3 Aleandri V, Spina V, Morini A (1996). The pineal gland and reproduction. *Hum Reprod Update.* **2**(3): 225–35.
- 4 Allegra M, Furtmüller PG, Regelsberger G *et al.* (2001). Mechanism of reaction of melatonin with human myeloperoxidase. *Biochem Biophys Res Commun.* **282**(2): 380–6.
- 5 Arushanian EB, Beier EV, Mastiagina OA *et al.* (2006). Melatonin effect on the hematological indices of healthy humans. *Eksp Klin Farmakol.* **69**(5): 36–8.
- 6 Baltaci AK, Mogulkoc R, Kul A *et al.* (2004). Opposite effects of zinc and melatonin on thyroid hormones in rats. *Toxicology.* **195**(1): 69–75.
- 7 Baydas G, Kutlu S, Naziroglu M *et al.* (2003). Inhibitory effects of melatonin on neural lipid peroxidation induced by intracerebroventricularly administered homocysteine. *J Pineal Res.* **34**(1): 36–9.
- 8 Bejarano I, Redondo PC, Espino J *et al.* (2009). Melatonin induces mitochondrial-mediated apoptosis in human myeloid HL-60 cells. *J Pineal Res.* **46**(4): 392–400.
- 9 Beydemir S, Gülçin I (2004). Effects of melatonin on carbonic anhydrase from human erythrocytes in vitro and from rat erythrocytes in vivo. *J Enzyme Inhib Med Chem.* **19**(2): 193–7.
- 10 Brömme HJ, Mörke W, Peschke D *et al.* (2000). Scavenging effect of melatonin on hydroxyl radicals generated by alloxan. *J Pineal Res.* **29**(4): 201–8.
- 11 Cacciapaglia F, Spadaccio C, Chello M *et al.* (2009). Apoptotic molecular mechanisms implicated in autoimmune diseases. *Eur Rev Med Pharmacol Sci.* **13**(1): 23–40.
- 12 Cardell M, Jung FJ, Zhai W *et al.* (2008). Acute allograft rejection and immunosuppression: influence on endogenous melatonin secretion. *J Pineal Res.* **44**(3): 261–6.
- 13 Cardinali DP, Del Zar MM, Vacas MI (1993). The effects of melatonin in human platelets. *Acta Physiol Pharmacol Ther Latinoam.* **43**(1–2): 1–13.
- 14 Caroleo MC, Doria G, Nisticò G (1994). Melatonin restores immunodepression in aged and cyclophosphamide-treated Mice. *Ann N Y Acad Sci.* **719**: 343–52.
- 15 Carrillo-Vico A, García-Peñañeda A, Naji L *et al.* (2003). Expression of membrane and nuclear melatonin receptor mRNA and protein in the mouse immune system. *Cell Mol Life Sci.* **60**(10): 2272–8.
- 16 Carrillo-Vico A, Calvo JR, Abreu P *et al.* (2004). Evidence of melatonin synthesis by human lymphocytes and its physiological significance: possible role as intracrine, autocrine, and/or paracrine substance. *Faseb J.* **18**(3): 537–9.
- 17 Carrillo-Vico A, Guerrero JM, Lardone PJ, Reiter RJ (2005). A review of the multiple actions of melatonin on the immune system. *Endocrine.* **27**(2): 189–200.
- 18 Champier J, Claustrat B, Besançon R, *et al.* (1997). Evidence for tryptophan hydroxylase and hydroxy-indol-O-methyl-transferase mRNAs in human blood platelets. *Life Sci.* **60**(24): 2191–7.
- 19 Cheng Y, Feng Z, Zhang QZ, Zhang JT (2006). Beneficial effects of melatonin in experimental models of Alzheimer disease. *Acta Pharmacol Sin.* **27**(2): 129–39.
- 20 Ciftçi M, Bilici D, Küfrevioğlu OI (2001). Effects of melatonin on enzyme activities of glucose-6-phosphate dehydrogenase from human erythrocytes in vitro and from rat erythrocytes in vivo. *Pharmacol Res.* **44**(1): 7–11.
- 21 Conti A, Conconi S, Hertens E *et al.* (2000). Evidence for melatonin synthesis in mouse and human bone marrow cells. *J Pineal Res.* **28**(4): 193–202.
- 22 Currier NL, Sun LZ, Miller SC (2000). Exogenous melatonin: quantitative enhancement in vivo of cells mediating non-specific immunity. *J Neuroimmunol.* **104**(2): 101–8.
- 23 Di Bella L, Rossi MT, Pellegrino N *et al.* (1969). Ruolo del sistema abenulo-epifisario nella regolazione del tasso piastrinico. *Boll Soc Ital Biol Sper.* Vol. XLV, N. 20 bis.
- 24 Di Bella L, Rossi MT, Scalera G (1979, a). Perspectives in pineal functions. *Prog Brain Res.* **52**: 475–8.
- 25 Di Bella L, Rossi MT, Gualano L, Scalera G (1979, b). Effect of the simultaneous action of melatonin and ADP in megakaryocytes in vitro. *Boll Soc Ital Biol Sper.* **55**(5): 389–93.
- 26 Di Bella L, Gualano L, Rossi MT, Scalera G (1979, c). The action of melatonin (MLT) on platelet metabolism in vitro. *Boll Soc Ital Biol Sper.* **55**(4): 323–6.
- 27 Di Bella L, Scalera G, Rossi MT, Gualano L (1979, d). L'Aggregazione Piastrinica in presenza di melatonina. *Bollettino società italiana di fisiologia. Riunione primaverile dell'anno sociale 1978/79.*
- 28 Di Bella L, Rossi MT (1980). Red Blood cells generation and melatonin. *International Symposium on melatonin. Bremen, 28–30 September 1980.*
- 29 Di Bella L, Bruschi C, Gualano L (2002). Melatonin effects on megakaryocyte membrane patch-clamp outward K<sup>+</sup> current. *Med Sci Monit.* **8**(12): BR527–31.
- 30 Di Bella L, Gualano L (2006). Key aspects of melatonin physiology: thirty years of research. *Neuro Endocrinol Lett.* **27**(4): 425–32.
- 31 Drazen DL, Bilu D, Bilbo SD, Nelson RJ (2001). Melatonin enhancement of splenocyte proliferation is attenuated by luzindole, a melatonin receptor antagonist. *Am J Physiol Regul Integr Comp Physiol.* **280**(5): R1476–82.
- 32 Droin N, Jacquelin A, Guery L *et al.* (2009). Various functions of caspases in hematopoiesis. *Front Biosci.* **14**: 2358–71.
- 33 Falcón J, Besseau L, Fuentès M *et al.* (2009). Structural and functional evolution of the pineal melatonin system in vertebrates. *Ann N Y Acad Sci.* **1163**: 101–11.
- 34 Feng Z, Qin C, Chang Y, Zhang JT (2006). Early melatonin supplementation alleviates oxidative stress in a transgenic mouse model of Alzheimer's disease. *Free Radic Biol Med.* **40**(1): 101–9.
- 35 Finocchiaro LM, Arzt ES, Fernández-Castelo S, Criscuolo M *et al.* (1988). Serotonin and melatonin synthesis in peripheral blood mononuclear cells: stimulation by interferon-gamma as part of an immunomodulatory pathway. *J Interferon Res.* **8**(6): 705–16.
- 36 Formigli L, Ibba Manneschi L, Tani A *et al.* (1997). Vitamin E prevents neutrophil accumulation and attenuates tissue damage in ischemic-reperfused human skeletal muscle. *Histol Histo-pathol.* **12**(3): 663–9.
- 37 Ghielmini M, Pagani O, Pampallona, de Jong J *et al.* (1999). Double-blind randomized study on the myeloprotective effect of melatonin in combination with carboplatin end etoposide in advanced lung cancer. *Br J Cancer.* **80**(7): 1058–61.
- 38 Goda K, Hamane Y, Kishimoto R, Ogishi Y (1999). Radical scavenging properties of tryptophan metabolites. Estimation of their radical reactivity. *Adv Exp Med Biol.* **467**: 397–402.
- 39 Haldar C, Häussler D, Gupta D (1992, a). Effect of the pineal gland on circadian rhythmicity of colony forming units for granulocytes and macrophages (CFU-GM) from rat bone marrow cell cultures. *J Pineal Res.* **12**(2): 79–83.
- 40 Haldar C, Häussler D, Gupta D (1992, b). Response of CFU-GM (colony forming units for granulocytes and macrophages) from intact and pinealectomized rat bone marrow to murine recombinant interleukin-3 (rIL-3), recombinant granulocyte-macrophage colony stimulating factor (rGM-CSF) and human recombinant erythropoietin (rEPO). *Prog Brain Res.* **91**: 323–5.
- 41 Hardeland R (1999). Melatonin and 5-methoxytryptamine in non-metazoans. *Reprod Nutr Dev.* **39**(3): 399–408.
- 42 Herrera J, Nava M, Romero F, Rodríguez-Iturbe B (2001). Melatonin prevents oxidative stress resulting from iron and erythropoietin administration. *Am J Kidney Dis.* **37**(4): 750–7.
- 43 Hirata F, Hayaishi O, Tokuyama T, Seno S (1974). In vitro and in vivo formation of two new metabolites of melatonin. *J Biol Chem.* **249**(4): 1311–3.

- 44 Hörl WH (2002). Adjunctive therapy in anaemia management. *Nephrol Dial Transplant*. **17** Suppl 5: 56–9.
- 45 Hu CL, Liu Z, Gao ZY *et al.* (2005). 2-iodomelatonin prevents apoptosis of cerebellar granule neurons via inhibition of A-type transient outward K<sup>+</sup> currents. *J Pineal Res*. **38**(1): 53–61.
- 46 Huan C, Zhou M, Wu M *et al.* (2001). Activation of melatonin receptor increases a delayed rectifier K<sup>+</sup> current in rat cerebellar granule cells. *Brain Res*. **917**(2): 182–90.
- 47 Ianăș O, Olinescu R, Bădescu I (1991). Melatonin involvement in oxidative processes. *Endocrinologie*. **29**(3–4): 147–53.
- 48 Jaworek J, Nawrot-Porabka K, Leja-Szpak A *et al.* (2007). Melatonin as modulator of pancreatic enzyme secretion and pancreatoprotector. *J Physiol Pharmacol*. **58** Suppl 6: 65–80.
- 49 Jung FJ, Yang L, Härter L *et al.* (2004). Melatonin in vivo prolongs cardiac allograft survival in rats. *J Pineal Res*. **37**(1): 36–41.
- 50 Kang JC, Ahn M, Kim YS *et al.* (2001). Melatonin ameliorates autoimmune encephalomyelitis through suppression of intercellular adhesion molecule-1. *J Vet Sci*. **2**(2): 85–9.
- 51 Kaptanoglu E, Palaoglu S, Demirpençe E *et al.* (2003). Different responsiveness of central nervous system tissues to oxidative conditions and to the antioxidant effects of melatonin. *J Pineal Res*. **34**(1): 32–35.
- 52 Kast RE, Altschuler EL (2006). Co-administration of ramelteon and flvoxamine to increase levels of interleukin-2. *Med Hypotheses*. **67**(6): 1389–90.
- 53 Kedziora-Kornatowska K, Szweczyk-Golec K, Czuczejko J *et al.* (2007). Effect of melatonin on the oxidative stress in erythrocytes of healthy young and elderly subjects. *J Pineal Res*. **42**(2): 153–8.
- 54 Kvetnoy IM (1999). Extrapineal melatonin: location and role within diffuse neuroendocrine system. *Histochem J*. **31**(1): 1–12.
- 55 Koc M, Buyukokuroglu ME, Taysi S (2002). The effect of melatonin on peripheral blood cells during total body irradiation in rats. *Biol Pharm Bull*. **25**(5): 656–7.
- 56 Konturek SJ, Konturek PC, Brzozowska I *et al.* (2007). Localization and biological activities of melatonin in intact and diseased gastrointestinal tract (GIT). *J Physiol Pharmacol*. **58**(3): 381–405.
- 57 Korf HW (1994). The pineal organ as a component of the biological clock. Phylogenetic and ontogenetic considerations. *Ann N Y Acad Sci*. **719**: 13–42.
- 58 Korkmaz A (2008). Melatonin as an adjuvant therapy in patients with rheumatoid arthritis. *Br J Clin Pharmacol*. **66**(2): 316–7.
- 59 Kornblihtt LI, Finocchiaro L, Molinas FC (1993). Inhibitory effect of melatonin on platelet activation induced by collagen and arachidonic acid. *J Pineal Res*. **14**(4): 184–91.
- 60 Labonia W, Rubio D, Arias C (2005). Melatonin corrects reticuloendothelial blockade and iron status in haemodialysed patients. *Nephrology (Carlton)*. **10**(6): 583–7.
- 61 Labunets IF, Butenko GM, Khavinson VKh (2004). Effects of bioactive factors of the pineal gland on thymus function and cell composition of the bone marrow and spleen in mice of different age. *Bull Exp Biol Med*. **137**(5): 510–2.
- 62 Launay JM, Lamaître BJ, Husson HP *et al.* (1982). Melatonin synthesis by rabbit platelets. *Life Sci*. **31**(14): 1487–94.
- 63 Leach CM, Thorburn GD (1980). A comparison of the inhibitory effects of melatonin and indomethacin on platelet aggregation and thromboxane release. *Prostaglandins*. **20**(1): 51–6.
- 64 Leaden PJ, Catalá A (2007). Melatonin and N-acetyl serotonin inhibit selectively enzymatic and non-enzymatic lipid peroxidation of rat liver microsomes. *Prostaglandins Leukot Essent Fatty Acids*. **77**(1): 29–35.
- 65 Lialiaris T, Lyratzopoulos E, Papachristou F *et al.* (2008). Supplementation of melatonin protects human lymphocytes in vitro from the genotoxic activity of melphalan. *Mutagenesis*. **23**(5): 347–54.
- 66 Lissoni P, Bastone A, Sala R *et al.* (1987). The clinical significance of melatonin serum determination in oncological patients and its correlations with GH and PRL blood levels. *Eur J Cancer Clin Oncol*. **23**(7): 949–57.
- 67 Lissoni P, Barni S, Brivio F *et al.* (1995, a). A biological study on the efficacy of low-dose subcutaneous interleukin-2 plus melatonin in the treatment of cancer-related thrombocytopenia. *Oncology*. **52**(5): 360–2.
- 68 Lissoni P, Barni S, Brivio F *et al.* (1995, b). Treatment of cancer-related thrombocytopenia by low-dose subcutaneous interleukin-2 plus the pineal hormone melatonin: a biological phase II study. *J Biol Regul Homeost Agents*. **9**(2): 52–4.
- 69 Lissoni P, Vigorè L, Rescaldani R *et al.* (1995, c). Neuroimmunotherapy with low-dose subcutaneous interleukin-2 plus melatonin in AIDS patients with CD4 cell number below 200/mm<sup>3</sup>: a biological phase-II study. *J Biol Regul Homeost Agents*. **9**(4): 155–8.
- 70 Lissoni P, Tancini G, Barni S *et al.* (1996). The pineal hormone melatonin in hematology and its potential efficacy in the treatment of thrombocytopenia. *Recenti Prog Med*. **87**(12): 582–5.
- 71 Lissoni P (2007). Biochemotherapy with immunomodulating pineal hormones other than melatonin: 5-methoxytryptamine as a new oncostatic pineal agent. *Pathol Biol (Paris)*. **55**(3–4): 198–200.
- 72 Liu F, Ng TB, Fung MC (2001). Pineal indoles stimulate the gene expression of immunomodulating cytokines. *J Neural Transm*. **108**(4): 397–405.
- 73 Lotufo CM, Yamashita CE, Farsky SH, Markus RP (2006). Melatonin effect on endothelial cells reduces vascular permeability induced by leukotriene B<sub>4</sub>. *Eur J Pharmacol*. **534**(1–3): 258–63.
- 74 Lu T, Galijasevic S, Abdulhamid I, Abu-Soud HM (2008). Analysis of the mechanism by which melatonin inhibits human eosinophil peroxidase. *Br J Pharmacol*. **154**(6): 1308–17.
- 75 Maestroni GJ (1995). T-helper-2 lymphocytes as a peripheral target of melatonin. *J Pineal Res*. **18**(2): 84–9.
- 76 Maestroni GJ (1998). Is hematopoiesis under the influence of neural and neuroendocrine mechanisms? *Histol Histopathol*. **13**(1): 271–4.
- 77 Maestroni GJ (1999). MLT and the immune-hematopoietic system. *Adv Exp Med Biol*. **460**: 395–405.
- 78 Majewska M, Zajac K, Zemelka M, Szczepanik M (2007). Influence of melatonin and its precursor L-tryptophan on Th1 dependent contact hypersensitivity. *J Physiol Pharmacol*. **58** Suppl 6: 125–32.
- 79 Markus RP, Ferreira ZS, Fernandes PA, Cecon E (2007). The immune-pineal axis: a shuttle between endocrine and paracrine melatonin sources. *Neuroimmunomodulation*. **14**(3–4): 126–33.
- 80 Martin V, Sainz RM, Antolin I *et al.* (2002). Several antioxidant pathways are involved in astrocyte protection by melatonin. *J Pineal Res*. **33**(4): 204–212.
- 81 Mauri R, Lissoni P, Resentini M *et al.* (1985). Effects of melatonin on PRL secretion during different photoperiods of the day in prepubertal and pubertal healthy subjects. *J Endocrinol Invest*. **8**(4): 337–41.
- 82 Mayo JC, Sainz RM, Uria H *et al.* (1998). Melatonin prevents apoptosis induced by 6-hydroxydopamine in neuronal cells: implications for Parkinson's disease. *J Pineal Res*. **24**(3): 179–92.
- 83 Mayo JC, Sainz RM, Antolin I *et al.* (2002). Melatonin regulation of antioxidant enzyme gene expression. *Cell Mol Life Sci*. **59**(10): 1706–1713.
- 84 Melchiorri D, Ortiz GG, Reiter RJ *et al.* (1998). Melatonin reduces paraquat-induced genotoxicity in mice. *Toxicol Lett*. **95**(2): 103–8.
- 85 Mogulkoc R, Baltaci AK, Aydin L *et al.* (2006). Pinelectomy increases oxidant damage in kidney and testis caused by hyperthyroidism in rats. *Cell Biochem Funct*. **24**(5): 449–53.
- 86 Morera AL, Abreu P (2005). Existence of melatonin in human platelets. *J Pineal Res*. **39**(4): 432–3.
- 87 Naranjo MC, Guerrero JM, Rubio A *et al.* (2007). Melatonin biosynthesis in the thymus of humans and rats. **64**(6): 781–90.
- 88 Paterson PY, Hanson MA (1969). Cyclophosphamide inhibition of experimental allergic encephalomyelitis and cellular transfer of the disease in Lewis rats. *J Immunol*. **103**(6): 1311–6.
- 89 Peña C, Rincon J, Pedreanez A *et al.* (2007). Chemotactic effect of melatonin on leukocytes. *J Pineal Res*. **43**(3): 263–9.

- 90 Persengiev SP, Kyurkchiev S (1993). Selective effect of melatonin on the proliferation of lymphoid cells. *Int J Biochem.* **25**(3): 441–4.
- 91 Pieri C, Marra M, Moroni F *et al.* (1994). Melatonin: a peroxyl radical scavenger more effective than vitamin E. *Life Sci.* **55**(15): PL271–6.
- 92 Poeggeler B, Reiter RJ, Tan DX *et al.* (1993). Melatonin, hydroxyl radical-mediated oxidative damage, and aging: a hypothesis. *J Pineal Res.* **14**(4): 151–68.
- 93 Pozo D, Delgado M, Fernandez-Santos JM *et al.* (1997). Expression of the Mel1a-melatonin receptor mRNA in T and B subsets of lymphocytes from rat thymus and spleen. *Faseb J.* **11**(6): 466–73.
- 94 Raikhlín NT, Kvetnoy IM (1976). Melatonin and enterochromaffin cells. *Acta Histochem.* **55**(1): 19–24.
- 95 Rapozzi V, Zorzet S, Comelli M (1998). Melatonin decreases bone marrow and lymphatic toxicity of adriamycin in mice bearing TLX5 lymphoma. *Life Sci.* **63**(19): 1701–13.
- 96 Regodón S, del Prado Míguez M, Jardín I *et al.* (2009). Melatonin, as an adjuvant-like agent, enhances platelet responsiveness. *J Pineal Res.* **46**(3): 275–85.
- 97 Reiter RJ, Tan DX, Cabrera J, D'Arpa D (1999). Melatonin and tryptophan derivatives as free radical scavengers and antioxidants. *Adv Exp Med Biol.* **467**: 379–87.
- 98 Reiter RJ, Acuña-Castroviejo D, Tan DX, Burkhardt S (2001, a). Free radical-mediated molecular damage. Mechanisms for the protective actions of melatonin in the central nervous system. *Ann NY Acad Sci.* **939**: 200–15.
- 99 Reiter RJ, Tan DX, Burkhardt S, Manchester LC (2001, b). Melatonin in plants. *Nutr Rev.* **59**(9): 286–90.
- 100 Reiter RJ, Tan DX (2003). What constitutes a physiological concentration of melatonin? *J Pineal Res.* **34**(1): 79–80.
- 101 Rosengarten H, Meller E, Friedhoff AJ (1972). In vitro enzymatic formation of melatonin by human erythrocytes. *Res Commun Chem Pathol Pharmacol.* **4**(2): 457–65.
- 102 Rossi MT, Ferrari R, Biral P, Di Bella L, Zini I (1974). Tasso leucocitario e trombopenia. *Boll Soc Ital Biol Sper.* Vol. L, N. 20 bis.
- 103 Rossi MT, Scalera G, Di Bella L (1976). Azione mielotropa della melatonina. *Boll Soc Ital Biol Sper.* Vol. LII, N. 18 bis.
- 104 Rossi MT, Di Bella L (1988). Melatonin in thrombocytogenesis. The pineal gland and cancer, edited by Gupta *et al.*: Brain Research Promotion. 183–94.
- 105 Rubio S, Estévez F, Cabrera J *et al.* (2007). Inhibition of proliferation and induction of apoptosis by melatonin in human myeloid HL-60 cells. *J Pineal Res.* **42**(2): 131–8.
- 106 Sandyk R (1990). Possible role of pineal melatonin in the mechanisms of aging. *Int J Neurosci.* **52**(1–2): 85–92.
- 107 Sandyk R (1993). Multiple sclerosis: the role of puberty and the pineal gland in its pathogenesis. *Int J Neurosci.* **68**(3–4): 209–25.
- 108 Santello FH, Frare EO, dos Santos CD *et al.* (2008). Suppressive action of melatonin on the TH-2 immune response in rats infected with *Trypanosoma cruzi*. *J Pineal Res.* **45**(3): 291–6.
- 109 Sarma A, Rodriguez MA, Cabanillas F *et al.* (2004). Recovery of natural killer cell counts after one course of CHOP chemotherapy is diminished in patients older than 60 compared to patients younger than 60.
- 110 Scott AE, Cosma GN, Frank AA *et al.* (2001). Disruption of mitochondrial respiration by melatonin in MCF-7 cells. *Toxicol Appl Pharmacol.* **171**(3): 149–56.
- 111 Shedpure M, Pati AK (1995). The pineal gland: structural and functional diversity. *Indian J Exp Biol.* **33**(9): 625–40.
- 112 Sutherland ER, Martin RJ, Ellison MC, Kraft M (2002). Immunomodulatory effects of melatonin in asthma. *Am J Respir Crit Care Med.* **166**(8): 1055–61.
- 113 Szczepanik M. (2007). Melatonin and its influence on immune system. *J Physiol Pharmacol.* **58** Suppl 6: 115–24.
- 114 Tan DX, Manchester LC, Reiter RJ *et al.* (1999). Identification of highly elevated levels of melatonin in bone marrow: its origin and significance. *Biochim Biophys Acta.* **1472**(1–2): 206–14.
- 115 Tesoriere L, D'Arpa D, Conti S *et al.* (1999). Melatonin protects human red blood cells from oxidative hemolysis: new insights into the radical-scavenging activity. *J Pineal Res.* **27**(2): 95–105.
- 116 Todisco M (2007). Melatonin makes splenectomy unnecessary in two patients with idiopathic thrombocytopenic purpura refractory to corticosteroids. *J Pineal Res.* **43**(2): 214.
- 117 Trubiani O, Recchioni R, Moroni F *et al.* (2005). Melatonin provokes cell death in human B-lymphoma cells by mitochondrial-dependent apoptotic pathway activation. *J Pineal Res.* **39**(4): 425–31.
- 118 Vacas MI, Del Zar MM, Martinuzzo M, Cardinali DP (1992). Binding sites for [3H]-melatonin in human platelets. *J Pineal Res.* **13**(2): 60–5.
- 119 Varga Z, Panyi G, Péter M Jr *et al.* (2001). Multiple binding sites for melatonin on Kv1. 3. *Biophys J.* **80**(3): 1280–97.
- 120 Vaziri ND, Oveisi F, Reyes GA, Zhou XJ (1996). Dysregulation of melatonin metabolism in chronic renal insufficiency: role of erythropoietin deficiency anemia. *Kidney Int.* **50**(2): 653–6.
- 121 Vijayalaxmi, Meltz ML, Reiter RJ, Herman TS (1999). Melatonin and protection from genetic damage in blood and bone marrow: whole-body irradiation studies in mice. *J Pineal Res.* **27**(4): 221–5.
- 122 Viviani S, Negretti E, Orazi A *et al.* (1990). Preliminary studies on melatonin in the treatment of myelodysplastic syndromes following cancer chemotherapy. *J Pineal Res.* **8**(4): 347–54.
- 123 Wheeler AG, Dansby D, Hawkins HC *et al.* (2004). A toxicologic and hematologic evaluation of cyclophosphamide in experimental animals. *Toxicol Appl Pharmacol.* **4**: 324–43.
- 124 Yau MY, Pang CS, Kravtsov G *et al.* (2002). 2[125I]iodomelatonin binding sites in guinea pig platelets. *J Pineal Res.* **32**(2): 97–105.
- 125 Yerer MB, Aydogan S, Yapislar H *et al.* (2003). Melatonin increases glutathione peroxidase activity and deformability of erythrocytes in septic rats. *J Pineal Res.* **35**(2): 138–9.
- 126 Yu Q, Miller SC, Osmond DG (2000). Melatonin inhibits apoptosis during early B-cell development in mouse bone marrow. *J Pineal Res.* **29**(2): 86–93.
- 127 Zhou MO, Jiao S, Liu Z *et al.* (2003). Luzindole, a melatonin receptor antagonist, inhibits the transient outward K<sup>+</sup> current in rat cerebellar granule cells. *Brain Res.* **970**(1–2): 169–77.