

Phenotypes of *SLC26A4* gene mutations – Pendred syndrome and hypoacusis with enlarged vestibular aqueduct

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Abstract

This paper presents the current views, regarding the pathomechanisms, which lead to the development of pathological symptoms in the enlargement of the vestibular aqueduct syndrome (EVAS) and the Pendred syndrome (PS). Associated phenotypes have been discussed and an attempt has been undertaken to correlate them with a corresponding genotype. Mutations of *SLC26A4* gene are one of the factors, which are at the base of congenital hearing losses. Inherited hearing loss occurs in these cases either as an isolated phenomenon with anatomical anomalies of the labyrinth in the background (EVAS) or with endocrine disorders (PS). The official name of *SLC26A4* gene is “solute carrier family 26, member 4”. Pendrin, the product of its expression, transports iodine beyond thyroid follicular cells, where it is linked with thyroglobulin and, then, used in hormone synthesis. Abnormal expression of *SLC26A4* gene results in disturbance of iodine organification. In the internal ear, pendrin transports bicarbonates to the endolymph, taking in this way an active part in pH control of the endolymph and providing proper functioning of KCNJ10 potassium channels and TRP5 calcium channels. Disorders of homeostasis in labyrinth fluids are responsible for abnormalities of its structure, such as enlargement of the vestibular aqueduct and of the endolymph sac. At present, the Human Gene Mutations database provides 124 recessive mutations of *SLC26A4* gene. In EVAS and PS, two missense mutations are most frequently observed: L236P and T416P, as well as the mutation, regarding abnormal splicing process, i.e., IVS8+1G-A, in a total of 55% of the patients with recognised mutation of *SLC26A4* gene; the remaining 45% of changes of this gene are unique mutations.

INTRODUCTION

The Pendred syndrome (PS, MIM 274600) is an autosomally recessively inherited disease, which is caused by mutation in *SLC26A4* gene (MIM 605646). Its diagnosis requires identification of the classical triad of symptoms, including hypoacusis, thyroid goitre and iodine organification defect in the thyroid, which may lead to hypothyroidism.

The familial occurring, isolated hypoacusis with enlarged vestibular aqueduct, was for the first time described in 1996 (*DFNB4* gene, MIM 600791) [7] and has since then been confirmed in a number of reports. It is also referred to as the enlarged vestibular aqueduct syndrome – EVAS (MIM 603545) or the pseudo-Pendred syndrome [12, 16, 34]. EVAS, in its clinical picture, involves bilateral receiving sensorineural hearing loss, up to deafness, as well as labyrinth structure disorders.

GENETIC BACKGROUND

SLC26A4 gene

Exactly one hundred years after the description of a disease by Vaughan Pendred in 1896 [29], referred to since then by his name as the Pendred syndrome, a region was identified on the long arm of chromosome 7 (7q22.3-q31.1), within an interval of 9-cM (centimorgan), between GATA 23F05 and D7S687 loci, the genetic changes of which were related to occurrence of the syndrome in question [7, 41]. A year later, Everett et al. [9] identified *SLC26A4* (*PDS*, *DFNB4*, MIM 605646) gene, the mutations of which were responsible for PS and EVAS. The official name of this gene is “solute carrier family 26, member 4”. This gene has got 21 exons and an open reading frame of 2343 bp in size. Messenger RNA specific for this gene is built of 4927 bp [39] (Fig. 1).

Product of *SLC26A4* gene expression

The product of *SLC26A4* gene expression is pendrin – a 780 amino acid protein with molecular mass of 85 722 Da [22]. This protein is an integral part of the cellular membrane, and its polypeptide chain is 12 times anchored in the cellular membrane, passing through its structure from one side onto the other. This configuration means the presence of 12 transmembrane domains, linked alternately by intra- and extracellular sections. Pendrin participates in active, sodium-dependent transportation of anions, such as: iodides, chlorides, and bicarbonates [8].

The occurrence of pendrin as a monomeric glycoprotein is found in its highest concentrations in the thyroid follicular cells, especially at their colloid adjoining apical surface [13, 39]. Pendrin transports iodine beyond the cell where it binds with thyroglobulin and is stored in colloid and used for the synthesis of thyroid hormones. *SLC26A4* mutations are associated with loss

of the protein ability to transport iodides and with abnormal pendrin localisation in cellular cytoplasm [37, 43]. It has been found that the degree of pendrin expression in the thyroid is not closely related with the functional activity of the gland [32] (Fig. 2).

Pendrin transports ions also within the ear. The complex function of the internal ear, including sound wave transformations and linear and angular accelerations into nervous impulses, depends on the presence of rest potential in cochlear and vestibular structures and on strictly determined electrolyte conditions in labyrinthine fluids. The *SLC26A4* gene undergoes expression within the labyrinth mainly in the regions, responsible for endolymph production, i.e., in the *stria vascularis*, the external spiral sulcus and the adjacent part of the spiral ligament, in Hensen’s cells, the spiral eminence and in the utricle. Based on immunohistochemical methods, its presence is also identified in Corti’s organ – the internal and external hair (auditory) cells, especially in their covering membranes, in supporting cells and the spiral ganglion of the cochlea [51]. In these structures of the internal ear, pendrin participates in HCO_3^- secretion into endolymph. It has been documented that in case of mutated, inactive pendrin, endolymph is acidified in result of decreased HCO_3^- concentration [47]. The increase of endolymph pH suppresses the functionality of KCNJ10 potassium channels and TRP5 and TRP6 calcium channels [47, 26]. The KCNJ10 channels, which transport K^+ to endolymph, play a significant role in inducing and maintaining cochlear rest potential. The KCNJ10 channels are located in intermediary cells, in the *stria vascularis* [46]. The perception of acoustic impulses requires low Ca^{2+} concentrations in endolymph.

The TRP5 and TRP6 epithelial calcium channels occur in the vestibular system and are responsible for Ca^{2+} reabsorption from endolymph [26]. The loss of internal ear functionality, observed in PS and EVAS, results from endolymph acidification, leading to K^+ concentration increase and fall of the intracochlear potential. It also leads to elevated Ca^{2+} concentrations and to loss of the hair cell sensitivity [46]. Using an animal model with murine cell line, deprived of any functional copy of *SLC26A4* gene, degeneration of the *stria vascularis* was found with hyperpigmentation and disorganisation of marginal cells, with secondary infiltration of macrophages [20] (Fig. 3.).

Regarding the kidneys, pendrin occurs in inclusive cells of the cortical collective tubules, where it takes part in the absorption of chlorides and secretion of bicarbonates [38]. The fact that no renal functional disorders are observed in the clinical picture indicates the existence of some pendrin-independent mechanisms of bicarbonate secretion.

Recently, experimental studies have been evaluating the role of pendrin in the renal process of blood pressure control and of arterial blood pH [45].

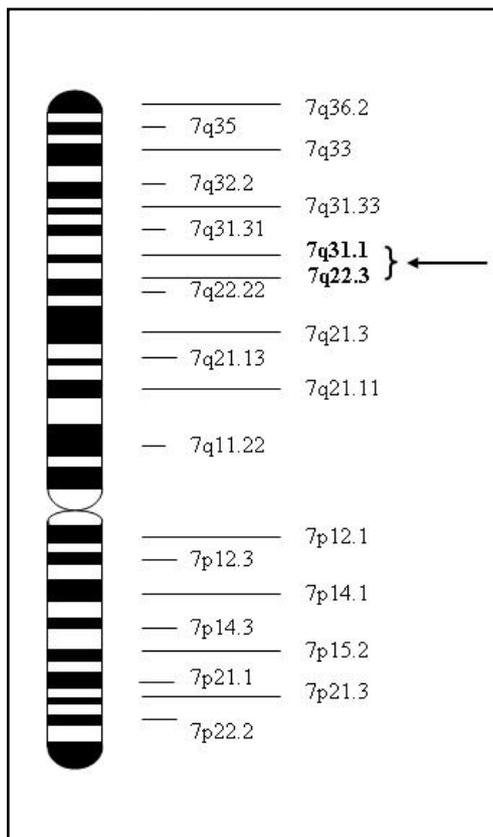


Fig. 1. Location of *SLC26A4* gene in long arm of chromosome 7

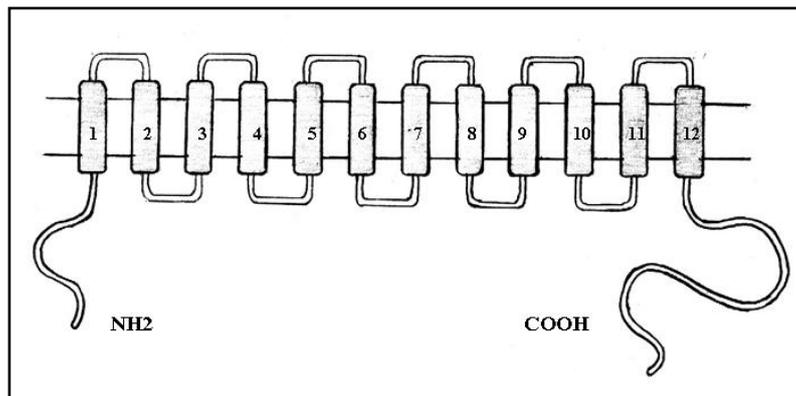


Fig. 2. A model of pendrin molecule

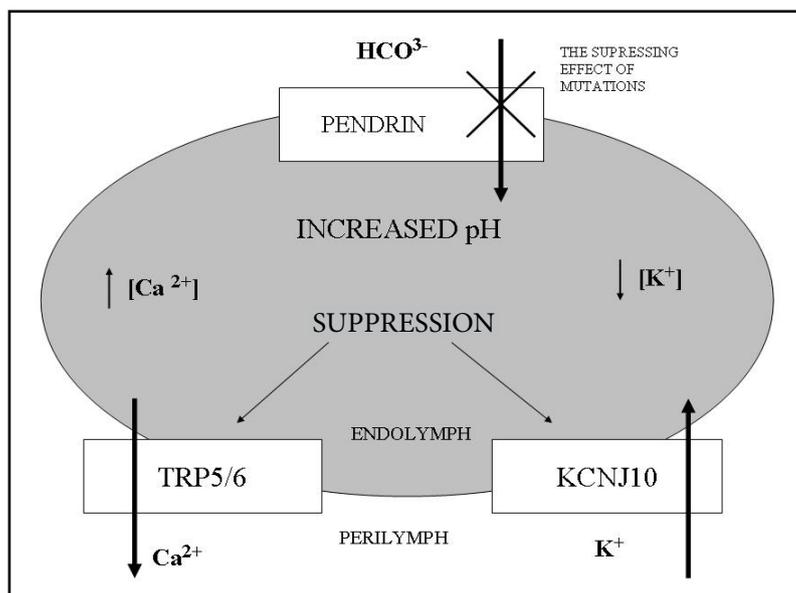


Fig. 3. Disturbances of homeostasis of the internal ear fluids in PS and EVAS

The specificity of pendrin activity in particular organs may result from the existence of its isotypes. It is possible that differences among these isotypes should be looked for beyond the epitope region for the actual antibodies and that is, perhaps, why they have not been identified so far. Another hypothesis, trying to explain it, assumes that pendrin is a component of a multiprotein complex, standing for a tonus-dependent channel for anions. Then, consequently, any differences in the other proteins, included in the structure of this complex, would control the selectivity for particular anions [48].

THE CLINICAL PICTURE

As it has been mentioned before, PS and EVAS are autosomally and recessively inherited diseases. It means that parents, whose child bears the disease, are associated with a 25% risk that another child will also have the same disease. They have also a 50% chance that the next child will be normal, being, however, a carrier of defec-

tive gene, and a 25% chance to have another baby which will not be either ill or carrier of the defective gene.

Disorders of hearing and of the balancing system

Hypoacusis in PS and EVAS occurs at the stage prior to speech formation and enhances in the course of speech development, revealing a progressive character in later stages of life. Sometimes, the degree of hypoacusis undergoes certain fluctuations. In general, hypoacusis is manifested as a sensorineural hearing loss, while only rarely with the transmission component [11]. Certain researchers try to explain the interaural difference in hypoacusis by asymmetry in the degree of labyrinth structure disorders [11, 24]. The cases of hypoacusis and genetic deafness are divided into isolated and concomitant with pathological symptoms in disease syndromes. PS is the most frequent genetic syndrome, causing 5–10% of cases of inherited deafness [11, 28, 35]. There are many genetic causes at the base of inherited isolated hypoacusis, which are difficult, if not totally impossible to be identified from the clinical point of view. Hypoacusis, resulting from disorder of *SLC26A4*

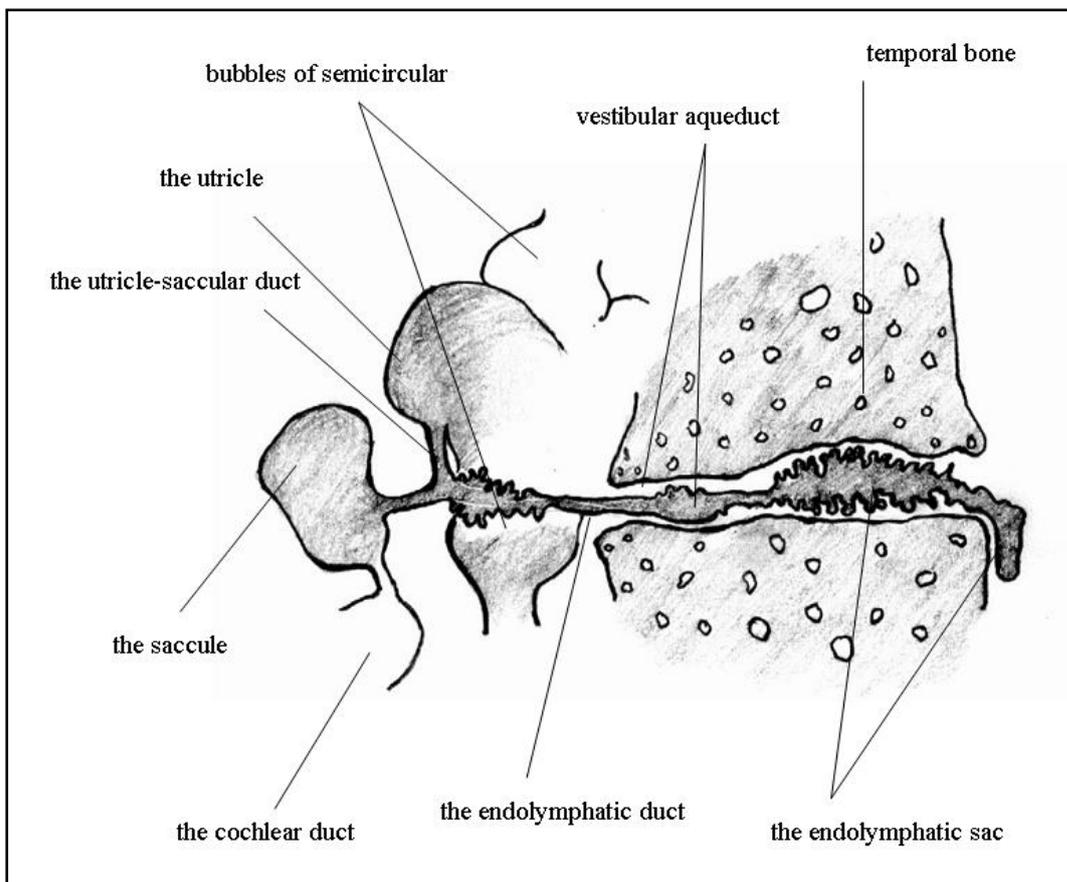


Fig. 4. The endolymphatic duct

gene, is distinguished by the abnormalities of the internal ear structure [6]. Fairly interesting are the results of audiological analysis, combined with radiological evaluation and a 9-year observation of 27 patients with PS and EVAS [5]. Almost the same numbers of persons from each group (about 30%) reported hypoacusis either of steady, progressive character or with stepping increase. Patients with PS had much deeper degree of hypoacusis [5]. No relationship was found between the degree of enlargement of the vestibular aqueduct and the stage of hypoacusis [5]. That observation has also been confirmed by other reports [27]. Dysfunction of the vestibular organ is observed in more than 50% of persons with PS – ranging from light, unilateral channel paresis to bilateral areactivity [35]. It has been found that patients with PS and vertigo more often manifest fluctuating hypoacusis with tinnitus. These subjects, who are homozygotes of *H723R* mutation, experience vertigo episodes much more often than heterozygotes [42].

Endocrine disorders

The defect of iodine organification in the thyroid is, in the clinical practice, identified in a test with potassium perchlorate (PPT). The potassium perchlorate test has been recognised as unreliable because of its low specificity and considerable differences among the criteria

for its interpretation. Abnormal PPT results are also found in patients with Hashimoto's disease, Graves' disease and with thyroid peroxidase (TPO) deficit in the gland, following earlier surgical intervention or after ^{131}I radioiodotherapy, also following long-term rich iodine supplementation in food [48]. Iodine organification leads – only in some cases – to slight suppression of thyroid hormone synthesis. For this reason, the screening examinations of newborns are not always effective to identify potential patients [31].

The goitre occurs in 73–80% of persons with PS, however, it is most often observed already in the second decade of life, and no disorders of thyroid hormone concentrations are usually observed [11, 35]. The goitre occurs despite applied therapy with thyroid hormones [22].

Anatomical anomalies

The vestibular aqueduct is a bone canal (diameter = 0.25 mm), localised in the pyramid of the temporal bone. It begins with an internal foramen in the elliptic recess section of the medial labyrinth wall and turns backwards on a 7–9 mm section, towards the posterior pyramid wall, where it is terminated with a slit-like external foramen. In the vestibular aqueduct, there is one of the five (5) main parts of the membranous labyrinth (beside the utricle, the saccule, the cochlear duct

and the semicircular canals), namely the endolymphatic duct. This duct, leading out of the saccule, joins the utricle-sacculus duct in the vestibular area. At this location, there are folds, separating – to some degree – the cochlear endolymphatic system from the semicircular ducts. Immediately after that juncture, the endolymphatic duct expands twice, forming, so-called – sinuses. Two thirds of the duct is located in the vestibular aqueduct, the terminal part of which extends to contain the third expansion, called the endolymphatic sac. The endolymphatic sac is an expansion, resulting not from inflation and diameter increase but from irregular folding of its inner surface, lined with cuboidal epithelium. The sinuses and the endolymphatic sac participate in the production and resorption of endolymph [23] (Fig. 4).

Enlargement of the vestibular aqueduct is identified when its diameter in the middle of the section between the common limb and the external foramen of the duct amounts to 1.5 mm. Identification of this anomaly is possible by CT or MRI scanning. Enlargement of the vestibular duct, including its content, is the most frequently observed malformation, recognised in the radiological diagnostics of sensorineural hearing loss. This malformation is diagnosed in as many as 12% of children with deafness [1] and in 80–100% with PS [14, 36]. This defect may coexist with incomplete cochlear development, i.e., the lack of apical turns, being referred to in this complex form as Mondini's malformation. Referring to the more and more common use of cochlear implants, CT of the internal ear becomes a routine diagnostic procedure in a growing number of persons with hearing loss, thus resulting in more and more frequent diagnosis of EVAS. Unlike other structural disorders of the internal ear, only EVAS and Mondini's malformation result from *SLC26A4* gene mutations [10]. Other types of labyrinth structure disorders, such as, for example, enlargement of the endolymphatic fossa, hypoplasia of the modiolus or of the horizontal semicircular canal are characteristic for mutations of *GJB2*, encoding connexin-26 protein. These mutations are at the base of more than 50% of cases of autosomally recessive, isolated hypoacusis [2, 33].

CORRELATION OF *SLC26A4* MUTATIONS AND PHENOTYPE

The *SLC26A4* gene is the only, documented so far gene, responsible for PS and EVAS occurrence. At present, the Database of Human Gene Mutations provides 124 recessive mutations of *SLC26A4* gene (Table 1) [19]. These are most often missense (replacement of a single nucleotide by another one, causing stop codon formation – prematurely terminating translations and causing shortened protein chain) and nonsense (replacement of a single nucleotide, causing codon sense modification and, in consequence, incorporation of another, wrong amino acid) mutations. Other types include splicing

Table 1. Type and number of *SLC26A4* mutations, as well as of different phenotypes [19] (modified).

Mutation type	The number of identified mutations
missense & nonsense	77
splicing	21
small deletions	15
small insertions	9
rearrangements	2
Phenotype	The number of cases
PS	68
EVAS	17

mutations small insertions and deletions of gene section and rearrangements (inversions). Because of the dispersed localisation of mutations in the gene, all the 21 exons should undergo sequencing to obtain complete genetic diagnostics [34]. Mutations in *SLC26A4* gene are phenotypically heterogenous, which means the occurrence of various clinical symptoms in carriers of the same mutation. Patients in mutations in *SLC26A4* represent at least two types of phenotypes – PS and, more rarely, EVAS. In either disease syndrome, disorders of the internal structure and hearing loss are observed. The factor, which differentiates PS from EVAS, is rather disturbed iodine organification than the presence of thyroid goitre, which is a not entirely penetrating feature in PS. Differential diagnostics of these diseases is rather difficult, especially in childhood. The search for *SLC26A4* mutation pattern, which would provide or confirm substantial clinical diagnosis, has been attempted for 10 years and is further continued (Tab. 1).

The detectability of *SLC26A4* gene mutations in PS and EVAS amounts to 90% and 70–78%, respectively [3, 44]. Both in persons with enlarged vestibular aqueduct and in those with Mondini's malformation, *SLC26A4* gene mutations are found in very high percents (70–86%) of studied patients [10, 36, 49]. In other reports, mutations could not be found in 1/3 of the cases of enlarged vestibular aqueduct, while in another one third, only a single mutation of *SLC26A4* allele mutation was identified [15]. The close similarity of clinical pictures and the occurrence of mutations in the same gene prompted some researchers to acknowledge PS and EVAS as two variants of the same disease [44]. So far, no correlation has been found between the type of *SLC26A4* mutation and the degree and type of abnormal formation of the labyrinth structures [25]. Genetic studies of patients with PS and EVAS demonstrate differences in the spectrum of *SLC26A4* gene mutations, depending on the race (white or yellow) of the studied group. The most frequent mutations for the white race subjects (L236P, T416P, IVS8+1G-A) are rare for the Japanese, for whom, H723R exceeds 50% of *SLC26A4* mutations. Therefore, the ethnic background should be

taken into account when searching for mutations in this gene [28, 44].

The *SLC26A4* mutations are responsible for 5–10% cases of congenital hearing loss. It has been confirmed in experimental studies that frequent mutations in PS, such as L236P, T416P or E384G cause a complete loss of pendrin dependent transportation of iodides and chlorides, while the mutations, found in EVAS: V480D, V653A, G497S provided maintenance of transportation of these ions, however, at a much lower level. A conclusion has been drawn from the study that this decreased activity of pendrin is satisfactory to eliminate thyroid goitre and disturbances of its functioning from the clinical picture [40].

A significant correlation was found between the phenotype and genotype of *SLC26A4*, while studying a group of 30 persons with enlarged vestibular aqueduct. In the study, the restrictively defined conditions of interpretation of the test with potassium perchlorate were especially emphasised, looking for reasons for the erroneous differentiation between PS and EVAS just in the clinical difficulty of thyroid phenotype assessment. The presence of single *SLC26A4* mutation concerned 61% of patients with EVAS. All the persons fulfilling the PS criteria had two mutated alleles in *SLC26A4* gene [34]. Also patients with PS turned out in other studies to be either homozygotes or compound (double) heterozygotes [4, 10]. Probably, finding of a single heterozygotic mutation in *SLC26A4* gene in patients with PS results from imperfection of the applied diagnostic methods and should encourage to perform further molecular studies [30]. A report is fairly interesting in which the same *T416P* mutation, occurring in three siblings, co-existed with various degrees of hearing loss of different clinical course and accompanied the development of different disorders of the labyrinth structure [27]. It may be explained by the presence of other mutations or by participation of environmental factors.

The failure in finding *SLC26A4* gene mutations in PS and EVAS probably results from limitations of the molecular techniques, applied for their identification, although it may also be an effect of participation of other genes in their aetiology.

FOXI 1 gene

The latest studies of Yang et al. [50] indicate that a gene of the *FOXI 1* (MIM 60 1093) transcriptive factor may participate in PS and EVAS pathogenesis. The *FOXI 1* gene is localised on the long arm of chromosome 5 (5q34). The studies of Larsson et al. [21] have provided evidence that the *FOXI 1* gene undergoes expression during embryonic development within the ectodermal, otic vesicle, which then develops into the membranous labyrinth. This gene is an early controller of the otic vesicle cell differentiation process into structures of the internal ear [21]. Studies on homozygotic mice with mutated *FOXI 1* gene showed the animals to be deaf, revealing dysfunction of the static system and considerable disorders in the internal ear structure [17]. In the

course of later studies, performed on an analogous animal model, a total lack of pendrin, the product of *SLC26A4* gene expression, was found in the epithelial cells of the endolymph duct and sac [18]. It was, in this way, demonstrated that the product of *FOXI 1* gene expression, as a transcriptive factor, controls the efficacy of *SLC26A4* gene transcription process. Yang et al. [50] have performed the first genetic studies, associating *FOXI 1* gene mutations with PS and EVAS in humans. In their research, they have identified a promotor fragment of *SLC26A4* gene, associated with the factor activating its transcription – i.e., with *FOXI 1*. In nine patients with PS and EVAS, they recognised a new mutation – 103 T-C – concerning just the fragment of *SLC26A4* gene. In other six patients, they demonstrated the presence of *FOXI 1* gene mutations, which prevented the activation of *SLC26A4* gene transcription. The members of one of the families with EVAS, included in the authors' studies, had both genes, i.e., *FOXI 1* and *SLC26A4*, mutated. Following the results of their studies, the authors have developed a hypothesis that both *FOXI 1* transcriptive factor gene mutations and mutations of *SLC26A4* gene, subordinate to *FOXI 1*, are involved in the pathogenesis of PS and EVAS, which has provided the first examples of bigene inheritance being the cause of hypoacusis.

CONCLUDING COMMENTS

A complex clinical evaluation: endocrine and audiological, together with radiological diagnostic imaging, supported by molecular studies of *SLC26A4* gene, are the procedures, necessary for complete and accurate diagnosis of PS and EVAS. Routine molecular diagnostics, available in European and American laboratories, involves a screening examination, searching the three most common mutations. When the mutations are not identified, a panel of 41 known mutations is applied and, eventually, sequencing of all the 21 exons of *SLC26A4* gene is possible in order to find any mutations at all. However, as it has already been mentioned, the routine diagnostics of *SLC26A4* gene allows for an identification of mutations in approximately 50% of patients with PS and EVAS. It is associated with limitations of the sequencing method, which does not identify all the defects of the cutting points, regulatory sequences of the gene or changes in introns (it is assumed, at present, that a sequence of introns influences correct functioning of the gene). It is not, however, excluded that changes in the genome, other than *SLC26A4* mutation, may be at the base of the pathologies. Taking into account the considerable phenotypic similarity of both syndromes and the defect of the same gene, being at the base of the syndromes, some researchers suggest considering both entities as variants (with or without endocrine symptoms) of the same disease. The application of molecular studies in the diagnostics of Pendred's syndrome and the syndrome of enlarged vestibular aqueduct provides significant data on the genetic aetiology of these diseases. It has been demonstrated that a

high percent of cases with PS and EVAS (70–90% in research studies, 50% in routine diagnostics) are caused by mutations of both alleles of *SLC26A4* gene, either in homo- or in heterozygotic system. The spectrum of observed mutations is similar for either syndrome, however in EVAS, also missense type mutations are observed, which is not met in PS. In both syndromes, the following two missense mutations: *L236P* and *T416P*, together with the mutation, concerning the abnormal splicing process, i.e., *IVS8+1G-A*, are most frequently found: in a total of 55% of patients with identified mutation of *SLC26A4* gene. The other 45% of changes in the gene include unique mutations, traced in very few families. Because of the phenotype heterogeneity of *SLC26A4* mutations, the result of molecular study does not yet allow for differentiation of the above-mentioned syndromes between each other. Neither can the finding of a single mutated *SLC26A4* allele be in any way decisive for the diagnosis without clinical evaluation towards PS/EVAS. Most probably, additional genetic factors (mutations and polymorphisms of other genes), as well as environmental factors, are responsible for various expressions of the disease phenotype in members of the families with the same defect of *SLC26A4*. The role of molecular studies will grow up, when the gene therapy becomes genotypically specific.

REFERENCES

- Arcand P, Desrosier M, Dube J, Abela A. The large vestibular aqueduct syndrome and sensorineural hearing loss in the pediatric population. *J Otolaryngol.* 1991; **20**: 247–50.
- Birkenhager R, Aschendorff A, Schipper J, Laszig R. Non-syndromic hereditary hearing impairment. *Laryngorhinotologie.* 2007; **86**: 299–313.
- Birkenhager R, Zimmer AJ, Maier W, Klenzner T, Aschendorff A, Schipper J. Evidence of a novel gene for the LAV-syndrome. *Laryngorhinotologie.* 2007; **86**: 102–6.
- Cho MA, Jeong SJ, Eom SM, Park HY, Lee YJ, Park SE, et al. The H323R mutation in the PDS/*SLC26A4* gene is associated with typical Pendred syndrome in Korean patients. *Endocrine.* 2006; **30**: 237–43.
- Colvin IB, Beale T, Harrop-Griffiths K. Long term follow-up of hearing loss in children and young adults with enlarged vestibular aqueducts: relationship to radiologic findings and Pendred syndrome diagnosis. *Laryngoscope.* 2006; **273**: 2–11.
- Courtman I, Mancilla V, Ligny C, Hilbert P, Mansbach AL, Van Maldergem L. Clinical findings and PDS mutations in 15 patients with hearing loss and dilatation of the vestibular aqueduct. *J Laryngol Otol.* 2007; **121**: 312–7.
- Coyle B, Coffey R, Armour JA, Gausden E, Hochberg Z, Grossman A, et al. Pendred syndrome (goitre and sensorineural hearing loss) map to the chromosome 7 in the region containing the nonsyndromic deafness gene *DFNB4*. *Nature Genet.* 1996; **12**: 421–3.
- Dawson PA, Markovich D. Pathogenesis of the human *SLC26A4* transporter. *Curr Med Chem.* 2005; **12**: 385–96.
- Everett LA, Glaser B, Beck JC, Idol JR, Buchs A, Heyman M, et al. Pendred syndrome is caused by mutations in putative sulphate transporter gene (PDS). *Nature Genet.* 1997; **17**: 411–22.
- Fitoz S, Sennaroglu L, Incesulu A, Cengiz Fb, Koc Y, Tekin M. *SLC26A4* mutations are associated with a specific inner ear malformation. *Int J Pediatr Otorhinolaryngol.* 2007; **71**: 479–86.
- Fraser GR. Association of congenital deafness with goitre (Pendred Syndrome) a study of 207 families. *Ann Hum Genet.* 1965; **28**: 201–49.
- Fugazzola L, Cerutti N, Mannavola D, Crino A, Cassio A, Gasparoni P, et al. Differential diagnosis between Pendred and pseudo-Pendred syndromes: clinical, radiological and molecular studies. *Pediatric Res.* 2002; **51**: 479–84.
- Gillam MP, Sidhaye AR, Lee EJ, Rutishauser J, Stephan CW, Kopp P. Functional characterization of pendrin in a polarized cell system. *J Biol Chem.* 2004; **279**: 13004–10.
- Goldfeld M, Glaser B, Nassir E, Gomori JM, Hazani E, Bishara N. CT of the ear in Pendred syndrome. *Radiology.* 2005; **253**: 537–40.
- Gonzales TO, Karamanoglu AO, Ceballos CJ, Vives VI, Ramirez RC, Gomez W, et al. Clinical and molecular analysis of three Mexican families with Pendred syndrome. *Eur. J Endocrinol.* 2001; **144**: 585–9.
- Griffith AJ, Arts A, Downs C, Innis JW, Shepard NT, Sheldon S, et al. Familial large vestibular aqueduct syndrome. *Laryngoscope.* 1996; **106**: 960–5.
- Hulander M, Wurst W, Carlsson P, Enerback S. The winged helix transcription factor Fkh 10 is required for normal development of the inner ear. *Nature Genet.* 1998; **20**: 374–6.
- Hulander M, Kiernan AE, Blomqvist SR, Carlsson P, Samuelsson EJ, Johansson BR, et al. Lack of pendrin expression leads to deafness and expansion of the endolymphatic compartment in inner ears of *FOXI1* null mutant mice. *Development.* 2003; **130**: 2013–25.
- Human Gene Mutation Database at the Institute of Medical Genetics in Cardiff. <http://www.hgmd.cf.ac.uk/ac/gene.php?gene=SLC26A4>, 2007-04-13.
- Jabba SV, Oelke A, Singh R, Maganti RJ, Fleming S, Wall SM, et al. Macrophage invasion contributes to degeneration of stria vascularis in Pendred syndrome mouse model. *BMC Med.* 2006; **22**: 4–7.
- Larsson C, Hellqvist M, Pierrou, S, White I, Enerback S, Carlsson P. Chromosomal localization of six human forkhead genes, *frac-1* (FKHL5), -3 (FKHL7), -4 (FKHL8), -5 (FKHL9), -6 (FKHL10), and -8 (FKHL12). *Genomics.* 1995; **30**: 464–9.
- Li XC, Everett LA, Lalwani AK, Desmukh D, Friedman RB, Green ED, et al. A mutation in PDS causes non-syndromic recessive deafness. *Nature Genet.* 1998; **18**: 215–7.
- Łasiński W. Narząd przedsionkowo-ślimakowy: Anatomia Człowieka [Vestibulocochlear organ: Human anatomy (In Polish)] Vol. V. Eds. Bochenek A, Reichert M. PZWL, Warszawa 1989.
- Madden C, Halsted M, Benton C, Greinwald J, Choo D. Enlarged vestibular aqueduct syndrome in the pediatric population. *Otol Neurotol.* 2003; **24**: 625–32.
- Naganawa S, Koshikawa T, Fukatsu H, Ishigaki T, Sato E, Sugiura M, et al. Enlarged endolymphatic duct and sac syndrome: relationship between MR findings and genotype of mutation in Pendred syndrome gene. *Magn Reson Imaging.* 2004; **22**: 25–30.
- Nakaya K, Harbidge DG, Wangemann P, Schultz BD, Green E, Wall SM, et al. Lack of pendrin *HCO3-* transport elevates vestibular endolymphatic $[Ca^{2+}]$ by inhibition of acid-sensitive TRPV5 and TRPV6 channels. *Am J Physiol Renal Physiol.* 2007; **292**: 1314–21.
- Napiontek U, Borck G, Muller-Forell W, Pfarr N, Bohnert A, Keilmann A, et al. Intrafamilial variability of the deafness and goiter phenotype in Pendred syndrome caused by T416P mutation in the *SLC26A4* gene. *J Clin Endocrinol Metab.* 2004; **89**: 5347–51.
- Park HJ, Shaukat S, Liu XZ, Hahn SH, Naz S, Ghosh M, et al. Origins and frequencies of *SLC26A4* (PDS) mutations in east and south Asians: global implications for epidemiology of deafness. *J Med Gen.* 2003; **40**: 242–8.
- Pendred V. Deaf-mutism and goiter. *Lancet.* 1896; **ii**: 532.
- Pfarr N, Borck G, Turk A, Napiontek U, Keilmann A, Muller-Forell W, et al. Goitrous congenital hypothyroidism and hearing impairment associated with mutations in the TPO and *SLC26A4*/PDS genes. *J Clin Endocrinol Metab.* 2006; **91**: 2678–81.
- Pniewska-Siark B, Jeziorowska A, Bobeff I, Lewiński A. Wrodzona niedoczynność tarczycy, aspekty kliniczne i genetyczne [Congenital hypothyroidism, clinical and genetic aspects (In Polish)]. *Klin Pediatr.* 2001; **9**: 217–24.
- Porra V, Bernier-Valentin F, Trouttet-Masson S. Characterization and semiquantitative analyses of pendrin expressed in normal and tumoral human thyroid tissues. *J Clin Endocrinol Metab.* 2002; **87**: 1700–7.

- 33 Propst EJ, Blaser S, Stockley TL, Harrison RV, Gordon KA, Papsin BC. Temporal bone imaging in GJB2 deafness. *Laryngoscope*. 2006; **116**: 2178–86.
- 34 Pryor SP, Madeo AC, Reynolds JC, Sarlis NJ, Arnos KS, Nance WE, et al. SLC26A4/PDS genotype/phenotype correlation in hearing loss with enlargement of the vestibular aqueduct (EVA): evidence that Pendred syndrome and non-syndromic EVA are distinct clinical and genetic entities. *J Med Genet*. 2005; **42**: 159–65.
- 35 Reardon W, Coffey R, Phelps PD, Luxon LM, Stephens D, Kendall-Taylor P, et al. Pendred syndrome – 100 years of underascertainment? *Q J Med*. 1997; **90**: 443–7.
- 36 Reardon W, Mahoney CF, Trembath R, Jan H, Phelps PD. Enlarged vestibular aqueduct: a radiological marker of Pendred syndrome and mutation of the PDS gene. *Q J Med*. 2000; **93**: 99–104.
- 37 Rotman-Piekielny P, Hirschberg K, Maruvada P, Suzuki K, Royaux IE, Green ED, et al. Retention of pendrin in the endoplasmic reticulum is a major mechanism for Pendred syndrome. *Hum Mol Genet*. 2002; **11**: 2625–33.
- 38 Royaux IE, Wall SM, Karniski LP, Everett LA, Suyuki K, Knepper MA, et al. Pendrin, encoded by the Pendred syndrome gene, resides in the apical region of renal intercalated cells and mediates bicarbonate secretion. *Proc Natl Acad Sci USA*. 2001; **98**: 4221–6.
- 39 Scott DA, Wang R, Kreman TM, Sheffield VC, Smith RJH, Karniski LP. The Pendred syndrome gene encodes a chloride-iodide transport protein. *Nature Genet*. 1999; **21**: 440–3.
- 40 Scott DA, Wang R, Kreman TM, Andrews M, McDonald JM, Bishop JR, et al. Functional differences of the PDS gene product are associated with phenotypic variation in patients with Pendred syndrome and non-syndromic hearing loss (DFNB4). *Hum Mol Genet*. 2000; **9**: 1709–15.
- 41 Sheffield VC, Kraiem Z, Nishimura D, Stone EM, Salameh M, Sadeh O, et al. Pendred syndrome maps to chromosome 7q21-34 and is caused by an intrinsic defect in thyroid iodine organification. *Nature Genet*. 1996; **12**: 424–6.
- 42 Sugiura M, Sato E, Nakashima T, Sugiura J, Furuhashi A, Yoshino T, et al. Long-term follow-up in patients with Pendred syndrome: vestibular, auditory and other phenotypes. *Eur Arch Otorhinolaryngol*. 2005; **262**: 737–43.
- 43 Taylor JP, Metcalfe RA, Watson PF, Weetman AP, Trembath RC. Mutations of the PDS gene, encoding pendrin, are associated with protein mislocalization and loss of iodide efflux: implications for thyroid dysfunction in Pendred syndrome. *J Clin Endocrinol Metab*. 2002; **87**: 1778–84.
- 44 Tsukamoto K, Suzuki H, Harada D, Namba A, Abe S, Usami S. Distribution and frequencies of PDS (SLC26A4) mutations in Pendred syndrome and nonsyndromic hearing loss associated with enlarged vestibular aqueduct: a unique spectrum of mutations in Japanese. *Eur J Hum Genet*. 2003; **11**: 916–22.
- 45 Wall SM. The renal physiology of pendrin (SLC26A4) and its role in hypertension. *Novartis Found Symp*. 2006; **273**: 231–9.
- 46 Wangemann P, Itza EM, Albrecht B, Wu T, Jabba SV, Maganti RJ, et al. Loss of KCNJ10 protein expression abolishes endocochlear potential and causes deafness in Pendred syndrome mouse model. *BMC Med*. 2004; **20**: 12–30.
- 47 Wangemann P, Nakaya K, Wu T, Maganti RJ, Itza EM, Sanneman JD, et al. Loss of cochlear HCO³⁻ secretion causes deafness via endolymphatic acidification and inhibition of Ca²⁺ reabsorption in Pendred syndrome mouse model. *Am J Physiol Renal Physiol*. 2007; **292**: 1345–53.
- 48 Wolff J. What is the role of pendrin? *Thyroid*. 2005; **15**: 346–8.
- 49 Yang J, Tsai C, Hsu H, Shiao JY, Su CC, Li SY. Hearing loss associated with enlarged vestibular aqueduct and Mondini dysplasia is caused by splice-site mutation in the PDS gene. *Hear Res*. 2005; **199**: 22–30.
- 50 Yang J, Vidarsson H, Rodrigo-Blomqvist S, Rosengren SS, Enerbäck S, Smith RJR. Transcriptional control of SLC26A4 is involved in Pendred syndrome and nonsyndromic enlargement of vestibular aqueduct (DFN B4). *Am J Hum Genet*. 2007; **80**: 1055–63.
- 51 Yoshino T, Sato E, Nakashima T, Teranishi M, Yamamoto H, Otake H, et al. Distribution of pendrin in the organ of Corti of mice observed by electron immunomicroscopy. *Eur Arch Otorhinolaryngol*. 2006; **263**: 699–704.