# Influence of radiotherapy on 6-sulphatoxymelatonin levels in the urine of brain cancer patients

## Vijayalaxmi,<sup>1</sup> Michael Selva,<sup>1</sup> Russel J. Reiter,<sup>2</sup> Martin L. Meltz,<sup>1</sup> Thomas J. Prihoda,<sup>3</sup> Jonathan Barnes,<sup>1</sup> Belinda Z. Leal,<sup>1</sup> Rajiv S. Dahiya<sup>1</sup> & Terence S. Herman<sup>1</sup>

1. Department of Radiation Oncology

2. Department of Co	ellular and Structural Biology
3. Department of Pa	athology
The University o	f Texas Health Science Center, San Antonio, TX 78229-7889, USA.
Correspondence to:	Vijayalaxmi, Ph.D., Associate Professor, Department of Radiation Oncology, The University of Texas Health Science Center, 7703 Floyd Curl Drive, San Antonio, TX 78229, USA. TEL: +1 210 567 5576 FAX: +1 210 567 3446 E-mail: vijay@uthscsa.edu
Submitted: Accepted:	February 10, 2000 March 27, 2000
Key words:	radiotherapy; 6-sulphatoxymelatonin; brain cancer;

Neuroendocrinology Letters 2000; 21:203–207 pii: NEL210300A03 Copyright © Neuroendocrinology Letters 2000

breast cancer; lung cancer

Abstract OBJECTIVES: The synthesis of melatonin, an endogenous compound synthesized by the pineal gland in the brain, is reported to be depressed in patients with primary cancers of the breast, prostate, stomach and rectum. It is not known whether patients with brain cancer exhibit altered melatonin synthesis. Also unknown is whether radiotherapy given to the region of the brain where the pineal gland is located affects the synthesis of melatonin. This information could be relevant to the clinician for the successful treatment of brain cancer patients since melatonin has been reported to be a potent oncostatic agent.

> METHODS: Urinary levels of 6-sulphatoxymelatonin, the chief metabolite of melatonin, are routinely used as an index of pineal melatonin production and secretion. In this study, the concentrations of 6-sulphatoxymelatonin (aMT6S) excreted in the urine before and during radiotherapy of patients with primary or metastatic brain cancer were determined and compared with the values obtained in breast or lung cancer patients who also received radiotherapy (excluding exposure of the brain where the pineal gland is located).

> RESULTS: The results showed a wide variation in the mean concentration of aMT6S excreted in the urine.

> CONCLUSION: The data from this preliminary study suggested that radiotherapy given to the region of human brain, where the pineal gland is located, does not significantly affect the excretion of aMT6S, the chief metabolite of melatonin.

#### Abbreviations used:

aMT6S - 6-sulphatoxymelatonin

# Introduction

Melatonin (5-methoxy-N-acetyltryptamine) is a highly conserved molecule, existing in organisms as diverse as algae and humans [1 4]. In mammals including man, it is synthesized and secreted by the pineal gland in the brain. It exhibits a distinct circadian rhythm with maximum production occurring during the night [2, 3, 5, 6]. Melatonin has been reported to participate in the regulation of a number of important physiological processes in mammals. It functions as a soporific [7] and as a timing signal [8]. Recent investigations have reported melatonin to be an antioxidant [9 12], capable of scavenging reactive species such as hydroxyl and peroxyl radicals and the peroxynitrite anion [13 18]. The radical scavenging activity of melatonin is hypothesized to be involved in its ability to reduce ionizing radiationinduced genetic damage in peripheral blood lymphocytes collected from healthy human volunteers [19 24]. The radioprotective ability of melatonin was also demonstrated in whole-body irradiated mice, where pre-treatment with melatonin increased survival after exposure to an  $LD_{50/30}$  radiation dose. Melatonin pre-treatment also decreased the extent of radiation-induced genetic damage in both peripheral blood and bone marrow cells of whole-body irradiated mice [25, 26].

The synthesis of melatonin decreases with increasing age of the individual [27, 28] and after exposure to bright light [29]. It has also been reported in some cases to decrease as a result of exposure to extremely low frequency electromagnetic fields [30, 31], intake of alcohol [32, 33], caffeine [34] and certain drugs, such as dexamethasone [35],  $\beta$ -blockers [36] and calcium antagonists [37]. Melatonin production has also been reported to be altered in individuals who experience headaches [38, 39] and in those who suffer from seasonal affective disorder [40]. It is not known if radiotherapy given to the region of the brain where the pineal gland is located affects the synthesis and/or degradation of melatonin. This information could be relevant to the clinician for the successful treatment of primary or metastatic brain cancer patients; melatonin has been shown to be a potent oncostatic agent and to inhibit the growth of experimental tumors in vivo [41], including those of the breast [42], liver [43] and prostate [44].

Urinary levels of 6-sulphatoxymelatonin are routinely used as an index of pineal melatonin production and secretion [8]. The current study was designed to investigate the concentrations of 6-sulphatoxymelatonin (aMT6S), the chief enzymatic metabolite of melatonin, excreted in urine before and during radiotherapy of patients with primary or metastatic brain cancer. The data are compared with the values obtained in breast or lung cancer patients who also received radiotherapy (excluding exposure of the brain where the pineal gland is located).

## Materials and methods

A protocol approved by the Institutional Review Board of the University of Texas Health Science Center at San Antonio was followed. Informed consent was obtained from patients with primary or metastatic brain cancer who received radiotherapy to the region of pineal gland at the Cancer Therapy and Research Center. Informed consent was also obtained from patients receiving radiotherapy for breast and lung cancers: these patients served as controls. Patients who were taking  $\beta$ - and calcium channel blockers or those who were unable to collect or unreliable to collect 24-hour urine samples were excluded from the study. The radiotherapy was given in fractionated doses, daily Monday through Friday. Each participating patient was asked to collect the first 24-hour urine sample before the radiation exposures began and a second sample one week after the start of radiotherapy, and weekly thereafter until the completion of the course of radiotherapy. A 24-hour urine sample was also collected from some patients at one month after completion of the therapy regimen.

All urine samples were stored at  $20+1^{\circ}C$  and coded before determining the concentration of 6-sulphatoxymelatonin (aMT6S) using a competitive binding radioimmunoassay [45, 46]. Briefly, an aliquot of each urine sample (containing aMT6S) was diluted, mixed with a specific antiserum to aMT6S raised in a sheep, and incubated for 30 minutes at room temperature. A trace amount of <sup>125</sup>I-aMT6S (Stockgrand Ltd., Guilford, UK) was then added to this mixture, which was incubated overnight at  $4\pm1^{\circ}$ C. The free and antibody-bound fractions of aMT6S were separated: the free aMT6S fraction was precipitated with dextran-coated charcoal suspension by incubation for 15 minutes at  $4\pm1^{\circ}$ C, and the radioactivity counted in a gamma counter. A standard curve using known amounts of aMT6S was constructed at the same time. The concentrations of aMT6S in the urine samples were calculated from this standard curve. The aMT6S binding was found to be 76% with a specificity of 2 pg/ml and a curve correlation of 0.9978. The values were expressed as ng of aMT6S per mg of creatinine.

**Statistical Analysis**: The data were collected from 9 brain, 7 breast and 2 lung cancer patients and compared between the three types of cancers investigated. The data were also analyzed for the regression of aMT6S excreted in the urine with the radiation dose received by individual cancer patients over their treatment period. These measures were also correlated adjusting for group and patient variability. In addition, based on all available data, the regression estimates of the amount of aMT6S excreted in the urine for each group of patients at 3000, 4000 and 5000 cGy radiation dose were computed.

#### **Results and discussion**

The results are presented in Table 1. Before, during and after radiotherapy, the mean concentration of aMT6S excreted in the urine was not significantly different between the three cancer patient groups (p=0.72) and showed wide variation. The regression slopes for aMT6S excreted in the urine with increasing radiation dose received by the three cancer patient groups were not significantly different (p=0.68). However, brain cancer patient #3 and #6 indicated a linear prediction of aMT6S excretion with

**Table 1.** Concentration of 6-Sulphatoxymelatonin (ng of aMT6S/mg of creatinine) in urine samples of brain cancer patients receiving radiotherapy (cumulative radiation dose in cGy) in the region of pineal gland.

		Patients									(*)	p value
1	cGy aMT6S	0 15.3	900 5.5	1800 2.8	2700 14.4	3600 10.6	4500 16.3	5040 9.0	5260 2.4		0.0	0.8
2	cGy aMT6S	180 7.4	900 8.6	1800 7.6	2700 11.2	3600 5.3	4320 7.7	5040 9.0	6300 10.7		0.1	0.5
3	cGy aMT6S	0 20.3	1260 25.9	1980 58.1	3060 28.9	3420 28.2	4380 37.8				0.8	0.0
4	cGy aMT6S	0 10.0	800 10.2	2600 6.5	3400 10.7	4000 6.8					0.2	0.6
5	cGy aMT6S	0 2.2	1000 11.0	2000 2.6	2800 3.5	4000 8.7					0.1	0.7
6	cGy aMT6S	400 10.2	1200 15.3	2200 17.2	3200 21.2						1.0	0.0
7	cGy aMT6S	0 9.5	1000 13.8	2600 21.2	3600 10.3						0.1	0.7
8	cGy aMT6S	0 3.5	2200 8.7	3400 2.0	4000 13.2						0.2	0.5
9	cGy aMT6s	0 10.7	1000 7.6	2800 12.6	3000 5.7						0.0	0.8
Br	east Cance	r Patients	;									
1	cGy aMT6S	0 13.7	720 19.0	1620 16.5	2520 12.0	3240 20.7	4140 16.4	5100 24.4	5900 19.3	** 29.6	0.3	0.2
2	cGy aMT6S	0	720 8.5	1800 10.1	2880 11.4	3600 9.8	4500 8.5	5400 7.3	6120 8.1	** 9.2	0.0	0.9
		7.1						5220	6120		0.2	0.0
3	cGy aMT6S	0 3.3	720	1620 4.8	2520 4.0	3420 5.4	4320 5.4	7.2	4.4		0.2	0.2
3	cGy	0	720			3420 5.4 4000 11.4				** 9.7	0.2	1.0
4	cGy aMT6S cGy	0 3.3 0	720 5.4 1000	4.8	4.0	5.4 4000	5.4	7.2				
_	cGy aMT6S cGy aMT6S cGy	0 3.3 0 9.2 0	720 5.4 1000 8.6 1200	4.8 2000 10.1 1600	4.0 3000 10.3 2800	5.4 4000 11.4 4000	5.4 5000 10.5 5600	7.2		9.7 **	0.0	1.0
4	cGy aMT6S cGy aMT6S cGy aMT6S cGy	0 3.3 0 9.2 0 1.4 0	720 5.4 1000 8.6 1200 7.8 1000	4.8 2000 10.1 1600 1.6 2000	4.0 3000 10.3 2800 5.4 2800	5.4 4000 11.4 4000 15.8 3800	5.4 5000 10.5 5600 10.9 4800	7.2 6000 7.5 6000		9.7 **	0.0	1.0 0.1
4 5 6 7	cGy aMT6S cGy aMT6S cGy aMT6S cGy aMT6S cGy	0 3.3 0 9.2 0 1.4 0 2.9 0 12.3	720 5.4 1000 8.6 1200 7.8 1000 6.9 800	4.8 2000 10.1 1600 1.6 2000 3.0 1800	4.0 3000 10.3 2800 5.4 2800 5.0 2600	5.4 4000 11.4 4000 15.8 3800 10.3 3600	5.4 5000 10.5 5600 10.9 4800	7.2 6000 7.5 6000		9.7 **	0.0 0.6 0.5	1.0 0.1 0.1
4 5 6 7	CGy aMT6S CGy aMT6S CGy aMT6S CGy aMT6S CGy aMT6S	0 3.3 0 9.2 0 1.4 0 2.9 0 12.3	720 5.4 1000 8.6 1200 7.8 1000 6.9 800	4.8 2000 10.1 1600 1.6 2000 3.0 1800	4.0 3000 10.3 2800 5.4 2800 5.0 2600	5.4 4000 11.4 4000 15.8 3800 10.3 3600	5.4 5000 10.5 5600 10.9 4800	7.2 6000 7.5 6000		9.7 **	0.0 0.6 0.5	1.0 0.1 0.1

the cumulative radiation dose received, with p-values of 0.03 and 0.02, respectively. When computed for the best fit for exposure to similar radiation doses of 3000, 4000 and 5000 cGy, the data indicated that the concentration of aMT6S excreted in the urine of brain cancer patients (12.7, 13.7 and 14.7, respectively) were higher than those observed in breast (9.6, 10.1 and 10.7, respectively) and lung cancer patients (10.4, 10.4 and 10.4, respectively). When the data were all pooled and analyzed by adjusting the two sources of variability (cumulative radiation dose and the type of cancer), the partial correlation of aMT6S and radiation dose was found to be 0.32 (p=0.003). Within groups, this partial correlation was 0.25 (p=0.12), 0.41 (p=0.007) and 0.37 (p=0.32) for brain, breast and lung cancer patients, respectively. Perhaps, the larger correlation observed in breast cancer patients (p=0.007) could be due to receiving higher doses of radiation. For some patients, a curvilinear fit was better compared to the linear fit, but several patients had only 4 urine collection points and a full analysis of this more complicated model was not possible.

A careful examination of the records of patients who participated in this study indicated intake of a variety of drugs (other than those which are shown to have an influence on melatonin synthesis, as mentioned in the introduction). These included ambien, azmacort, dilantin, diazepam, tagamet, tylenol and zocor. Four brain cancer patients (#1, 2, 8 and 9) and one lung cancer patient (#1) were taking dilantin in various dose regimens. The possible effect of this drug on the mean aMT6S level excreted in the urine was evaluated using the analysis of variance test. The radiation exposure was used as a covariate and the drug was tested as a factor and for interaction with the type of cancer. The drug was found to have no significant effect on the mean aMT6S level excreted in the urine.

The data from this preliminary study suggests that radiotherapy given to the region of the human brain where the pineal gland is located does not significantly affect the excretion of the chief melatonin metabolite during treatment. The evidence for a linear relationship between aMT6S excretion and cumulative radiation dose in two brain cancer patients (#3 and 6) could be due to an enhanced synthesis of melatonin or due to an increase in the metabolic degradation of melatonin (resulting in greater amounts of aMT6S excretion in the urine). The urinary excretion level of aMT6S was the highest recorded for patient #3 (58.1 ng/mg creatinine) and the patient was observed to have difficulty to stay awake during weekly checkups: based on this one patient, it is not possible to conclude the increased excretion of aMT6S is due to increased synthesis of melatonin, which in turn is the result of the radiotherapy given to the region of the brain where the pineal gland is located. Investigations with larger numbers of cancer patients and long-term follow-ups are necessary to determine possible differences in melatonin synthesis and degradation.

#### REFERENCES

- 1 Reiter RJ. The pineal gland and its hormones in the control of reproduction in mammals. Endocrine Rev 1980; **1**:109–131.
- 2 Reiter RJ. Melatonin: The chemical expression of darkness. Mol Cell Endocrinol 1991; **79**:c153–c159.
- 3 Reiter RJ. Pineal melatonin: Cell biology of its synthesis and of its physiological interactions. Endocrine Rev 1991; **12**:151–180.
- 4 Poeggler B. Melatonin and the light-dark zeitgeber in vertebrates, invertebrates and unicellular organisms. Experientia 1993; **49**:611–613.
- 5 Ebadi M. Regulation of the synthesis of melatonin and its significance to neuroendocrinology. In: The Pineal Gland (Ed. Reiter, RJ.). New York: Raven Press; 1984. p 1–38.
- 6 Waldhauser F, Dietzel M. Daily and annual rhythms in human melatonin secretion: Role in puberty control. Ann NY Acad Sci 1985; **453**:205–214.
- 7 Ian JE, Espezel H, Gaulden KJ. Melatonin in sleep disorders in children with developmental disabilities. In: "Melatonin in psychiatric and neoplastic disorders" (Eds. Shaffi M and Shaffi SL). Washington: American Psychiatric Press; 1998. p 169–190.
- 8 Arendt J. Melatonin and the mammalian pineal gland. London: Chapman and Hall; 1995.
- 9 Reiter RJ, Melchiorri D, Sewerynek E, Poeggler B, Barlow-Walden LR, Chang JI, et al. A review of the evidence supporting melatonin's role as antioxidant. J Pineal Res 1995; **18**:1–11.
- 10 Benot S, Goberna R, Reiter RJ, Garcia-Maurino S, Osuna C, Guerrero JM. Physiological levels of melatonin contribute to the antioxidant capacity of human serum. J Pineal Res 1999; 27:59–64.
- 11 Tesoriere L, D'Arpa D, Conti S, Giaccone V, Pintaudi AM, Livrea MA. Melatonin protects human red cells from oxidative hemolysis: new insights into radical-scavenging activity. J Pineal Res 1999; 27:95–105.
- 12 Reiter RJ. Oxidative damage to nuclear DNA: amelioration by melatonin. Neuroendocrine Lett 1999; **20**:145–150.
- 13 Tan DX, Chen LD, Poeggeler B, Manchester LC, Reiter RJ. Melatonin: a potent, endogenous hydroxyl radical scavenger. Endocr J 1993; **1**:57–60.
- 14 Tan DX, Manchester LC, Reiter RJ, Plummer BF, Hardies LJ, Weintraub ST, et al. A novel melatonin metabolite, cyclic 3-hydroxymelatonin: A biomarker of in vivo hydroxyl radical generation. Biochem Biophys Res Commun 1998; **253**:614–620.
- 15 Pieri C, Marra M, Moroni F, Recchioni R, Marcheselli F. Melatonin: a peroxyl radical scavenger more effective than vitamin E. Life Sci 1994; **55**:PL271–276.
- 16 Gilad E, Cuzzocrea S, Zingarelli B, Salzman AL, Szaba C. Melatonin is a scavenger of peroxynitrite. Life Sci 1997; **60**:PL169–174.
- 17 Reiter RJ. Antioxidant actions of melatonin. Adv Pharmacol 1997; **38**:103–117.

- 18 Cuzzocrea S, Zingarelli B, Costintino G, Caputi AP. Protective effect of melatonin in non-septic shock model induced by zymosan in the rat. J Pineal Res 1998; **25**:24–33.
- 19 Vijayalaxmi, Reiter RJ, Meltz ML. Melatonin protects human blood lymphocytes from radiation-induced chromosome damage. Mutat Res 1995; **346**:23–31.
- 20 Vijayalaxmi, Reiter RJ, Sewerynek E, Poeggeler B, Leal BZ, Meltz ML. Marked reduction of radiation-induced micronuclei in human blood lymphocytes pretreated with melatonin. Radiat Res 1995; **143**:102–106.
- 21 Vijayalaxmi, Reiter RJ, Leal BZ, Meltz ML. Effect of Melatonin on Mitotic and Proliferation Indices, and Sister Chromatic Exchange in Human Blood Lmphocytes. Mutat Res 1996; **351**:187–192.
- 22 Vijayalaxmi, Reiter RJ, Herman TS, Meltz ML. Melatonin and radioprotection from genetic damage: In vivo/In vitro studies with human volunteers. Mutat Res 1996; **371**:221–228.
- 23 Vijayalaxmi, Reiter RJ, Herman TS, Meltz ML: Melatonin reduces gamma radiation-induced primary DNA damage in human blood lymphocytes. Mutat Res 1998; **397**:203–208.
- 24 Vijayalaxmi, Reiter RJ, Meltz ML, Herman TS. Melatonin: Possible mechanisms involved in its "Radioprotective Effect." Mutat Res 1998; **404**:187–189.
- 25 Vijayalaxmi, Meltz ML, Reiter RJ, Herman TS, Sree Kumar K. Melatonin and protection from whole-body irradiation: Survival studies in mice. Mutat Res 1999; **425**:21–27.
- 26 Vijayalaxmi, Meltz ML, Reiter RJ, Herman TS. Melatonin and protection from genetic damage in blood and bone marrow: Whole-body irradiation studies in mice. J Pineal Res 1999; 27:221–225.
- 27 Sack RL, Lewy AJ, Singer CM. Human melatonin production decreases with age. J Pineal Res 1986; **3**:379–388.
- 28 Reiter RJ. The ageing pineal gland and its physiological consequences. BioEssays 1992; **14**:169–175.
- 29 Lewy AJ, Wehr TA, Goodwin FK. Light suppresses melatonin secretion in humans. Science 1980; **210**:1267–1269.
- 30 Wilson BW, Stevens RG, Richardson LE. Neuroendocrine mediated effects of electromagnetic-field exposure: Possible role of the pineal gland. Life Sci 1989; **45**:1319–1332.
- 31 Reiter RJ. Melatonin suppression by static and extremely low frequency electromagnetic fields: relationship to the reported increased incidence of cancer. Rev Env Health 1994; **10**:171–186.
- 32 Ekman AC, Leppaluoto J, Vakkuri O. Ethanol inhibits melatonin secretion in healthy volunteers in a dose-dependent randomized double blind cross-over study. J Clin Endocrinol Metab 1993; **77**:780–783.
- 33 Badia P, Murphy PJ, Wright KP. Alcohol ingestion and nighttime melatonin levels. Sleep Res 1994; **23**:477–000.
- 34 Wright KP, Badia P, Myers BL, Hakel M. Effects of caffeine, bright light, and their combination on nighttime melatonin and temperature during two nights of sleep deprivation. Sleep Res 1995; **24**:458–000.
- 35 Demisch L, Demisch K, Nickelsen T. Influence of dexamethasone on nocturnal melatonin production in healthy adult subjects. J Pineal Res 1988; **5**:317–322.
- 36 Brismar K, Hylander B, Wetterberg L. Melatonin secretion related to side-effects of  $\beta$ -blockers from the central nervous system. Acta Med Scand 1988; **223**:525–530.
- 37 Meyer AC, Nieuwenhuis JJ, Meyer BJ. Dihydropyridine calcium antagonists depress the amplitude of the plasma melatonin cycle in baboons. Life Sci 1986; **39**:1563–1569.
- 38 Waldenlind E, Ekborn K, Wetterberg L, Fanciullaci M, Marabini

S, Cuteri FS, et al. Lowered circannual urinary melatonin concentrations in episodic cluster headache. Cephalagia 1994; **14**:199–204.

- 39 Brun J, Claustrat B, Saddier P, Chazot G. Nocturnal melatonin excretion is decreased in patients with migraine without aura attacks associated with menses. Cephalagia 1995; 15:136–139.
- 40 McIntyre IM, Norman TR, Burrows GD. Melatonin supersensitivity to dim light in seasonal affective disorder. Lancet 1990; **335**:488–000.
- 41 Bartsch C, Bartsch H, Jain AK, Laumas KR, Wetterberg L. Urinary melatonin levels in human breast cancer patients. J Neural Transm 1981b; **52**:281–294.
- 42 Tamarkin L, Cohen M, Roselle D, Reichert C, Lippman M, Chabner B. Melatonin inhibition and pinealectomy enhancement of 7,12-dimethylbenz(a)anthracene-induced mammary tumors in the rat. Cancer Res 1981; **41**:4432–4436.
- 43 Blask DE, Sauer LA, Dauchy R, Holowachuk EW, Ruhoff MS. New actions of melatonin on tumor metabolism and growth. Biol Sig and Rep 1999; **8**:49–55.
- 44 Philo R, Berkowitz AS. Inhibition of Dunning tumor growth by melatonin. J Urol 1988; **139**:1099–1102.
- 45 Arendt J, Bojkowski C, Franey C, Wright J, Marks V. Immunoassay of 6-hydroxymelatonin sulphate in human plasma and urine: abolition of the 24-hour rhythm with atenolol. J Clin Endocrinol Metab 1985; **60**:1166–1173.
- 46 Aldhous ME, Arendt J. Radioimmunoassay for 6-hydroxymelatonin in urine using an iodinated tracer. Ann Clin Biochem 1988; **25**:298–303.