

Streptococcus group B serotype distribution in anovaginal isolates of women in term pregnancy

Małgorzata ROMANIK^{1,2}, Krzysztof NOWOSIELSKI², Ryszard PORĘBA²,
Urszula SIOMA-MARKOWSKA³, Gayane MARTIROISIAN¹, Jan GROBORZ¹

¹ Department of Medical Microbiology, Medical University of Silesia, Katowice, Poland

² Department of Gynecology and Obstetrics, Specialist Teaching Hospital in Tychy, Poland

³ Department of Women's Health, Faculty of Health Sciences Medical University of Silesia, Katowice, Poland

Correspondence to: Krzysztof Nowosielski, MD., PhD.
Department of Gynecology and Obstetrics, Specialist Teaching Hospital in Tychy,
Edukacji 102, 43-100 Tychy, Poland.
E-MAIL: krzysnowosilcow@yahoo.com

Submitted: 2014-06-05 *Accepted:* 2014-07-02 *Published online:* 2014-07-15

Key words: **group B Streptococcus serotyping; pregnancy; prophylaxis**

Neuroendocrinol Lett 2014; **35**(4):301–305 PMID: 25038604 NEL350414A05 © 2014 Neuroendocrinology Letters • www.nel.edu

Abstract

BACKGROUND: To evaluate Streptococcus group B (GBS) serotype distribution in anovaginal isolates of women in term pregnancy and to assess the correlation of the distribution with socio-epidemiological variables and neonatal outcomes.

DESIGN: An observational study.

SETTINGS: Department of Gynecology and Obstetrics, Specialist Teaching Hospital in Tychy, Poland.

POPULATION: 80 women between 37 and 40 gestation weeks with preserved fetal membranes and who had not been treated with antibiotics for at least two weeks before the study.

MATERIAL AND METHODS: The specimens from the vagina and the rectum of pregnant women were collected. GBS colonization tests were conducted in compliance with Centers for Disease Control and Prevention recommendations. Serotyping of the isolates was performed using the Essum GBS Serotyping Kit (Umea, Sweden) according to manufacturer's instruction. Main outcome measures. GBS serotype distribution in the population of Polish women in term pregnancy.

RESULTS: In the studied group of 80 pregnant women GBS colonization rate was 28.7%. Four GBS serotypes were observed (Ia, V, III and II). Serotype Ia was the most predominant – 43.47%. For GBS Ia, V and III serotypes, no significant difference in the prevalence of diabetes mellitus and neonatal outcomes was observed. Only in one case early-onset sepsis was diagnosed in the neonate and serotype Ia was determined.

CONCLUSIONS: 1) From among four identified GBS serotypes in the population of Polish pregnant women, serotype Ia was the most dominant. 2) For GBS serotypes, no significant difference in the prevalence of diabetes mellitus and neonatal outcomes was observed. 3) Active immunization aimed for preventing GBS colonization in mothers should include not only serotypes V, II and III but also Ia in order to be an effective and safe in preventing life threatening neonatal infections.

INTRODUCTION

Streptococci group B (GBS), colonizing the anogenital region of pregnant women, are frequently involved in the neonatal (51% early-onset sepsis – EOS and 49% late-onset invasive disease – LOD) and maternal (post-partum sepsis, chorioamnionitis, endometritis, urinary tract infections) infections worldwide (Gray *et al.* 2007; Lin *et al.* 2011; Stoll *et al.* 2011; Romanik *et al.* 2011; Kaambwa *et al.* 2010; Häkansson & Källén 2006; Lin *et al.* 1998). The intrapartum maternal chemoprophylaxis of colonized gravidas with β -lactams is currently recommended by Centers for Disease Control and Prevention (CDC) (MMWR 2010). However, common use of antibiotics in prophylaxis in pregnant women may lead to the following complications:

- development of GBS strains resistant to antibiotics administered in intrapartum maternal prophylaxis,
- growth of other bacterial species resistant to antibiotics,
- increase the contribution rate of other bacteria species in neonatal infections,
- development of the new microorganisms that are pathogenic for the newborns and neonates (Moore *et al.* 2003).

The intrapartum antibiotic therapy of GBS recommended by CDC may not prevent the late complications of GBS infections in neonates or may be the cause of allergic reactions in mothers; for that reasons in might not be the proper method of prophylaxis. The use of active immunization for preventing the GBS colonization of gravidas may be, in the nearest future, effective and safe way of life-threatening neonatal infection prophylaxis (Shen *et al.* 2000; Melin 2011). However, the knowledge of GBS serotypes distribution in pregnant women is crucial to prepare such vaccines.

The aim of this study was to evaluate the GBS serotype distribution and to assess the correlation of that distribution with socio-epidemiological variables and neonatal outcomes.

MATERIAL AND METHODS

One hundred and sixty specimens from the vagina and the rectum of 80 pregnant women between 37 and 40 gestation weeks hospitalized in the Maternity Ward of the Department of Gynecology and Obstetrics, Medical University of Silesia in Tychy, Poland, were examined. All study samples (two from each patient) were collected by physicians and transported to the laboratory within one hour. All tested gravidas had preserved fetal membranes and had not been treated with antibiotics for at least two weeks before the test (the complete characterization of studied group was presented in our previous study) (Romanik *et al.* 2011).

GBS colonization tests were conducted in compliance with CDC recommendations, using liquid Todd Hewitt Broth (bioMérieux, France) and then subcul-

tured onto sheep blood agar plates. Additionally, solid selective differentiation Granada medium (bioMérieux, France) was used, onto which specimens collected from vaginal vestibule and rectum were inoculated directly, as previously described (Romanik *et al.* 2011).

Isolated strains were identified as GBS based on the following criteria: their morphological features (colony characteristics including a narrow zone of β -hemolysis and Gram positive staining), serological (Slidex Strepto-Kit bioMérieux, France) and biochemical (rapid ID 32 Strep bioMérieux, France) tests, and the growth and orange pigment formation on Granada agar (bioMérieux, France).

Serotyping of the isolates was performed using the Essum GBS Serotyping Kit (Umea, Sweden) according to manufacturer's instruction. The kit recognizes the nine polysaccharide antigens: Ia, Ib, II, III, IV, V, VI, VII, VIII.

Statistical analysis was performed with the Statistica 8,0 computer software (StatSoft, Krakow, Poland). Fisher's exact test was used for qualitative variables and Kruscal-Wallis test – for quantitative variables. The p -level ≤ 0.05 were accepted as statistically significant.

RESULTS

In the investigated group of 80 pregnant women GBS colonization rate was 28.7%. Four GBS serotypes, specified by the polysaccharide capsule, were observed (Ia, V, III and II). Serotype Ia was the most predominant – 43.47%, followed by V (7 from 23) and III (5 from 23). We compared the occurrence of GBS serotypes isolation from anovaginal region with the selected socio-demographic characteristics and medical history (Table 1). Serotype II was not included in the analysis as only one gravida was colonized with GBS serotype II. For GBS Ia, V and III serotypes, no significant difference in the prevalence of diabetes mellitus and neonatal outcomes was observed. Only in one case early-onset sepsis was diagnosed in the neonate and serotype Ia was determined.

DISCUSSION

The data on the rate of GBS colonization in Polish gravidas vary from 3.3% up to 30% (Romanik *et al.* 2011; Brzychczy-Włoch *et al.* 2010; Brzychczy-Włoch *et al.* 2011). Additionally, the presence of GBS in the vagina is transient and was seen mainly in young women and did not correlate with vaginal symptoms (vaginal erythema, vaginal desquamation, itching) (Romanik *et al.* 2007; Romanik *et al.* 2011; Wiechuła *et al.* 2007; Friedek *et al.* 2005; Kiely *et al.* 2011). In the authors' previous research, 27.8% of gravidas were GBS colonized with the mean age of 28 years old; our results are in accordance with other European studies (Brzychczy-Włoch *et al.* 2010; Brzychczy-Włoch *et al.* 2011; Daniels *et al.* 2011; Kunze *et al.* 2011).

In our study in the 80 pregnant women before delivery the most frequent GBS serotypes was serotype Ia and V (74%), whereas serotypes Ib, IV, and VI–VIII were not identified. Sadowy *et al.* and Brzychczy – Włoch showed in their studies that among seven observed serotypes Ia and V were identified in 37.4% and 37% GBS strains, respectively (Brzychczy-Włoch *et al.* 2010; Brzychczy-Włoch *et al.* 2011; Sadowy *et al.* 2010).

We determined EOD in one case – the infection was found to be associated with isolates of GBS serotype Ia as seen by others (Sadowy *et al.* 2010; Martins *et al.* 2007). Interestingly, serotype II in our study was observed only in one pregnant women, whereas in other researches this serotypes showed a tendency to be associated with non-invasive infections (Sadowy *et al.* 2010; Bisharat *et al.* 2005).

The differences between GBS serotypes distribution varies with geographic area, ethnic origin, virulence and antibiotics resistance of clinical isolates (Kiely *et al.* 2011; Dadvand *et al.* 2011; Ippolito *et al.* 2010; Lachenauer *et al.* 1999; Corvec *et al.* 2011). A number of studies have reported that serotype III was the most prevalent in LOD cases (Sadowy *et al.* 2010; Martins *et al.* 2007; Martins *et al.* 2011). This serotypes is also associated with meningitis; increased invasiveness of this serotype has also been suggested (Grey *et al.* 2007; Berg *et al.* 2000).

Data from Polish and German studies showed that 35% and 28% of pregnant women, respectively, were colonized with GBS serotypes III. This serotype is also very often isolated from neonates and adults in Sweden (from neonates – 62% and from adults – 29%) (Brzychczy-Włoch *et al.* 2010; Brzychczy-Włoch *et al.* 2011; Kunze *et al.* 2011; Sadowy *et al.* 2010; Berg *et al.* 2000). Additionally, serotypes III and V predominated in erythromycin – resistant GBS isolates in Southern Ireland (Kiely *et al.* 2010). Domelier *et al.* showed correlation between erythromycin resistance (*erm*) and GBS serotypes III, IV, V; *erm* serotype III strains were significantly more frequent for those isolated from vaginal carriage (30/136, 22%) and colonized neonates (3/17, 18%) than for those from EOD (0/25) (Domelier *et al.* 2008). However, in Japan, serotypes VI and VIII were the most common serotypes isolated from healthy pregnant women; these serotypes seem to be absent in central Europe (Lachenauer *et al.* 1999). Sadowy *et al.* among the seven serotypes observed in one hundred and fourteen GBS isolates from 43 centers from 28 towns during the period 1996–2005, identified serotype VI as the less prevalent (Sadowy *et al.* 2010). Serotyping shifts have been reported by Kiely *et al.* between two samplings period: 2004 and 2006. The authors observed increased in prevalence of serotype Ia (from 18.6% to 28.5%) and IV (from 7.6% to 15.2%), while serotype V decreased from 20.9% to 11.9% (Kiely *et al.*

Tab.1. Association between GBS serotypes and socioepidemiological data and neonatal outcomes.

Variable	Serological type of GBS			p-value*
	Ia (n=10)	III (n=5)	V (n=7)	
Age (years) (mean±SD, range)	27.3±4.81 (18–33)	33.2±6.50 (26–41)	27.0±5.03 (18–32)	0.41
Parity (median, range)	3 (1–2)	3 (1–3)	1 (1–5)	0.25
Miscarriage (median, range)	0 (0–1)	0 (0–1)	0 (0–2)	0.93
Residency (% ,n)				
Rural	40 (4)	20 (1)	14.3 (1)	0.46
Small city	10 (1)	40 (2)	14.3 (1)	
Large city	50 (5)	40 (20)	71.4 (5)	
GDM (n, %)	10 (1)	20 (1)	0 (0)	0.55
Diabetes mellitus (% ,n)	0 (0)	20 (1)	0 (0)	0.36
Hypothyreosis (% ,n)	0 (0)	20 (1)	0 (0)	0.66
Fetal weight (g) (mean±SD, range)	3377.0±405.49 (2700–3820)	3392.0±373.33 (2940–3890)	3458.57±504.56 (3030–4460)	0.51
Fetal hypotrophy (% ,n)	10 (1)	20 (1)	0 (0)	0.36
Fetal makrosomia (% ,n)	0 (0)	0 (0)	14.3 (1)	0.48
Apgar scores (median, range)	10 (6–10)	10 (7–10)	10 (10–10)	0.62
Blood loss during labor (ml) (mean±SD, range)	375.0±116.07 (200–500)	410.0±102.47 (250–500)	268.57±172.9 (150–600)	0.15

* Fisher's exact test for qualitative variables; Kruskal-Wallis test for quantitative variables; GBS – *Streptococcus* group

2011) in contrast, according to Ippolito et al., the seroprevalence of GBS have remained relatively stable in the United States (Martins *et al.* 2011).

Currently, monovalent and polyvalent vaccines containing capsular polysaccharides of GBS conjugated with tetanus toxoid have been tested. Those vaccines are well tolerated and might be administered in both non-pregnant women in the reproductive age as well as in gravidas in the III trimester of pregnancy. After the vaccination, initially, postvaccinal immune response with IgM is induced. High level of IgM is transient. However, the serum level of IgG increases after approximately 4–8 weeks after the initial immunization and last over 26 weeks. In case of both mono- and polyvalent vaccines the 4-fold or higher rise in IgG titer was observed in 80–90% of cases. The presence of specific IgG in the mother's blood leads to the acquisition of the passive response in neonates (Kasper *et al.* 1996; Backer *et al.* 2003; Backer *et al.* 2004).

Active immunization aimed for preventing GBS colonization in mothers including not only serotypes V, II and III but also Ia GBS serotype may be an effective and safe way to prevent life threatening neonatal infections in the future, especially that the presence of specific IgG antigens in the mothers' blood creates an opportunity for acquisition of passive immunity by the neonate. Studies show that vaccinating pregnant and non-pregnant women at reproductive age is an effective way of preventing GBS infections in neonates (Melin *et al.* 2011).

CONCLUSIONS

From among four identified GBS serotypes in the population of Polish pregnant women, serotype Ia was the most dominant.

For GBS serotypes, no significant difference in the prevalence of diabetes mellitus and neonatal outcomes was observed.

Active immunization aimed for preventing GBS colonization in mothers should include not only serotypes V, II and III but also Ia in order to be an effective and safe in preventing life threatening neonatal infections.

REFERENCES

- Backer CJ, Paoletti LC, Rench MA, Guttormsen HK, Edwards MS, Kasper DL 2004. Immune response of healthy women to 2 different group B streptococcal type V capsular polysaccharide – protein conjugate vaccines. *J Infect Dis.* 189: 1103–12.
- Backer CJ, Rench MA, Fernandez M, Paoletti LC, Kasper DL, Edwards MS 2003. Safety and immunogenicity of bivalent group B streptococcal conjugate vaccine for serotypes II and III. *J Infect Dis.* 188: 66–73.
- Berg S, Trollfors B, Lagergård T, Zackrisson G, Claesson BA 2000. Serotypes and clinical manifestations of group B streptococcal infections in western Sweden. *Clin Microbiol Infect.* 6: 9–13.
- Bisharat N, Jones N, Marchaim D, Block C, Harding RM, Yagupsky P, *et al.* 2005. Population structure of group B streptococcus from a low-incidence region for invasive neonatal disease. *Microbiology.* 151: 1875–81.
- Brzychczy-Włoch M, Gosiewski T, Bodaszewska-Lubas M, Adamski P, Heczko PB 2011. Molecular characterization of capsular polysaccharides and surface protein genes in relation to genetic similarity of group B streptococci isolated from Polish pregnant women. *Epidemiol Infect.* 14: 1–8.
- Brzychczy-Włoch M, Gosiewski T, Bodaszewska M, Pabian W, Bulanda M, Kochan P, *et al.* 2010. Genetic characterization and diversity of *Streptococcus agalactiae* isolates with macrolide resistance. *J Med Microbiol.* 59: 780–786.
- Centers for Disease Control and Prevention 2010. Prevention of perinatal group B streptococcal disease: Revised Guidelines from CDC. *MMWR.* 59: 1–39.
- Corvec S, Illiaquer M, Touchais S, Boutoille D, van der Mee-Marquet N, Quentin R, *et al.* 2011. Clinical features of group B *Streptococcus* prosthetic joint infections and molecular characterization of isolates. *J Clin Microbiol.* 49: 380–2.
- Dadvand P, Basagana X, Figueras F, Sunyer J, Nieuwenhuijsen MJ 2011. Climate and group B streptococci colonisation during pregnancy: present implications and future concerns. *BJOG.* 118: 1396–400.
- Daniels JP, Gray J, Pattison HM, Gray R, Hills RK, Khan KS GBS Collaborative Group 2011. Intrapartum tests for group B streptococcus: accuracy and acceptability of screening. *BJOG.* 118: 257–65.
- Domelier AS, van der Mee-Marquet N, Arnault L, Mereghetti L, Lanotte P, Rosenau A, *et al.* 2008. Molecular characterization of erythromycin-resistant *Streptococcus agalactiae* strains. *J Antimicrob Chemother.* 62: 1227–33.
- Friedek D, Ekiel A, Romanik M, Chelmicki Z, Wiechula B, Wilk I, *et al.* 2005. Co-occurrence of urogenital mycoplasmas and group B streptococci with chlamydial cervicitis. *Pol J Microbiol.* 54: 253–5.
- Gray KJ, Bennett SL, French N, Phiri AJ, Graham SM 2007. Invasive group B streptococcal infection in infants, Malawi. *Emerg Infect Dis.* 13: 223–9.
- Håkansson S, Källén K 2006. Impact and risk factors for early-onset group B streptococcal morbidity: analysis of a national, population-based cohort in Sweden 1997–2001. *BJOG.* 113: 1452–8.
- Ippolito DL, James WA, Tinnemore D, Huang RR, Dehart MJ, Williams J, *et al.* 2010. Group B streptococcus serotype prevalence in reproductive-age women at a tertiary care military medical center relative to global serotype distribution. *BMC Infect Dis.* 10: 336.
- Kaambwa B, Bryan S, Gray J, Milner P, Daniels J, Khan KS, *et al.* 2010. Cost-effectiveness of rapid tests and other existing strategies for screening and management of early-onset group B streptococcus during labour. *BJOG.* 117: 1616–27.
- Kasper DL, Paoletti LC, Wessels MR, Guttormsen HK, Carey VJ, Jennings HJ, *et al.* 1996. Immune response to type III group B Streptococcal polysaccharide – tetanus toxoid conjugate vaccine. *J Clin Invest.* 98: 2308–14.
- Kiely RA, Cotter L, Mollaghan AM, Cryan B, Coffey A, Lucey B 2011. Emergence of group B *Streptococcus* serotype IV in women of child-bearing age in Ireland. *Epidemiol Infect.* 139: 236–8.
- Kiely RA, Lucey B, Cotter L 2010. Analysis of phenotype, genotype and serotype distribution in erythromycin-resistant group B streptococci isolated from vaginal flora in Southern Ireland. *Epidemiol Infect.* 138: 286–91.
- Kunze M, Ziegler A, Fluegge K, Hentschel R, Proempeler H, Berner R 2011. Colonization, serotypes and transmission rates of group B streptococci in pregnant women and their infants born at a single University Center in Germany. *J Perinat Med.* 39: 417–22.
- Lachenaier CS, Kasper DL, Shimada J, Ichiman Y, Ohtsuka H, Kaku M, *et al.* 1999. Serotypes VI and VIII predominate among group B streptococci isolated from pregnant Japanese women. *J Infect Dis.* 179: 1030–3.
- Lin CY, Hsu CH, Huang FY, Chang JH, Hung HY, Kao HA, *et al.* 2011. The changing face of early-onset neonatal sepsis after the implementation of a maternal group B *Streptococcus* screening and intrapartum prophylaxis policy—a study in one medical center. *Pediatr Neonatol.* 52: 78–84.

- 23 Lin FY, Clemens JD, Azimi PH, Regan JA, Weisman LE, Philips JB 3rd, *et al.* 1998. Capsular polysaccharide types of group B streptococcal isolates from neonates with early-onset systemic infection. *J Infect Dis.* **177**: 790–2.
- 24 Martins ER, Andreu A, Correia P, Juncosa T, Bosch J, Ramirez M, *et al.* 2011. Group B streptococci causing neonatal infections in Barcelona are a stable clonal population: 18-year surveillance. *J Clin Microbiol.* **49**: 2911–8.
- 25 Martins ER, Pessanha MA, Ramirez M, Melo-Cristino J 2007. Analysis of group B streptococcal isolates from infants and pregnant women in Portugal revealing two lineages with enhanced invasiveness. *J Clin Microbiol.* **45**: 3224–9.
- 26 Melin P 2011. Neonatal group B streptococcal disease: from pathogenesis to preventive strategies. *Clin Microbiol Infect.* **17**: 1294–303.
- 27 Moore MR, Schrag S J, Schuchat A 2003. Effects of intrapartum antimicrobial prophylaxis for prevention of group B streptococcal disease on the incidence and ecology of early – onset natal sepsis. *Lancet Infect Dis.* **3**: 201 –213.
- 28 Romanik M, Kafel J, Lagergård T, Martirosian G 2007. Streptococcus group B--association with Aerobic vaginitis and ability to human cell lines activation. *Med Dosw Mikrobiol.* **59**: 85–91.
- 29 Romanik M, Nowosielski K, Martirosian G, Poręba R, Sioma-Markowska U 2011. Identification of pregnant women at risk of Streptococcus group B colonisation. *Neuro Endocrinol Lett.* **32**: 308–12.
- 30 Sadowy E, Matynia B, Hryniewicz W 2010. Population structure, virulence factors and resistance determinants of invasive, non-invasive and colonizing Streptococcus agalactiae in Poland. *J Antimicrob Chemother.* **65**: 1907–14.
- 31 Shen X, Lagergård T, Yang Y, Lindblad M, Fredriksson M, Holmgren J 2000. Systemic and mucosal immune responses in mice after mucosal immunization with group B streptococcus type III capsular polysaccharide-cholera toxin B subunit conjugate vaccine. *Infect Immun.* **68**: 5749–55.
- 32 Stoll BJ, Hansen NI, Sánchez PJ, Faix RG, Poindexter BB, Van Meurs KP, *et al.* 2011. Early onset neonatal sepsis: the burden of group B Streptococcal and E. coli disease continues. *Pediatrics.* **127**: 817–26.
- 33 Wiechula BE, Friedek DA, Ekiel AM, Romanik MK, Martirosian G 2007. Human neutrophil peptides in vaginitis/cervicitis of different etiology. *Pol J Microbiol.* **56**: 185–9.