H₁-antihistamines and oxidative burst of professional phagocytes

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OBJECTIVES: We analysed and compared the effect of five H₁-antihistamines on stimulated oxidative burst at extra- and intracellular level of isolated and stimulated human polymorphonuclear leukocytes.

DESIGN: Oxidative burst of isolated human neutrophils was studied by means of luminol and isoluminol enhanced chemiluminescence.

RESULTS: The following rank order of potency for H₁-antihistamines to decrease chemiluminescence was evaluated extracellularly: dithiaden> loratadine> chlo-rpheniramine> brompheniramine> pheniramine and at intracellular site: loratadine> dithiaden.

CONCLUSION: H_1 -antihistamines differ substantially according to their chemical structure in suppressing oxidative burst both at extra- and intracellular site of isolated stimulated human neutrophils.

INTRODUCTION

Abstract

Antihistamines are often viewed as selective antagonists of the histamine H_1 -receptor with relatively few other actions. Besides their antihistaminic activities, H_1 -receptor antagonists possess other pharmacological properties: anti-inflammatory action, inhibition of blood platelet function and antioxidative effects (Drabikova *et al.* 2002; Nosal & Jancinova, 2002, Nosal *et al.* 2002; Macickova *et al.* 2008). The mechanism(s) responsible for the nonspecific non-receptor operated activity of H_1 antihistamines is not fully understood and may operate both at receptor and non-receptor site of target cells (Church, 2001). The antioxidative effect in cell-free system differs substantially from antiplatelet and antiphagocyte activities, indicating that the H_1 - antihistamines studied act both at extracellular and intracellular level. In this study we compared the effect of five H_1 -antihistamines and histamine on stimulated chemiluminescence (CL) at extra- and intracellular site of human neutrophils. Moreover, we evaluated the EC₅₀ for histamine and the antihistamines studied at extraand intracellular site of stimulated neutrophils.

MATERIAL AND METHODS

Chemicals and drugs tested. Luminol, isoluminol, PMA (4-phorbol-12-myristate-13-acetate), superoxide dismutase and histamine dihydrochloride were obtained from Sigma Aldrich Chemie (Diesenhofer, Germany), horseradish peroxidase (HRP) and catalase from Merck (Darmstadt,Germany).

Abbreviations & Units:		
CL	chemiluminescence	
PA	pheniramine	
BPA	brompheniramine	
CPA	chlorpheniramine	
DIT	dithiaden	
HIST	histamine	
LOR	loratadine	
PMA	4-phorbol-12-myristate-13-acetate	
HRP	horse radish peroxidase	
EC ₅₀	concentration providing 50% effect	

All other chemicals of analytical grade were obtained from available commercial sources. Drugs tested: pheniramine maleate (PA; Hoechst Franfurt/M,Germany), chlorpheniramine maleate (CPA; Glaxo, Brentdorf, England), brompheniramine maleate (BPA; Wyeth Laboratories, Berks, England), dithiaden (DIT; Leciva Praha, Czech Republic), loratadine (LOR; Slovakofarma Hlohovec, Slovak Republic).

Isolation of polymorphonuclear leukocytes (PMNL). Human blood was collected in 2×9 ml citrate tubes, purified by 3% dextrane centrifugation and separated on Lymphoprep (Fresenius, Norway). The erythrocytes were removed with hypotonic, cold haemolysis. Cells were washed with phosphate-buffered saline before counting on Beckman Coulter. Concentration of cells in stock suspension was 10×10^6 cells /ml (for details see Drabikova *et al.* 2002)

Extra- and intracellular chemiluminescence. dCL was measured in a microtitre plate computer driven luminometer LM-01T (Immunotech, Czech Republic) at 37°C, using human isolated neutrophils. Measurement of extracellular CL was initiated by addition of PMA (50 μ l, final concentration 0.05 μ mol/L) to the reaction mixture, which consisted of isoluminol (5 µmol/L), neutrophils (5×10⁵), drug tested (0.01–100 μ mol/L) and horseradish peroxidase (HRP, 8 U/ml), in 50 µl aliquots. In experiments where the intracellular light emission was recorded, luminol (5 µmol/L) was used as luminophore, and superoxide dismutase (25 µl, 100 U/ ml) and catalase (25 µl, 2000 U/ml) were added instead of HRP. All concentrations are final. CL was recorded continuously for 30 min to obtain kinetic curves and evaluated on the basis of peak values (Drabikova et al. 2006; Jancinova et al. 2006a).

RESULTS

Figure 1 demonstrates the effect of H_1 -antihistamines and histamine on extracellular CL of human neutrophils stimulated with PMA. All drugs tested decreased dose-dependently CL of neutrophils with significant inhibition at 10 µmol/L concentration. Pheniramines and histamine decreased CL by 15%, loratadine by 39%. The most potent was dithiaden, decreasing CL by 58%. Increase of the concentration to 100 µmol/l resulted in further suppression of neutrophil CL. The effect of H_1 -antihistamines on intracellular CL of isolated human neutrophils is demonstrated in *Figure 2*. Pheniramines in concentrations of 0.1 and 10 µmol/L did not change intracellular CL, in 100 µmol/l concentration PA, CPE and BPE potentiated CL by 11, 23 and 13 %, respectively. Histamine did not change intracellular CL in any concentration used. DIT and LOR decreased significantly intracellular chemiluminescence in 10 µmol/L concentration by 42 and 37%, respectively. Increase of the concentration of DIT and LOR to 100 µmol/L resulted in the inhibition of intracellular CL by 95 and 96%, respectively.

Figure 3 shows EC_{50} values for pheniramines, HIST, DIT and LOR calculated for inhibition of extra- and intracellular CL. The respective rank order of potency for EC_{50} of BPH, CPH, DIT and LOR was 50.9, 98.4, 8.6 and 12.1 µmol/L. EC_{50} values for PA and histamine were over 100 µmol/L. Since BPH, CPH and PA potentiated intracellular CL, the EC_{50} values were irrelevant (negative). The same was true for HIST.

DISCUSSION

All H₁-antihistamines tested decreased dose-dependently the CL of whole human blood, both when stimulated by membrane-operating or by membranebypassing stimuli (Jancinova et al. 2006c; Kralova et al. 2008b; Nosal et al. 2002b). Inhibition of CL at extracellular site of neutrophils seems to be the result of scavenging free radicals due to molecules of H₁-antihistamines and HIST, as demonstrated for drugs studied in a cell free system (Kralova et al. 2008b). DIT and LOR significantly suppressed stimulated CL inside neutrophils, indicating an interaction with the regulatory pathway stimulated with PMA. This may suggest an interaction at protein kinase C, as recently demonstrated for diferuloylmethane (Jancinova et al. 2009). As expected, HIST operated extracellularly and was not effective inside neutrophils since it does not enter living cells. On the other hand, pheniramines potentiated stimulated CL of neutrophils at intracellular site in the highest concentration used (100 µmol/L). The wide variety in the suppressive effect of H₁-antihistamines on stimulated neutrophil CL is demonstrated by EC_{50} values of the drugs tested and HIST.

When examining the possible mechanisms by which antihistamines exert their anti-inflammatory effects, both receptor-independent and receptor-dependent mechanisms need to be taken into consideration. Although the number of recently published results (Jancinova *et al.* 2006c, Kralova *et al.* 2006, Nosal *et al.* 2002b, 2005, 2006) demonstrated a very important role of DIT and other H₁-antihistamines in the regulation of reactive oxygen species production by professional phagocytes and in the regulation of myeloperoxidase activity (Kralova *et al.* 2008a,b), further information is needed to provide insight into the effect of H₁-antihis-





Figure 2: Dose-dependent effect of pheniramine (PA), chlorpheniramine (CPA), brompheniramine (BPA), dithiaden (DIT), loratadine (LOR) and histamine (HIST) on intracellular human neutrophil isoluminolenhanced chemiluminescence after stimulation with PMA. n=6; x±SEM, **p<0.01

Figure 3: EC₅₀ values calculated for extracellular (extracell) and intracellular (intracell) chemiluminescence of human neutrophis exposed to pheniramine (PA), chlorpheniramine (CPA), brompheniramine (BPA), dithiaden (DIT), loratadine (LOR) and histamine (HIST) and stimulated with PMA.

tamines on the mechanism of reactive oxygen species generation.

The effect of the pheniramines tested on CL of oxidative burst of whole human blood was structure-dependent and related to lipophilicity, partition coefficient and other physico-chemical parameters (Jancinova *et al.* 2006b). BPA and CPA differ from PA by halogenisation of the molecule with Br and Cl. Moreover, the affinity and binding kinetics of H_1 -AH were found to be differently affected by an acidic environment, which can be encountered during inflammation (Gillard & Cahdelain, 2006).

Thus it appears likely that antihistamines exert antiinflammatory effects by both receptor-dependent and receptor-independent mechanisms. The receptor-independent mechanisms, which require higher drug concentrations, appear to include the release of preformed mediators from inflammatory cells, such as HIST and eosinophil proteins, eicosanoid generation, and particularly oxygen free radical production (Church, 2001). The mechanism of CL inhibition of professional phagocytes due to H_1 -antihistamines differs according to their chemical structure. Suppression of intracellular CL suggests interaction of H_1 -antihistamines with regulatory pathways responsible for oxidative burst of neutrophils, like proteinkinase C (Jancinova *et al.* 2009), NADPH-oxidase and others.

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REFERENCES

- Church MK (2001). H₁-Antihistamines and Inflammation. Clin Exper Allergy. **31**: 1341–1343.
- 2 Drabikova K, Nosal R, Jancinova V, Ciz M, Lojek A (2002). Reactive oxygen metabolite production is inhibited by histamine and H₁-antagonist dithiaden in human PMN leukocytes. Free Rad Res **36**: 975–980.
- 3 Drabikova K, Jancinova V, Nosal R, Pecivova J, Macickova T. (2006). Extra- and intracellular oxidant production in phorbol myristate acetate stimulated human polymorphonuclear leukocytes: modulation by histamine and H₁-antagonist loratadine. Inflamm Res. **55**: S19–20.
- 4 Gillard M, Cahdelain P. (2006). Changes in pH differently affect the binding of histamine H₁- receptor antagonists. Eur J Pharma-col. **530:** 205–214.
- 5 Jancinova V, Drabikova K, Nosal R, Rackova L, Majekova M, Holomanova D (2006a). The combined luminol/isoluminol chemiluminescence method for differentiating between extracellular and intracellular oxidant production by neutrophils. Redox Report. 11: 110–116.

- 6 Jancinova V, Drabikova K, Nosal R, Majekova M, Rackova L, Holomanova D (2006b). Antiradical effects of antihistamines in human blood. Structure-activity relationship. Inflamm Res. 55: S85–86.
- 7 Jancinova V, Drabikova K, Nosal R, Holomanova D (2006c). Extraand intracellular formation of reactive oxygen species by human neutrophils in the presence of pheniramine, chlorpheniramine and brompheniramine. Neuroendocrinol Lett. **27:** 141–143.
- 8 Jancinova V, Perecko T, Nosal R, Kostalova D, Bauerova K, Drabikova K (2009). Decreased activity of neutrophils in the presence of diferululoylmethane (curcumin) involve protein kinase C inhibition. Eur J Pharmacol 612: 161–166.
- 9 Kralova J, Ciz M, Nosal R, Lojek A (2006). Effect of H₁-antihistamines on the oxidative burst of rat phagocytes. Inflamm Res. 55: S15–16.
- 10 Kralová J, Pekarova M, Drabikova K, Jancinova V, Nosal R, Ciz M, Lojek A (2008a). The effect of dithiaden on nitric oxide production by RAW 264.7 cells. Interdisc Toxicol. 1: 214–217.
- 11 Kralova J, Nosal R, Drabikova K, Jancinova V, Denev P, Moravcova A, et al. (2008b). Comparative investigations of the influence of H₁-antihistamines on the generation of reactive oxygen species by phagocytes. Inflamm Res 57: S49–50.
- 12 Macickova T, Pecivova J, Nosal R, Lojek A, Pekarova M, Cupanikova D (2008). Inhibition of superoxide generation and myeloperoxidase release by carvedilol after receptor and nonreceptor stimulation of human neutrophils. Neuroendocrinol Lett 29: 790–793.
- 13 Nosal R, Jancinova V (2002a). Cationic amphiphilic drugs and platelet phospholipase A₂ (cPLA₂). Thromb Res. **105**: 339–345.
- 14 Nosal R, Drabikova K, Ciz M, Lojek A, Danihelova E (2002b). Effect of H₁-antagonist Dithiaden on human PMN-leukocyte aggregation and chemiluminescence is stimulus dependent. Inflamm Res **51:** 557–562.
- 15 Nosal R, Drabikova K, Jancinova V, Petrikova M, Fabryova V (2005). Antiplatelet and antiphagocyte activity of H₁-antihistamines. Inflamm Res. 54: S19–20.
- 16 Nosal R, Drabikova K, Jancinova V, Macickova T, Pecivova J, Holomanova D (2006). On the pharmacology and toxicology of neutrophils. Neuroendocrinol Lett. 27: 148–151.